VOL. XIX APRIL, 1935 No. 2.

Entered as second-class matter September 14, 1934, at the post office at Río Piedras, Puerto Rico, under the Act of August 24, 1912.

THE JOURNAL OF AGRICULTURE

of the

UNIVERSITY OF PUERTO RICO

In continuation of The Journal of the Department of Agriculture of Puerto Rico

MELVILLE T. COOK, Editor

New Cerambycid Beetles from Puerto Rico
By *W. S. Fisher*..... 51

New Eucnemid Beetles from Puerto Rico
By *W. S. Fisher*..... 65

New Species of Scarabaeidae (Coleoptera) from Puerto Rico and the Virgin Islands
By *Edward A. Chapin*..... 67

Nitification Studies with Soil Types of Northern Puerto Rico
By *Juan A. Bonnet*..... 73

Heat sterilization of Mangoes and Guavas for Fruit Flies
By *F. Sein Jr*..... 105

The First record of the Mosaic Disease of Sugar-Cane in Puerto Rico
By *G. N. Wolcott*..... 117

Diseases of Sugar-Cane in Puerto Rico
Part I. Normal Structure of Roots
By *Melville T. Cook*..... 121
Part II. A New Parasitic Fungus in the Root of Sugar Cane
By *Melville T. Cook*..... 124

First Supplement to Partial Bibliography of Virus Diseases of Plants
By *José I. Otero* and *Melville T. Cook*..... 129

PUBLISHED BY
THE AGRICULTURAL EXPERIMENT STATION
RÍO PIEDRAS, P. R.

ISSUED AUGUST 1935

UNIVERSITY OF PUERTO RICO

CARLOS E. CHARDON, M. S., D. S.—Chancellor
H. G. PARKINSON, M. S.—Dean, College of Agriculture and Mechanic Arts

AGRICULTURAL EXPERIMENT STATION STAFF

F. A. LÓPEZ DOMÍNGUEZ, B. S.—Director

DIVISION OF SOILS

JUAN AMEDÉE BONNET, M. S.—Soil Expert
FERNANDO A. VILLAMIL, B. S.—Soil Chemist
LEONARDO IGARAVÍDEZ, B. S.—Assistant Chemist

DIVISION OF AGRICULTURAL ENGINEERING

MANUEL L. VICENTE, C. E., M. E.—Engineer
JOSÉ ADSUAR, B. S.—Assistant Engineer

DIVISION OF CHEMISTRY

RAFAEL ARROYO, B. S., S. E.—Chemist
JOSÉ H. RAMÍREZ, B. S.—First Analytical Chemist
HÉCTOR CRUZ MONCLOVA, B. S.—Second Analytical Chemist

DIVISION OF ENTOMOLOGY

GEORGE N. WOLCOTT, Ph. D.—Entomologist
FRANCISCO SEÍN, JR., B. S.—Assistant Entomologist

DIVISION OF PHYTOPHATOLOGY

MELVILLE T. COOK, Ph. D.—Phytopathologist
ARTURO ROQUE, M. S.—Assistant Phytopathologist

DIVISION OF ZOOTECHNICS

FRANK PICÓ, B. S.—Zootechnician

DIVISION OF PHYTOTECHNICS

PEDRO RICHARDSON KUNTZ, B. S.—Sugar Cane Specialist
FERNANDO CHARDON, B. S.—Tobacco Specialist
FERDINAND MÉNDEZ, B. S.—Cocoanut Specialist
E. MOLINARY SALÉS, B. S.—Horticulturist
J. PASTOR RODRÍGUEZ, B. S.—Cotton Specialist
A. RIOLLANO, B. S.(1)—Specialist in Minor Crops

DIVISION OF AGRICULTURAL ECONOMICS

J. E. MCCORD, M. S.—Economist, Chief of the Division
S. L. DESCARTES, M. S.—Assistant Economist
JORGE SERRALLÉS, JR., B. S.—Assistant Economist

LIBRARY AND PUBLICATIONS

JOSÉ I. OTERO—Librarian

OFFICE

ARTURO RIVERA—Chief Clerk and Accountant

ISABELA SUB-STATION

LUIS A. SERRANO, B. S.—Agronomist in Charge
CARLOS J. CLAVELL, B. S.—Assistant Agronomist

(1) At Isabela.

The Journal of Agriculture of the University of Puerto Rico

In continuation of The Journal of the Department of Agriculture of Puerto Rico

Published Quarterly: January, April, July and October of each year

MELVILLE T. COOK, EDITOR

VOL. XIX. APRIL 1935 No. 2.

NEW CERAMBYCID BEETLES FROM PUERTO RICO

By W. S. FISHER

Associate Entomologist, Bureau of Entomology and Plant Quarantine, United States Department of Agriculture

Among the material received during the past year from Puerto Rico for identification, the following twelve new species of Cerambycidae were found.

Tilloclytus puertoricensis, new species

Small, elongate, slightly depressed above, nearly glabrous, subopaque; head black; pronotum black anteriorly, reddish at base; elytra reddish, with a broad, transverse, white fascia at middle, the fascia arcuately emarginate in front at sutural margins and bordered anteriorly by a narrow, black fascia, each elytron with an elongate, black spot along lateral margin extending from the white fascia backward to near apex and inward to middle of elytron; antenna and under side of body black or reddish brown, with bases of tibiae and femora, posterior part of prosternum, anterior part of mesosternum, and basal joints of antennae yellowish or reddish.

Head with the front wider than long, flat between the antennal tubercles, which are widely separated, but not elevated; surface densely, finely granulose, coarsely, confluently punctate, sparsely clothed with short, recumbent, inconspicuous, white hairs, with a few long, erect, white hairs around the eyes; eyes small, oblong, strongly convex, but not emarginate. Antenna extending to basal third of elytra, the outer joints slightly enlarged; first joint long, cylindrical, slightly arcuate, slightly expanded toward apex, subequal in length to the third and fourth joints united; third joint twice as long as the fourth, which is distinctly shorter than the fifth.

Pronotum one-half longer than wide, distinctly narrower at base than at apex, widest along middle; sides nearly parallel at middle, slightly rounded anteriorly, strongly constricted at base; disk strongly convex anteriorly, transversely depressed on basal third; surface finely, densely granulose, coarsely, deeply alveolate-punctate, clothed with a few long, erect, stiff, whitish hairs. Scutellum small, elongate, rounded at apex, slightly pubescent.

Elytra nearly one and three-fourths times as long as pronotum, at base slightly wider than pronotum at middle; sides nearly parallel from base to behind middle, where they are feebly, arcuately expanded, then arcuately narrowed to the tips, which are conjointly broadly rounded; disk strongly flattened on basal halves, strongly convex on apical halves; surface coarsely, deeply alveolate-punctate, with a few long, erect, stiff, white hairs posteriorly, each elytron with

a narrow vitta of short, recumbent, yellowish white hairs along sutural margin at apex.

Abdomen beneath smooth, shining, nearly glabrous; last segment broadly rounded at apex. Prosternum very sparsely, feebly punctate, feebly, transversely rugose, nearly glabrous; prosternal process very narrow between the coxal cavities, strongly expended posteriorly, truncate at apex. Metasternum and mesosternum with a small spot of dense, recumbent, whitish pubescence on each side. Legs rather long, smooth, clothed with a few scattered, long, erect, white hairs; femora strongly, abruptly, clavate toward apices, petiolate at bases; tibiae long, subcylindrical.

Length, 3 — 4.25 mm; width, 0.88 — 1.3 mm.

Type locality—Guánica, Puerto Rico.

Type and paratypes.—U. S. National Museum, No. 51019.

Remarks.—Described from three specimens (one type) collected in decaying wood on the Borinquen Forest Reservation, October 3, 1934, by R. G. Oakley (San Juan No. 5854).

This species is closely allied to *minutus* Fisher, but it differs from that species in having the basal part of the pronotum and basal halves of elytra reddish.

Lamproclytus oakleyi, new species

Elongate, nearly parallel, moderately convex above, strongly shining, above and beneath uniformly dark reddish brown, the pronotum slightly darker, each elytron ornamented with a transverse, eburneous fascia.

Head with the front strongly transverse, flat between the antennal tubercles, which are widely separated and slightly elevated; surface coarsely alveolate-punctate, sparsely clothed with long, semierect, white hairs. Antenna extending just beyond base of elytron, basal joints cylindrical, slightly expanded toward apices, clothed with a few long, erect, white hairs, apical joints broader, slightly flattened, rather densely clothed with short, recumbent pubescence, with a few long, erect intermixed; first joint robust, cylindrical, about twice as long as the second joint, which is two-thirds as long as the third; tenth joint oval, acutely rounded at apex.

Pronotum distinctly longer than wide, slightly wider at apex than at base, widest at middle; sides strongly, arcuately rounded from apex to basal sixth, where they are strongly constricted and parallel; disk strongly convex, feebly, transversely flattened along anterior margin, strongly, transversely constricted at base; surface coarsely alveolate-punctate, sparsely clothed with short, inconspicuous pubescence, and numerous long, erect, stiff, white hairs. Scutellum elongate-triangular, rather densely clothed with short, semierect pubescence.

Elytra twice as long as pronotum, at base subequal in width to pronotum at middle; sides nearly parallel, feebly, very broadly, arcuately constricted at middle, the tips separately broadly rounded or subtruncate; disk feebly, broadly, transversely depressed at middle, vaguely, broadly elevated along sutural margins behind scutellum; surface very coarsely, deeply, irregularly punctate, clothed with a few long, erect, stiff, white hairs, each elytron ornamented behind the

middle with a rather broad, transverse, eburneous fascia, extending from lateral margin to near the sutural margin.

Abdomen beneath very sparsely, feebly punctate, clothed with a few long, erect, white hairs; last segment broadly rounded at apex. Prosternum sparsely, coarsely punctate, rather densely clothed with moderately long, recumbent, whitish pubescence; prosternal process narrow, strongly expanded behind the coxal cavities. Legs sparsely clothed with long, erect, stiff, white hairs; anterior pair slightly shorter than middle and posterior pairs; femora strongly, abruptly clavate toward the tips, petiolate at bases; tibiae straight and subcylindrical.

Length, 3.25 mm; width, 0.85 mm.

Type locality.—Ponce, Puerto Rico.

Type.—U. S. National Museum, No. 51020.

Remarks.—Described from a single specimen collected on *Tabebuia* sp. at the Guánica Central Finca, September 21, 1933, by R. G. Oakley (San Juan No. 4693).

This species is closely allied to *elegans* Fisher, but it differs from that species in being uniformly dark reddish brown.

Leptostylus albosignatus, new species

Strongly robust, moderately convex, slightly flattened above, reddish brown to brownish black, rather densely clothed with brownish white and dark brown pubescence, the elytra ornamented with a large, distinct, irregular, square, whitish pubescent spot at middle, common to both elytra.

Head with the front quadrate, slightly convex, feebly depressed between the antennal tubercles, which are strongly elevated and obliquely divergent, with a narrow, longitudinal groove extending from occiput to epistoma; surface indistinctly punctate, densely, irregularly clothed with long, recumbent, brownish white pubescence, nearly concealing the surface, causing a slightly variegated appearance; eyes separated from each other on the top by twice the width of the upper lobe. Antenna about one-third longer than the body, variegated with white and brownish pubescence, the outer joints annulated with brown pubescence.

Pronotum three-fourths wider than long, subequal in width at base and apex, widest at middle; sides sinuate anteriorly, parallel posteriorly, tumid on each side at middle; disk feebly, tranversely depressed along base and anterior margin, with seven obtusely rounded tubercles arranged in two transverse rows on disk, four in front and three behind, the three median tubercles more strongly elevated than the four exterior ones; surface coarsely, deeply, irregularly punctate between the tubercles, and with a row of coarser punctures in the basal and anterior depressions, densely clothed with long, recumbent, brownish white pubescence, and ornamented with six small, dark brown pubescent spots, two along anterior margin and four along base. Scutellum transverse, broadly rounded at apex, rather densely pubescent.

Elytra three-fourths longer than wide, slightly wider than pronotum at middle; humeri prominent, slightly elevated; sides nearly parallel from base to apical fourth, then arcuately narrowed to the tips, which are separately narrowly, obliquely truncate internally, the exterior angle obtuse and feebly produced;

disk uneven, with numerous irregularly distributed, rather acute tubercles, those in the basal region more strongly developed; surface coarsely, deeply, irregularly punctate, the punctures becoming finer and sparser toward apices, densely clothed with long, recumbent, brownish white and dark brown pubescence, ornamented with an irregular, square, white pubescent spot at middle, common to both elytra, and with a few small, dark brown pubescent spots posteriorly.

Beneath nearly smooth, rather densely clothed with long, recumbent, brownish white pubescence, the legs variegated with whitish and brownish pubescence; tibiae feebly annulated with brown pubescence; prosternal process as wide as the coxal cavity.

Length, 9.5 — 12 mm; width, 4 — 4.75 mm.

Type locality.—Ponce, Puerto Rico.

Type and paratype.—U. S. National Museum, No. 51021.

Remarks.—Described from two specimens (one type). The type was collected at the type locality during December 1933 by C. Rinaldi, and the paratype was collected at light at the Lesesne Finca, Bayamón, Puerto Rico, November 12, 1933, by Rae Lesesne and C. G. Anderson (San Juan No. 4855).

This species is allied to *antillarum* Fisher, but it differs from that species in having a distinct, large, white, pubescent spot on the elytra.

Leptostylus oakleyi, new species

Elongate, slightly flattened above, reddish brown to brownish yellow, densely clothed with brownish yellow pubescence, and ornamented with dark brown or black pubescence.

Head with the front quadrate, slightly convex, feebly depressed between the antennal tubercles, which are strongly elevated and obliquely divergent, with a narrow, longitudinal groove extending from occiput to epistoma; surface indistinctly punctate, rather densely, irregularly clothed with long, recumbent, brownish white pubescence, nearly concealing the surface, causing a variegated appearance; eyes separated from each other on the top by the width of the upper lobe. Antenna about one-fourth longer than the body, variegated with brown and white pubescence, the joints annulated with dark brown pubescence.

Pronotum nearly twice as wide as long, subequal in width at base and apex, widest at middle; sides obliquely expanded from base to near middle, arcuately expanded at middle, parallel posteriorly; disk feebly, transversely flattened along base and anterior margin, with three vague, round protuberances, two arranged transversely in front and one behind; surface coarsely, deeply, irregularly punctate, with a row of similar punctures along base and anterior margin, densely clothed with long, recumbent, brownish white pubescence, irregularly ornamented around the dorsal protuberances with brownish pubescence, and with a distinct brown pubescent vitta on each side below the lateral protuberance. Scutellum broadly triangular, broadly rounded at apex, sparsely pubescent.

Elytra twice as long as wide, distinctly wider than pronotum at middle; humeri not prominent; sides parallel to behind middle, then arcuately narrowed to the tips, which are separately obliquely truncate internally; disk uneven, with

a few inconspicuous tubercles clothed with blackish hairs near the base; surface densely clothed with long, recumbent, brownish white pubescense, and each elytron ornamented with black pubescence as follows: A large, elongate spot along lateral margin; a short, narrow, oblique fascia along sutural margin behind middle; a narrow, arcuate fascia at lateral margin at apical fifth; an irregular, inconspicuous spot near apex; and with a few small, irregularly distributed, round spots.

Beneath obsoletely granulose, rather densely clothed with long, recumbent, whitish pubescence, the sternum and legs variegated with white and brown pubescence; tibiae annulated with brown pubescence; femora strongly pedunculate; prosternal process one-half as wide as the coxal cavity.

Length, 4.5 mm; width, 1.8 mm.

Type locality.—Bayamón, Puerto Rico.

Type.—U. S. National Museum, No. 51022.

Remarks.—Described from a single specimen collected at light at the Lesesne Finca, March 10, 1934, by Rae Lesesne and C. G. Anderson (San Juan No. 5257).

This species is allied to *gundlachi* Fisher, but it differs from that species in the different arrangement of the brown pubescence on the elytra, and in having a broad, dark brown, pubescent vitta on each side of the pronotum.

Leptostylus nigricans, new species

Small, elongate, moderately flattened above, brownish yellow, with the basal halves of elytra, disk of pronotum, tips of antennal joints, tarsi, and parts of head, tibiae, and femora, black or dark brown and rather densely pubescent.

Head with the front quadrate, slightly convex, feebly depressed between the antennal tubercles, which are rather strongly elevated and obliquely divergent, with a narrow, longitudinal groove extending from occiput to epistoma; surface indistinctly punctate, rather sparsely, irregularly clothed with long, recumbent, brownish yellow pubescence, with a few white and dark brown hairs intermixed, the brown hairs more numerous on the occiput; eyes separated from each other on the top by twice the width of the upper lobe. Antenna about one-fourth longer than body, variegated with white and brown pubescence, the joints annulated with dark brown pubescence.

Pronotum nearly twice as wide as long, subequal in width at base and apex, widest at basal third; sides feebly, obliquely expanded to basal third, then strongly, arcuately narrowed to near the base, parallel and strongly constricted at base; disk even, narrowly, tranversely grooved along base and anterior margin, the margins elevated; surface indistinctly punctate, rather densely, irregularly clothed with long, recumbent, black and reddish brown pubescence. Scutellum triangular, narrowly rounded at apex, densely pubescent.

Elytra nearly twice as long as wide, slightly wider than pronotum at basal third; humeri not prominent; sides parallel to behind middle, then arcuately narrowed to the tips, which are separately narrowly rounded; disk even, with a distinct tubercle clothed with long, black hairs near base of each elytron; surface

coarsely, densely, deeply punctate from bases to apices, the basal black area with the posterior margin extending obliquely backward from the sutural margin at middle to lateral margin at apical third, sparsely clothed with short, black hairs, with an irregular spot beneath the humerus and a few small spots at middle clothed with brownish yellow pubescence, the apical pale area densely clothed with long, recumbent, brownish yellow pubescence, and narrowly margined anteriorly with white pubescence.

Beneath feebly granulose, rather densely clothed with long, recumbent, whitish pubescence, the sternum and legs variegated with white and brown pubescence; tibiae annulated with brown pubescence; femora strongly peduneulate; prosternal process three-fourths as wide as coxal cavity.

Length, 4 mm; width, 1.75 mm.

Type locality.—Villalba, Puerto Rico.

Type.—U. S. National Museum, No. 51023.

Remarks.—Described from a single specimen collected in vegetative debris at the Insular Government Finca, June 18, 1934, by R. G. Oakley (San Juan No. 5666).

This species is allied to *dorsalis* Fisher, but it differs from that species in being shorter and more robust, and in having the pronotum broader, with the sides more broadly rounded at the basal third, and each elytron armed with a distinct basal tubercle and narrowly rounded at the apex.

Leptostylus puertoricensis, new species

Small, elongate, moderately flattened above, brownish black to brownish yellow, rather densely pubescent.

Head with the front slightly transverse, slightly convex, feebly depressed between the antennal tubercles, which are rather strongly elevated and obliquely divergent, with a narrow, longitudinal groove extending from occiput to epistoma; surface indistinctly punctate, sparsely, irregularly clothed with long, recumbent, brownish yellow pubescence, the pubescence dark brown on the occiput; eyes separated from each other on the top by twice the width of the upper lobe. Antennae about as long as the body, variegated with white and brown pubescence, the joints annulated with dark brown pubescence.

Pronotum twice as wide as long, subequal in width at base and apex, widest at basal third; sides obliquely expanded to basal third, then strongly, arcuately narrowed to near the base, parallel and strongly constricted at base; disk even, narrowly, transversely grooved along base and anterior margin; surface indistinctly punctate, densely clothed with long, recumbent, brownish yellow and brownish black pubescence. Scutellum triangular, narrowly rounded at apex, densely pubescent.

Elytra two-thirds longer than wide, and slightly wider than pronotum at basal third; humeri not prominent; sides parallel to behind middle, then arcuately narrowed to the tips, which are separately obliquely subtruncate internally; disk even, with a small tubercle clothed with black hairs near base of each elytron; surface rather densely, finely punctate from bases to apices, rather densely clothed

with long, recumbent, brownish yellow pubescence, each elytron with a more or less distinct, broad, whitish pubescent fascia, extending obliquely backward from the sutural margin at middle to the lateral margin at apical third, and ornamented with brownish black pubescence as follows: A large, semioval spot along lateral margin in front of midddle; an irregular area around basal tubercle; a narrow, irregular, interrupted, oblique fascia behind the whitish pubescent fascia; and a few small, irregularly distributed, round spots.

Beneath feebly granulose, rather densely clothed with long, recumbent, whitish pubescence, the sternum and legs variegated with white and brown pubescence; tibiae annulated with brown pubescence; femora strongly pedunculate; prosternal process one-half as wide as coxal cavity.

Length, 4 mm; width, 1.75 mm.

Type locality.—Adjuntas, Puerto Rico.

Type.—U. S. National Museum, No. 51024.

Remarks.—Described from a unique specimen collected on an unknown tree at the Pietri Finca, June 10, 1933, by R. G. Oakley (San Juan No. 4304).

This species is allied to *planicollis* Fisher, but it differs from that species in having the upper surface clothed with brownish yellow and brownish black pubescence, and each elytron ornamented near the middle with a broad, oblique, whitish pubescent fascia.

Eugamandus oakleyi, new species

Short, oblong, strongly convex, subopaque, uniformly reddish or yellowish brown, each elytron ornamented laterally with an irregular, black pubescent vitta.

Head with the front strongly transverse, nearly flat, flat between the antennal tubercles, which are widely separated and feebly elevated, with a vague, longitudinal groove extending from occiput to epistoma; surface sparsely, coarsely, uniformly punctate, rather densely clothed wtih short, scalelike, golden yellow and brownish hairs, with a few longer, semierect, whitish hairs intermixed; eyes small, narrow, rather deeply emarginate separated from each other on the top by nearly four times the width of the upper lobe. Antenna nearly as long as the body, annulated with short, recumbent, yellowish white and dark brown pubescence; first joint robust, subcylindrical, flattened beneath at base, subequal in length to the fourth joint, which is slightly shorter than the third, the following joints shorter and nearly equal in length.

Pronotum distinctly wider than long, subequal in width at base and apex, widest at middle; sides arcuately expanded at middle, parallel posteriorly; disk strongly convex, slightly uneven, more or less transversely gibbose at middle, broadly, transversely flattened posteriorly; surface coarsely, sparsely, deeply punctate, the punctures denser toward the sides, irregularly variegated with dense, short, scalelike, brownish yellow and dark brown hairs, with a few longer, semierect, whitish hairs intermixed. Scutellum strongly transverse, broadly rounded at apex, broadly concave, and clothed with scalelike hairs, which are dark brown at the middle and whitish on each side.

Elytra two and two-thirds times as long as pronotum, at base distinctly

wider than pronotum; sides nearly parallel from base to behind middle, then arcuately narrowed to the tips, which are seperately narrowly rounded; disk strongly convex, uneven, slightly flattened above, obliquely declivous behind middle, slightly elevated near apices; surface coarsely, sparsely, deeply punctate, variegated with dense, short, scalelike, brownish yellow and dark brown hairs, the hairs slightly paler toward apices, with a few long, erect, whitish hairs toward lateral margins, each elytron ornamented with a black pubescent vitta extending along lateral margin from humeral angle to basal third, then inward and backward to posterior declivity, and with numerous, strongly elevated, irregularly distributed tubercles, the largest located on middle of elytron at posterior declivity.

Abdomen beneath finely granulose, coarsely, densely, deeply punctate, rather densely clothed with short, scalelike, yellowish and whitish hairs, with a few longer, erect, whitish hairs intermixed; last segment broadly rounded at apex.

Length, 5.75 mm; width, 3 mm.

Type locality.—Matrullas Dam, near Orocovis, Puerto Rico

Type.—U. S. National Museum, No. 51025.

Remarks.—Described from a single specimen collected in decaying wood at the Insular Government Finca, October 8, 1934, by R. G. Oakley (San Juan No. 5861).

This species is allied to *schwarzi* Fisher, but it differs from that species in being more strongly convex and in having the elytra distinctly tuberculate.

Eugamandus brunneus, new species

Small, short, oblong, strongly convex, subopaque, uniformly dark reddish brown with the tarsi, antennae, and tibiae, in part, yellowish, and above irregularly variegated with black and brownish, scalelike hairs.

Head with the front strongly transverse, nearly flat, flat between the antennal tubercles, which are widely separated and feebly elevated, with a vague, narrow, longitudinal groove extending from occiput to epistoma; surface coarsely, sparsely, uniformly punctate, rather densely clothed with short, scalelike, golden yellow and brownish hairs, with a few longer, erect, whitish hairs intermixed; eyes small, narrow, deeply emarginate, separated from each other on the top by three times the width of the upper lobe. Antenna nearly as long as the body, rather densely clothed with short, recumbent pubescence; first joint robust, subcylindrical, flattened beneath at base, subequal in length to the fourth joint, which is three-fourths as long as the third, the following joints shorter and nearly equal in length.

Pronotum distinctly wider than long, subequal in width at base and apex, widest at middle; sides arcuately expanded at middle, parallel posteriorly; disk strongly convex, slightly uneven, more or less transversely gibbose at middle, broadly, transversely flattened posteriorly; surface coarsely, deeply, sparsely, irregularly punctate, the punctures denser toward the sides, irregularly variegated with dense, short, scalelike, brownish yellow and dark brown hairs. Scutellum similar to *oakleyi* Fisher.

Elytra nearly three times as long as pronotum, and at base distinctly wider than pronotum; sides nearly parallel from base to behind middle, then arcuately narrowed to the tips, which are separately narrowly rounded or subtruncate; disk strongly convex, uneven, slightly flattened above, obliquely declivous behind the middle; surface coarsely, sparsely, deeply punctate, variegated with dense, short, scalelike, brownish yellow and dark brown hairs, each elytron with an indistinct, irregular, blackish pubescent spot toward lateral margin, and ornamented with two longitudinal rows of tubercles, the inner row distinct and composed of three tubercles, the outer one less distinct.

Abdomen beneath finely granulose, feebly, coarsely, sparsely punctate, sparsely clothed with short, scalelike, whitish hairs, with a few longer, erect hairs of the same color intermixed; last segment broadly subtruncate at apex.

Length, 3.75 mm; width, 1.85 mm.

Type locality.—Yauco, Puerto Rico.

Type.—U. S. National Museum, No. 51026.

Remarks.—Described from a single specimen collected in vegetative debris at the Augustin Finca, in the mountains north of Yauco, June 15, 1934, by R. G. Oakley (San Juan No. 5654).

This species is very closely allied to *oakleyi* Fisher, but it differs from that species in being much smaller and more slender, and in having the tubercles on each elytron arranged in two longitudinal rows.

Eugamandus flavipes, new species

Small, short, oblong strongly convex, feebly shining, uniformly pale reddish brown, with the legs and the antennae yellowish, above more or less variegated with black and brownish, scalelike hairs.

Head with front strongly transverse, nearly flat, flat between the antennal tubercles, which are widely separated and feebly elevated, with a vague, narrow, longitudinal groove on occiput and vertex; surface coarsely, sparsely, uniformly punctate, densely clothed in front with moderately long, recumbent, yellow pubescence; eyes small, narrow, deeply emarginate, separated from each other on the top by four times the width of the upper lobe. Antenna nearly as long as the body, densely clothed with short, recumbent pubescence; first joint robust, subcylindrical, flattened beneath at base, subequal in length to the third and fourth joints, the following joints shorter and gradually diminishing in length.

Pronotum distinctly wider than long, narrower at apex than at base, widest at middle; sides arcuately expanded at middle, slightly parallel posteriorly; disk strongly convex, more or less gibbose at middle, rather broadly, transversely flattened along base and anterior margin; surface coarsely, deeply, sparsely punctate, more densely toward the side, rather densely clothed with short, scalelike, black hairs on median part, sparsely clothed with scalelike, whitish hairs at sides, and ornamented with a narrow, more or less interrupted, median vitta of similar white hairs. Scutellum similar to that of *oakleyi* Fisher.

Elytra two and one-half times as long as pronotum, at base distinctly wider than pronotum; sides nearly parallel from base to behind middle, then arcuately

narrowed to the tips, which are conjointly broadly rounded; disk strongly, uniformly convex, arcuately declivous posteriorly; surface coarsely, rather densely, deeply punctate, variegated with rather dense, short, scalelike, yellowish brown and dark brown hairs, each elytron ornamented with two irregular-shaped, black pubescent spots near middle, and two strongly elevated tubercles, one at basal fourth, the other just behind the middle.

Body beneath coarsely, rather densely punctate, sparsely clothed with short, recumbent, whitish hairs, with a few scalelike hairs intermixed; last segment broadly rounded or subtruncate as apex.

Length, 2.75 mm; width, 1.25 mm.

Type locality.—Villalba, Puerto Rico.

Type.—U. S. National Museum, No. 51027.

Remarks.—Described from a single specimen collected in vegetative debris at the Insular Govenment Finca, June 18, 1934, by R. G. Oakley (San Juan No. 5667).

This species is allied to *oakleyi* Fisher, but it differs from that species in having the elytra evenly convex, arcuately declivous posteriorly, and each elytron armed with only two distinct tubercles.

Cyrtinus eugeniae, new species

Very small, elongate, feebly shining; head, pronotum, and abdomen dark reddish brown; antennae, legs, and elytra slightly paler, the latter dark reddish brown along lateral margins.

Head with the front strongly transverse, feebly convex, flat between the antennal tubercles, which are widely separated and scarcely elevated, without a distinct, longitudinal groove; surface coarsely, deeply, confluently punctate, rather densely clothed with long, semierect, whitish hairs; eyes small, coarsely granulated, distinctly divided, separated from each other on the top by twice the width of the upper lobe, which is slightly smaller than the lower lobe. Antenna 11-jointed, about as long as the body, sparsely clothed with long and short hairs; first joint extending to apical third of pronotum, gradually expanded toward apex, one-half longer than third joint, which is subequal in length to the fourth, the following joints gradually diminishing in length.

Pronotum slightly longer than wide, distinctly narrower at base than at apex, widest along apical fourth; sides feebly expanded anteriorly, strongly constricted at basal fourth, then parallel to base; disk strongly convex anteriorly, narrowly, transversely flattened along base; surface coarsely, deeply, confluently punctate, sparsely, clothed with rather short, recumbent, whitish hairs.

Elytra twice as long as wide, slightly wider than pronotum near apex; sides nearly parallel, slightly, arcuately expanded behind middle, then arcuately narrowed to the tips, which are conjointly broadly rounded; disk transversely flattened on basal half, moderately convex posteriorly; surface coarsely, deeply, densely punctate basally, the punctures becoming obsolete near apices, sparsely, irregularly clothed with moderately long, recumbent, white hairs, with a few long, erect hairs intermixed.

Abdomen beneath vaguely punctate, sparsely clothed with short, inconspicuous hairs; last segment broadly rounded at apex. Legs sparsely clothed with short, recumbent, whitish hairs; femora strongly, abruptly clavate toward apices, the club scarcely flattened laterally.

Length, 2 mm; width, 0.63 mm.

Type locality.—Aibonito, Puerto Rico.

Type.—U. S. National Museum, No. 51028.

Remarks.—Described from a single specimen collected on *Eugenia* sp. at the Díaz Finca, October 6, 1933, by R. G. Oakley (San Juan No. 4768).

This species is allied to *hubbardi* Fisher, but it differs from that species in being much smaller, feebly shining, and coarsely, confluently punctured on the upper surface.

Cyrtinus subopacus, new species

Small, elongate, subopaque; head, pronotum, tarsi, and abdomen black or dark reddish brown; antennae, legs and elytra pale brownish yellow, the latter with a large black spot on each side toward lateral margin.

Head with the front flat, strongly transverse, flat between the antennal tubercles, which are widely separated and scarcely elevated, with a vague, narrow, longitudinal groove extending from epistoma to occiput; surface densely, finely granulose, coarsely, deeply, confluently punctate, sparsely clothed with long, recumbent, whitish hairs; eyes small, coarsely granulated, distinctly divided, separated from each other on the top by six times the width of the upper lobe, which is considerably smaller than the lower lobe. Antenna 11-jointed, about as long as the body, clothed with a few long and short hairs; first joint extending to apical third of pronotum, gradually expanded toward apex, twice as long as the third joint, which is subequal in length to the fourth; the following joints gradually decreasing in length.

Pronotum slightly longer than wide, distinctly narrower at base than at apex, widest near apex; sides feebly narrowed from apex to basal fourth, where they are strongly constricted, then parallel to base; disk strongly convex anteriorly, narrowly, transversely flattened along base, with a narrow, transverse groove at basal fourth; surface coarsely, deeply, confluently punctate, sparsely clothed with short, recumbent, whitish hairs.

Elytra twice as long as wide, slightly wider than pronotum near apex; sides parallel from base to apical fourth, then arcuately narrowed to the tips, which are conjointly broadly rounded; disk transversely flattened on basal half, moderately convex posteriorly; surface coarsely, deeply, confluently punctate, the punctures elongate, becoming obsolete near apices, sparsely, irregularly clothed with moderately long, recumbent, withish pubescence.

Abdomen beneath feebly, coarsely, sparsely punctate, sparsely clothed with fine, semierect hairs; last segment broadly rounded or subtruncate at apex. Legs sparsely clothed with short, recumbent, white hairs; femora strongly, abruptly clavate toward apices, the club slightly flattened laterally.

Length, 2.5 mm; width, 0.75 mm.

Type locality.—Adjuntas, Puerto Rico.

Type.—U. S. National Museum, No. 51029.

Remarks.—Described from a single specimen collected flying, at the Pietri Finca, April 13, 1933, by R. G. Oakley (San Juan No. 3984).

This species is allied to *eugeniae* Fisher, but it differs from that species in being larger and subopaque, and in having the punctures on the elytra elongate.

Cyrtinus oakleyi, new species

Very small, elongate, strongly shining; head and pronotum black or dark reddish brown, the latter with the base narrowly brownish yellow; elytra pale brownish yellow, each with a large, irregular black or reddish brown spot behind the middle, and a similar, but smaller, spot at apex; antenna pale brownish yellow with the joints darker at apices; beneath dark reddish brown, with the tibiae and tarsi pale brownish yellow.

Head with the front strongly transverse, feebly convex, flat between the antennal tubercles, which are widely separated and scarcely elevated, without a distinct longitudinal groove; surface vaguely granulose, coarsely, densely, rather deeply punctate, sparsely clothed with long, recumbent, whitish hairs; eyes small, coarsely granulated, distinctly divided, separated from each other on the top by about four times the width of the upper lobe, which is considerably smaller than the lower lobe. Antenna 11-jointed, about as long as the body, clothed with a few long and short hairs; first joint extending to middle of pronotum, gradually expanded toward apex, about twice as long as the third joint, which is slightly longer than the fourth, the following joints gradually decreasing in length.

Pronotum as wide as long, distinctly narrower at base than at apex, widest near apex; sides feebly narrowed from apex to basal fourth, where they are strongly constricted, then parallel to the base; disk strongly convex anteriorly, narrowly, transversely flattened along base, with a narrow, transverse groove at basal fourth; surface densely, coarsely, deeply punctate, and nearly glabrous.

Elytra twice as long as wide, slightly wider than pronotum near apex; sides parallel from base to apical third, then arcuately narrowed to the tips, which are conjointly broadly rounded; disk slightly flattened on basal half, moderately convex posteriorly; surface coarsely, rather densely punctate basally, the punctures becoming obsolete toward apices, sparsely, irregularly clothed with short, recumbent, white hairs, with a few long, erect hairs intermixed.

Abdomen beneath vaguely punctate, nearly glabrous; last segment subtruncate at apex. Legs sparsely clothed with short, recumbent, whitish hairs, more densely pubescent on outer margin of femora, which are strongly, abruptly clavate toward apices, the club slightly flattened laterally.

Length, 1.75 mm; width, 0.6 mm.

Type locality.—Yauco, Puerto Rico.

Type and paratype.—U. S. National Museum, No. 51030.

Remarks.—Described from two specimens (one type) collected in decaying plants at the Augustin Finca, in the mountains north of Yauco, June 2, 1934, by R. G. Oakley (San Juan No. 5625).

This species is allied to *eugeniae* Fisher, but it differs from that species in being strongly shining, and in having the pronotum as wide as long, and each elytron ornamented with two reddish brown spots.

NEW EUCNEMID BEETLES FROM PUERTO RICO

By W. S. FISHER

Associate Entomologist, Bureau of Entomology & Plant Quarantine,
United States Department of Agriculture

Among the material received during the past year from Puerto Rico for identification, the following four new species of Eucnemidae were found.

Dirhagus puertoricensis, new species

Body narrow, subcylindrical, attenuate posteriorly, reddish black, legs yellow, pubescence inconspicuous, ornamented with whitish pubescence as follows: Sides and base of pronotum, a broad, transverse fascia at base and a similar, more conspicuous fascia at apical third of elytra. Head convex in front, coarsely, confluently punctate. Antenna extending to middle of abdomen, joint 3 elongate, joint 4 triangular, joints 5 to 11 pectinate. Pronotum wider than long, parallel, depressed posteriorly, carinate in front of scutellum, densely ocellate-punctate; posterior angles carinate. Elytra attenuate posteriorly, depressed at base, densely punctate. Beneath coarsely, densely punctate. Length, 2.75 mm.

Type in United States National Museum, collected on coffee at the Paraiso Finca, Ponce, P. R. July 13, 1933, by R. G. Oakley (San Juan No. 4315).

Differs from *D. phyrrhopus* Chevrolat in being nearly black, and in having the antennae pectinate and the elytra ornamented with two transverse, whitish pubescent fasciae.

Dirhagus oakleyi, new species

Body narrow, subcylindrical, attenuate posteriorly, black, with posterior angles of pronotum, scutellum, basal two-thirds of sutural margins and basal halves of elytra, and sides of mesosternum and metasternum reddish brown, legs yellow, sparsely clothed on sides of pronotum with whitish pubescence, a vague, irregular, transverse fascia of whitish pubescence near middle of elytra, each elytron with a broad, transversely oblique fascia of golden yellow pubescence at apical third. Head convex in front, confluently ocellate-punctate. Antenna extending to middle of abdomen, joint 3 narrow, elongate, joint 4 triangular, joints 5 to 11 flabellate. Pronotum wider than long, parallel, carinate in front of scutellum, densely ocellate-punctate; posterior angles carinate. Elytra attenuate posteriorly, densely punctate. Beneath coarsely, densely punctate. Length, 2.6 mm.

Type in United States National Museum, collected on *Eugenia* sp. at the Rivera Finca, Aibonito, P. R., May 31, 1934, by R. G. Oakley (San Juan No. 5617).

Differs from *D. puertoricensis* Fisher in having the elytra bicolored, with a golden yellow pubescent fascia at apical third, and the antennae flabellate.

Nematodes puertoricensis, new species

Body very narrow, subcylindrical, dark reddish brown, legs and antennae paler, pubescence uniform, yellowish white. Head convex in front, uniformly, finely, densely punctate. Antenna with joints 2, 4 and 5 subequal, joints 6 to 10 slightly longer, broader, subequal, last joint longer, acute at apex. Pronotum parallel, slightly longer than wide, vaguely biimpressed on each side, finely, confluently punctate. Elytra strongly attenuate posteriorly, indistinctly striate, finely, densely punctate. Beneath finely, densely punctate, antennal grooves indistinct, posterior coxae strongly angulated. Length, 5 mm.

Type and paratype in United States National Museum, collected on weeds at the Insular Government Finca, Matrullas Dam, near Orocovis, P. R. October 8, 1934, by R. G. Oakley (San Juan No. 5859).

Differs from *N. simulans* Chevrolat in being much smaller, and in having the intermediate antennal joints as wide as long, and the elytra not distinctly striate.

Adelothyreus insularis, new species

Body narrow, subcylindrical, black, with an elongate, brownish yellow spot covering the exterior three-fourths of each elytron, the spot not extending to base or apex, tarsi and palpi yellow, pubescence sparse, uniform, whitish. Head convex in front, confluently ocellate-punctate. Antenna extending to posterior coxae, joint 3 slightly serrate, joints 5 to 11 strongly transverse. Pronotum quadrate, parallel, feebly flattened, depressed posteriorly, with 3 elevations at base, densely ocellate-punctate; posterior angles carinate. Elytra nearly parallel, depressed at bases, rugose basally, coarsely, densely punctate posteriorly. Beneath densely, coarsely punctate, more feebly on abdomen. Length, 3—3.25 mm.

Type and paratype in United States National Museum collected by R. G. Oakley. The type was collected on *Eugenia* sp. at the Rivera Finca, Aibonito, P. R., May 31, 1934 (San Juan No. 5617), and the paratype was collected at Castaner Finca, Adjuntas, P. R., February 5, 1934 (San Juan No. 5206).

Differs from *A. dufaui* Fleutiaux in having the antennal joints strongly transverse, the elytra more parallel, pubescence whitish, with a large brownish yellow spot covering the exterior three-fourths of each elytron.

NEW SPECIES OF SCARABAEIDAE (COLEOPTERA) FROM PUERTO RICO AND THE VIRGIN ISLANDS

By EDWARD A. CHAPIN,
Curator, Division of Insects, U. S. National Museum.(1)

As it will probably be some considerable time before a projected revision of the Puerto Rican and Virgin Island species of Scarabaeidae can appear, it seems desirable to establish names for certain new species which have come to my attention. I am indebted to Dr. Stuart T. Danforth, Mr. R. G. Oakley and to the American Museum of Natural History for the gift or loan of the material described below.

Canthochilum hispidum, new species

Piceous, legs dark castaneous, antennae and mouth parts paler. Entire dorsal surface set with very short, capitate hairs, most densely placed on pronotum and along the elytral striae. Head nearly smooth, minutely punctured except for two oval patches of coarser punctures on the vertex, one patch on either side of the slightly elevated median line. Eyes large, transverse diameter of eye equal to one-fifth distance between eyes. Clypeus quadridentate with middle pair of teeth acute and slightly recurved, genal angles also slightly prominent, each tooth or prominence with a tuft of setae above and below. Pronotum transverse, almost twice as wide as long, basal margin very broadly curved, sides parallel to apical fifth where they turn suddenly toward the median line, anterior angles distinct, posterior angles wanting, underside deeply excavated in anterior third for reception of anterior femora, excavation bounded by a strong carina. Surface strongly shagreened, densely and moderately coarsely punctured. Elytra more shining, less convex than pronotum, very finely striate, seventh stria bounded almost to apex by the strong lateral carina which is broadly interrupted just before its extremity, third interspace with a strong, blunt, subapical spine. Eight striae composed of large punctures and located on the inflexed portion along a second carina. Under surface smooth, without setae, sternites crowded along median line, pygidium subtriangular, margin strongly beaded. Anterior femora strong, not alate, middle femora small and slender, tarsi of anterior legs feeble, strongly compressed, those of other legs well developed, not noticeably compressed, heavily clothed with pale pubescence. Claws minute.

Male: Pronotum strongly gibbous, anterior tibia slender, somewhat bent and enlarged in apical fourth, with three lateral teeth, of which the basal is a little remote from the others and with a single ventral tooth near insertion of tarsus; posterior femora broadly expanded in middle portion.

(1) Published with permission of the Secretary of the Smithsonian Institution.

Female: Pronotum moderately convex, anterior tibia moderately broad, with three lateral teeth situated in the apical third but without a ventral tooth.

Length: 5.5 — 6.2 mm.

Type locality: Villalba, Puerto Rico.

Type and twelve paratypes: U. S. Nat. Mus. Cat. No. 51079.

Material examined: Eight males and five females from the Insular Government Finca at Villalba, collected Sept. 7, 1934 by Mr. R. G. Oakley.

Because of the vestiture of capitate hairs, all specimens seen of this species have been almost completely covered with a layer of dirt, most difficult to remove.

Canthochilum andyi, new species

Piceous to deep black, anterior margin of head and legs castaneous, antennae and mouth parts paler. Resembles *C. histeroides* Harold but larger and with more strongly punctured elytral striae. Head almost smooth, most finely shagreened and with a patch of minute punctures on vertex between eyes. Clypeus quadridentate, the middle teeth more slender than the lateral, almost equidistantly spaced, genal angles slightly prominent, each tooth or prominence with hair tufts above and below. Eyes large, nearly divided by canthus. Pronotum transverse, visibly shagreened only at sides, not visibly punctured except under high magnification; basal margin very broadly rounded, sides parallel to apical fourth, thence sharply convergent to the obtuse anterior angles. Under side excavated in anterior third for reception of femora, excavation bounded posteriorly by carina. Elytra distinctly shagreened, striae well defined and moderately coarsely punctured, the punctures larger toward apical extremity. Seventh stria bounded by lateral carina which ends near apical fourth. Eight stria on inflexed portion and bounded by a carina which reaches almost to extreme apex of suture. Under parts and pygidium as in preceding species. Anterior femora thickened, anterior tibiae coarsely denticulate in addition to the usual three teeth.

Male: Pronotum somewhat gibbous, posterior femora broadly expanded in middle portion, anterior tibiae rather slender, the teeth moderately long and acute in fresh specimens, middle tibiae somewhat, posterior tibiae strongly, arcuate and gradually widened to apices.

Female: Pronotum not more convex than is usual, posterior femora not expanded, anterior tibia moderately broad, middle and posterior tibiae feebly arcuate and gradually widened to apices.

Length: 5 — 5.5 mm.

Type locality: Matrullas, Puerto Rico.

Type and fifteen paratypes: U. S. Nat. Mus. Cat. No. 51080.

Material examined: Four males and twelve females from the Insular Government Finca at Matrullas Dam, near Orocovis, collected October 8, 1934 by Mr. R. G. Oakley.

Cyclocephala danforthi, new species

Similar to *Cyclocephala vincentiae* Arrow in size and form. Testaceous, vertex piceous. Lower portion of frons and upper portion of clypeus moderately strongly and confluently punctured, lower portion of clypeus shining, clypeal margin feebly reflexed laterally, strongly so at the broadly rounded anterior margin. Antenal club of male very large, one third longer than distance between eyes. Pronotum with lateral margins subangulate at middle, sparsely, and rather finely punctured. Scutellum very finely punctured at sides. Elytra with faintly indicated rows of punctures, the interspaces rather coarsely and sparsely punctured. Underparts slightly darker in color than upper, metasternum finely and closely punctured except at middle and on either side adjacent to hind coxae. Pygidium closely and finely granulate. Anterior tibiae tridentate, anterior tarsus of male short and stout, the third to fifth segments with striated areas on inner faces, fifth segment strongly inflated. Claws of anterior tarsus very dissimilar in size, the larger broad and bifurcated in apical fourth, the rami dissimilar and widely divergent.

Length: 11 mm.

Type locality: Great Bay, St. Martin, Virgin Islands.

Type: U. S. Nat. Mus. Cat. No. 51081; paratype in collection of Dr. S. T. Danforth.

Material examined: Two males from the above-mentioned locality colected Dec. 23, 1927, by Dr. S. T. Danforth.

Epiphileurus puertoricensis, new species

Near *Epiphileurus cribratus* Chevrolat but larger with less coarsely punctured pronotum and with quite different aedeagus. Head sparsely set with ill defined punctures, frons of male with a deep, circular pit which is puncture-free, of female slightly concave with a few coarse punctures. Apex of clypeus acute and reflexed, horns over the antennal insertion short, stout and obtuse. Pronotum with an ill-defined median groove which bears a few medium-sized ocellate punctures, rest of surface sparsely punctured, the punctures coarse on disc either side of median groove and very fine along the lateral and basal margin. Anterior angles acute and produced, posterior angles rounded. Elytra with rows of ocellate punctures, the rows somewhat paired, so that the sutural, second, fourth and sixth interspaces are more elevated than the first, third and fifth. Lateral portions of underside with large, ocellate punctures. Pygidium coarsely and rather sparsely punctured, strongly convex in male, less so in female. Anterior tibia tridentate, the teeth slender and acute. Anterior tarsi similar in the sexes.

Length: 15 — 16 mm.

Type locality: Villalba, Puerto Rico; also from Barranquitas, P. R.

Type and nine paratypes: U. S. Nat. Mus. Cat. No. 51082.

Material examined: One male from the Insular Government, Finca, Villalba, collected Oct. 26, 1933, by Mr. R. G. Oakley; seven

males and one female from same locality, June 29, 1934, Oakley; one male and three females from Barranquitas, collected Dec. 1930 by Mr. R. Colón and received for study from Dr. S. T. Danforth and American Museum of Natural History. Two paratypes, male and female, returned to Dr. Danforth, one female paratype returned to the American Museum.

Phyllophaga yunqueana, new species

Large, subdepressed, color yellow brown with head, pronotum and elytra rich reddish castaneous with a dense bluish-white bloom conspicuous only in certain lights. Legs pale with tibiae and tarsi slightly darker. Head moderately coarsely, evenly but sparsely punctured, clypeo-frontal suture deep, clypeus slightly tumid, a little more sparsely punctured than frons, margin gradually reflexed, median indentation broad and shallow. Antenna nine-segmented, club shorter than stem. Pronotum with sides subangulate at middle, anterior angles not at all produced, basal angles rounded, lateral marginal bead broken at intervals, a long hair arising from each break. Elytra with sutural margin slightly tumid, apices of suture minutely mucronate, discal costae obsolete, punctures similar in size to, but more densely placed than those on pronotum. Epipleura very narrow except at base. Pygidium strongly convex in male, coarsely and subconfluently punctured. Underparts rather densely and finely punctured at sides, sparsely so at middle. Anterior tibia strongly tridentate in both sexes, the upper tooth somewhat remote from middle. Tarsi moderately long, posterior tibia slender, hardly expanded at apex, calcaria slender, the longer noticeably longer than the first tarsal segment. Claws strongly curved, the median tooth strong.

Length: 23 mm.

Type locality: El Yunque Mt., Luquillo Range, Puerto Rico.

Type: U. S. Nat. Mus. Cat. No. 51083. Paratype in American Museum of Natural History.

Material examined: A male collected February 1900 by Dr. Leonhard Stejneger; a female collected Oct. 4, 1919, in the collection of the American Museum of Natural History.

Phyllophaga discalis, new species

Below medium size, yellowish to reddish brown, elytra and flanks of pronotum reddish brown, head and disc of pronotum deep castaneous. Head with frons coarsely and confluently punctured, with a deeply impressed median line, clypeo-frontal suture deep, clypeus with punctures similar to those on frons, margin narrowly reflexed, median indentation broad, shallow and not angulate. Antennae nine-segmented, club shorter than stem. Pronotum notably transverse and convex, sides subangulate at middle, anterior angles obtuse, not produced, basal angles rounded, lateral marginal bead broken in a few places and with long hairs at breaks. Elytra with sutural margin slightly tumid, apices of sutural bead minutely mucronate, discal costae absent, rather more finely and densely punctured than pronotum. Epipleura very narrow. Pygidium only slightly convex in male, sparsely and not coarsely punctured.

Under parts finely and densely punctured at sides, very sparsely so at middle. Anterior tibia tridentate in both sexes, the basal tooth a little remote from middle. Posterior tibia slender, not greatly expanded apically. Calcaria of posterior tibia narrow, the longer half again as long as the first tarsal segment. Tarsi moderately long, claws not strongly curved, tooth stout, situated a little beyond middle.

Length: 13 — 15 mm.

Type locality: Yauco, Puerto Rico.

Type and paratypes: U. S. Nat. Mus. Cat. No. 51084.

Material examined: A male from Natalie Finca, in the mountains north of Yauco, Aug. 9, 1934, collected in the ground by Mr. R. G. Oakley; one male and one female from Añasco, P. R., Oct. 16, 1930, by Mr. J. A. Zalduondo.

The aedeagus of this species departs radically from the usual type found among the West Indian species. It is elongate tubular, with the apical margin somewhat modified and bears a striking resemblance to the type commonly found among the species of the Philippines. I have checked the specimens against our entire Asiatic series without success. The similarity may be due to convergent evolution or it may be that at some time Asiatic cane has been introduced into the island.

NITRIFICATION STUDIES WITH SOIL TYPES IN NORTHERN PUERTO RICO

By JUAN AMEDEE BONNET, *Chief of Soils Division,*
Agricultural Experiment Station, University of Puerto Rico, Río Piedras, P. R.

SCIENTIFIC POINTS RELATIVE TO SOIL NITRIFICATION

The suggestion of Pasteur in 1862 that nitrification is due to bacterial action led Winogradsky in 1890 to isolate the organisms concerned in nitrification processes. Since then, nitrification studies have been a subject of investigation in various soils located in countries of different climatic conditions. Among the various aspects studied is worth while mentioning: Nitrification as influenced by soil moisture, temperature, seasonal variations, plant growth, carbon-nitrogen ratio, buffering agents, application of fertilizers and amendments; occurrence, isolation, limiting and optimum pH of soil nitrifying bacteria; nitrification and solubilization of certain soil inorganic elements; methods of studying soil nitrification and nitrifying capacity of a soil as an index of fertility.

It is a well known fact that the soil nitrifying organisms are of an autotrophic nature characterized by the property of obtaining carbon from the carbon dioxide of the atmosphere and their energy by the oxidation of simple inorganic compounds in the form of ammonium salts, and that the process is carried on in two stages: first by a group of bacteria oxidizing ammonia to nitrites (nitrosomonas, nitrosococcus), and then by another group of bacteria oxidizing nitrites to nitrates (nitrobacter). But Gopala Rao and Dhar (13) claim that nitrification in the soil is at least partly photochemical, taking place without the agency of bacteria under the action of sunlight at the surface of various soil photocatalysts like alumina and titania. In further experiments Gopala Rao (12) says: "The slow oxidation of ammonia in aqueous solution to nitrite has been shown to take place in transparent silica vessels under the action of light from a quartz mercury vapor lamp, copper arc or more slowly in sunlight." He also calls the attention to the fact that nitrification in the atmosphere and seasonal variations of the nitrate-nitrite contents in favor of the summer months can be successfully explained on the basis of the photochemical view. Fraps and Sterges (**10 b**) have shown that photonitrification is of little or no practical importance.

Two laboratory methods have been used for the study of nitrification: solution or sand method and the most used soil or tumbler method. Although various inorganic and organic nitrogenous sources have been used for such studies, ammonium sulphate and dried blood have been preferred. The solution or sand method supplies information as to the absence or presence of nitrifying bacteria, influence of stimulating substances present in the soil, etc. This method is limited to the use of nitrogenous inorganic sources; since organic materials produce soluble organic compounds and ammonia which are toxic to the nitrifying bacteria. The sensitivity is more pronounced to organic substances in solution than in soil.

It is of great importance in nitrificaton studies to control the concentration of the nitrogenous salt, the pH range, the time and temperature of incubation and the optimum content of soil moisture. The usual concentration recommended for ammonium sulphate is 30 milligrams of nitrogen per 100 grams dry soil in the presence or absence of 210 milligrams of calcium carbonate which is equivalent to an addition of the theoretical amount of base necessary for the complete neutralization of all the nitric and sulphuric acids formed from the complete oxidation of the nitrogen added. For nitrification of organic nitrogenous materials, 0.25 per cent of organic matter with a high nitrogen content (10–12 per cent), such as dried blood, or 0.5–1.0 per cent of organic materials of a low nitrogen content (cotton-seed meal, soy-bean meal, alfalfa meal) should be employed. The usual incubation time and temperature is 30 days at 25–28°C. For the solution or sand method a temperature of 28 to 30°C, is recommended. The optimum amount of water used in the tumbler method is from 50 to 60 per cent saturation.

Conditions which tend to promote nitrate formation in the soil are: temperature of 27.5°C., an abundant supply of air (oxygen), proper moisture supply, a favorable reaction (pH greater than 4.6), presence of carbonates or other buffering agents and absence of large quantities of soluble organic matter and alkali salts in the soil. The nature of the crop and the season also influences the nitrate content of the soil.

Although a definite correlation between the nitrifying power of a soil and its crop productivity has been observed by various investigators, some others reported that there may or may not be a correlation, and that continuous cropping, especially without fertilization, reduces the nitrifying capacity of the soil. The subject is fully discussed by Waksman (**32, 33**). Such a correlation may be limited

by some factor other than the nitrogen supply, such as moisture, temperature, aeration.

Fraps and Sterges (**10 a**) summarize studies on low nitrification capacity of soils as follows: "Soils which do not nitrify ammonium sulphate may be caused to nitrify it by addition of cultures of actively nitrifying soil, of calcium carbonate, or of both nitrifying culture and calcium carbonate. Nitrifying organisms may remain in a dry soil for many years. Nitrites may be produced from ammonium sulphate when calcium carbonate is added, though few nitrates may be produced at the same time."

From the physiological point of view nitrates is no longer considered to be the only form in which nitrogen is absorbed by certain plants. Tiedjens and Robins (**29**) found that ammonium hydroxide was a much better source of nitrogen than either sulphate of ammonia or calcium nitrate for the tomato and soybean and that ammonia was no more toxic to plants than were nitrates. They further comment: "Pirschle grew wheat, oats, corn, tobacco, cucurbits, peas, beans, soybeans and rape with ammonium sulphate, ammonium chloride, potassium nitrate, and calcium nitrate. He found that in a neutral culture some plants produced as good growth with ammonia as with nitrate salts or even better, but that an acid and even slightly alkaline reaction, nitrates were superior in most cases. Shive and Stahl state that seedlings absorb more ammonia than nitrate nitrogen but that mature plants require more nitrate nitrogen." In a more recent publication, Tiedjens (**30**) discusses experimental evidence on factors affecting assimilation (synthesis to amino acids and other organic nitrogenous materials) of nitrogen.

Nitrification Studies With Tropical Soils

Antipov–Karataev (**2**) in nitrification studies with soil from the Nikita orchard, Crimea, found: "Intensity of nitrification in orchard soils is similar to that of fallow in the chernozem district. Tobacco and virgin soils give the same amount of nitrates as chernozem soils under corn or sorghum after a fallow. In the shaly soil, low in lime, the process of nitrification was slow. During the period of intensive growth of tobacco no accumulation of nitrates was noted."

Prescott (**26**) and Roche (**28**) studied nitrification of Egyptian soils. Prescott says: "Nitrification is well ahead of the needs of the cotton crop, and probably entirely accounts for the fact that nitrogenous fertilizers produce no effect on this crop. There is no accumulation of nitrates in the soil when wheat and maize are grown."

The work of Prescott also covers observations of seasonal variations of nitrates and pot experiments showing the effect of the growth of maize and wheat on the accumulation of nitrates. The work of Roche includes some observations of rate of nitrification in soils under irrigation.

Peck (**23, 24**) carried some nitrification experiments with soils of Hawaii, using the tumbler method. His results showed that blackstrap molasses from sugar cane removes temporarily part of the available nitrates. Nitrificaton of ammonium sulphate was checked and nitrification of nitrogenous organic fertilizers was retarded by adding molasses. Kelley (**20**) and Burgess (**8**) also studied nitrification in soils of Hawaii. Kelley found that nitrification was as active in the manganese and titanium soils as in other soils, but magnesium carbonate was especially toxic. Burgess reports the following data:

Productivity of Soil	Nitrifying power (Mgm. nitrogen per 100 gms. dry soil)	
	Dried blood	Alfalfa meal
Best	20.8	15.2
Very good	15.2 — 20.0	9.6 — 12.8
Poorer	4.0 — 13.6	7.2 — 9.0
Poorest	4.0	4.5

In 1927, Bal (**4**) found in a black soil in India, planted to cotton, that concentrations of over 100 milligrams of ammoniacal nitrogen from ammonium hydroxide are definitely injurious to the process of soil nitrification. Plymen and Bal (**25**) studied nitrification rates of different nitrogenous organic manures in some typical soils of Berar, India. Hutchinson (**17**) summarizes investigations on nitrification in India soils. Joshi (**19**) reports on rate of nitrification of different green manures and influence of crop residues on nitrification in India soils. Walton (**35**) studied the influence of alkali salts on nitrification in some India soils and also (**34**) the rate of nitrification of calcium cyanamide as compared with ammonium sulphate and mustard cake. Batham (**5**) compares nitrification rate of certain amino acids and ammonium sulphate in India soils.

Itano and Arakawa (**18**) report studies in the rice fields of Japan on relation of nitrification to crop yield of rice, seasonal variations and soil specific buffer capacity.

Wilcox (**37**) presents a soil nitrification map of the Bandjaratma concession of Java as prepared by Arrhenius who recommends, as sound soil management for a sugar-cane plantation, a map, to show the distribution of the soil's nitrifying power. Arrhenius correlated soil nitrifying power with sugar-cane yields and showed that soil reaction does not parallel nitrate production.

Gerretsen (**11**) in work with Java soils found that the concentration of ammonium salts through soil adsorption may be so high that nitrification becomes impossible. He claims that the intensity of nitrification in different soils does not necessarily have any direct effect on crop production, and its importance as an index of fertility has often been overrated, although the two frequently run parallel.

Pañganiban (**22**) reports nitrification studies, with ammonium sulphate and dried blood, in Philippine soils planted to yautias, corn, banana and cogon. Aquino and Javier (**3**) report nitrification studies in eighteen different soil types of the Philippine Islands, and Alicante (**1**) found: "Treatment of citrus soils constitute conclusive evidence that nitrification is an index of crop production. In general, soils devoted to sugar cane converted their original nitrogen into available form more slowly than did those devoted to rice, abacá, tobacco, citrus, etc. Soils containing a high percentage of clay nitrified either the original or the added nitrogen very poorly." He gives data on the average nitrifying power of some of the Philippine soils as compared to those of Hawaii and the United States.

Watt (**36**) studied nitrification in Transvaal soils. Martin and Massey (**21**) studied the effect of seasonal variation on the nitrification rate of Sudan soils. Hall (**14**) studied nitrification rate of some South African soils supplemented with additions of limestone, dried blood, dried cowpea hay, bone meal, ammonium sulphate, whale guano, calcium cyanamide, crayferine and sewage soil. He also studied seasonal variations of nitrates in virgin, cropped and cultivated lands and nitrate variation with soil depth. In further studies, Hall (**15**) studied nitrification in some acid soils of South Africa treated with superphosphate, and ammonium sulphate in the presence or absence of lime. He also reports nitrification data in tobacco soils.

Chardón (**9**) studied nitrification rate of dried blood in the presence or absence of calcium carbonate in an acid clay around the College of Agriculture at Mayagüez, Puerto Rico. Ramírez (**27**)

studied the rate of nitrification of filter press cake residue from a sugar-cane factory added to the red acid clay close to the Insular Experiment Station at Río Piedras, Puerto Rico. He found that nitrate nitrogen from the application to a soil of 15, 25 and 50 tons of filter press cake per acre begins to increase on the third month, goes to a maximum on the fifth month, then the tendency towards the sixth or seventh month is to decrease.

Experimental

Part I contains some of the unpublished research work done by the author in the New Jersey Agricultural Experiment Station, under the direction of the soil microbiologist, Dr. Selman A. Waksman, as contained in a thesis presented in partial fulfillment of the requirement for the degree of Master of Science. The main part of said thesis entitled "Nitrogen transformations in the decomposition of sugar-cane trash, with special bearing upon Puerto Rico soil problems" has been published (**6**) elsewhere. It also contains some essential data from that thesis reported in a published paper (**7**) entitled "Preliminary microbiological studies in certain soils of the San Juan area, Puerto Rico".

Part II contains the complete laboratory data of the work done from 1930–1934 in fulfillment of the requirements of a research project of the Insular Experiment Station entitled: "Nitrification studies for a typical soil type in each classified soil series in Puerto Rico." The limitations of our means allowed only to undertake studies with soils from the northern coast of Puerto Rico.

Soils

The preliminary soil survey maps and reports which are a part of the Soil Survey of Puerto Rico as undertaken since 1928 by the United States Bureau of Chemistry and Soils in cooperation with the Insular Experiment Station were used as a basis for soil location, classification and description. The soil types were taken from a list, tentatively called "North Coast Area of Puerto Rico", which was correlated and approved by the Soil Survey Division of the United States Bureau of Chemistry and Soils on March 16, 1934.

Soil Types: The following soil types are included in these studies:

1. Catalina clay. This soil is a deeply weathered soil of the mountain and hill lands. It is derived from andesitic tuffs. The surface ranges from rolling to steep; however, most of the hills are

rounded with gently sloping ravines. It varies in color from light red to reddish-brown. This soil is well drained but is affected by sheet erosion. It has a friable clay surface whose depth varies with surface relief from about four to eight inches. The subsoil is a reddish-brown or light-red, slightly heavy, but friable clay. At a depth of about 24 inches the soil becomes a deep red clay which continues to great depths. This soil is usually acid. It is used for a variety of crops as cane, citrus, pineapples, coffee and minor crops; and produces profitable yields. This is one of the best coffee soils.

2. Catalina clay level phase. This is the Catalina clay with a nearly level to slightly undulating relief. It is more deeply weathered than the main type as it has not been affected by sheet erosion. Its surface soil is deeper. Farmers prefer it to the main type for most all crops. It is used mostly for cane, citrus and pineapples; all do very well on it.

3. Coto clay light textured phase. This type occurs on nearly level surface relief. It is characterized by a dark grayish-brown, permeable, friable clay about a foot thick underlain by a reddish-brown, slightly compact, permeable clay subsoil and a yellowish-brown, non-plastic clay lower subsoil which usually rests on hard limestone at about 6 feet, but may vary from 26 inches to 12 feet. It is just slightly acid. The most profitable crop adapted to this soil is sugar cane. Other crops producing a good profit are Spanish pepper, tomatoes, corn, ñames, cotton, beans and bananas.

4. Espinosa clay. This type is the acid counterpart of the Coto clay, but has a less yellowish subsoil more mottled in the lower subsoil. In many places the subsoil is quite red. It occurs on gently undulating surfaces in large and small valleys and generally in very well drained positions. It is characterized by a 10-inch surface horizon of light brown to reddish yellow somewhat stiff and finely cloddy clay upper subsoil, and 15 to 30 inches or more of reddish-yellow friable clay which is sometimes mottled red in the deeper horizons. Limestone, the original parent material, occurs at greatly varying depths. It is used almost entirely for sugar-cane production.

5. Lares clay loam. This soil belongs to a group of deeply weathered soils with medium friable subsoils that occur on terraces or on terrace-like positions, with level to slightly undulating relief. It is affected by gully erosion. It is characterized by a heavy clay loam surface with a brown to reddish-brown color that crushes fairly easily into granules and some rounded quartz grains. At a depth

of about 6 inches the soil changes to red or brownish red, heavy, medium plastic clay that cracks some on drying; in the lower part of this layer usually at a depth of only about 14 inches is mottled red and yellowish-medium friable clay. At about 40 inches is a friable red, yellow and brown clay that continues to great depth. In places it looks very much like disintegrated tuff. A characteristic feature of this soil is the noticeable amount of water rounded rocks on this soil and throughout the profile. Many of the rocks are flat and pitted. The rocks near the surface are coated with a layer about 1/16 of an inch thick, consisting of a dark mineral, like that composing the perdigon, although very few perdigons are noticed except where this soil grades into Sabana Seca. In the lower depths there will often be gravel layers resembling water deposited gravel. These rounded gravel are of shale, andesite and tuffs. This soil is acid in all layers. It has been influenced by materials from other soil series (Río Piedras and Cialitos). It is used mostly for sugar cane and citrus.

6. Múcara silty clay loam. This undulating to rolling hill soil is easily recognized by its brown surface and light-brown subsoil over a brown igneous rock. It is a medium weathered soil characterized by a dark brownish gray or grayish-brown, gritty silty clay loam surface that is stained lightly in the lower part with gray. This layer is underlain at a depth of about 9 inches by a yellowish, slightly plastic clay or clay loam that has grayish or rust colored mottles. In places small angular fragments of partly weathered rock material occur in this layer and in places the clay is moderately tough and compact. The lower subsoil is a yellowish, light textured, transitional layer consisting of gritty clay loam mixed with partly decomposed rocks. The depth to rock, depending upon the slope, varies from 14 inches to 3 feet. It is affected by sheet erosion. Sugar cane, beans, pigeon peas, corn, yautía, etc., are among the crops planted. Soil depth limits crop yields.

7. Múcara silt loam. This soil differs from the silty clay loam type in having a lighter texture surface and slightly less heavy subsoil.

8. Sabana Seca clay. This is a fertile soil but not very productive because of its heavy stiff subsoil that resists ready penetration of air, plant roots and water. It occurs on nearly level to undulating surface relief, is very acid, and is probably derived from marine deposits. It is characterized by a dark brown, medium compact clay

surface. At a depth of about 5 to 7 inches there is a thin sub-surface layer about 3 inches thick of light yellow, heavy, medium compact clay; this abruptly changes at about 10 inches to an extremely heavy, stiff, plastic, mottled red, rich brown, gray and yellow clay. This layer continues to a depth of about 30 inches where it becomes gradually lighter in texture and slightly less compact; however, at depths below six feet the soil is still heavy mottled and compact. Cane and pasture are the two best crops grown.

9. Toa silt loam. This is an alluvial, well drained, loose and porous soil developed from material washed from the shale and limestone hills. It is the best sugar-cane soil in the North Coast of Puerto Rico. It is characterized by a grayish-brown color, nearly uniform in both color and texture to a depth of several feet and has very little mottling in the subsoil.

Location of soil samples: The samples used in part I were taken to a depth of six inches with a shovel, from a single representative spot not under cultivation.

Soil Types Used in Part I	Location
Catalina clay level phase	Km. 2.25 sideroad from Trujillo Alto mainroad to Leper Colony.
Lares clay loam	Km. 2.2 Sanatorium road, Río Piedras.
Múcara silty clay loam	Km. 2.8 Carolina-Juncos road.
Toa silt loam	Km. 10.3 San Juan-Bayamón road.

The next chart indicates the location of soil samples used in part II. Samples numbered 1–15 inclusive were taken also to a depth of six inches with a shovel in representative spots not under cultivation. Samples 16 and 17 were taken all over the field to a depth of twelve inches with a soil auger; since these samples were also used for other chemical work.

Sample Number	Date	Soil types used in part II	Location	Sampling places
1	August 5, 1930	Lares clay loam	Km. 2.2 Sanatorium road	1
2	August 5, 1930	Lares clay loam	Km. 2.0 Bayamón–Slaughterhouse road	1
3	August 5, 1930	Lares clay loam	Km. 2.8 Bayamón–Comerío road	1
4	March 11, 1931	Toa silt loam	Julia farm close to Central Constancia, Toa Baja	6
5	March 5, 1931	Toa silt loam	Central Canóvanas, opposite main office	3
6	March 11, 1931	Toa silt loam	Nevares farm at km. 1.8 of side road to Central Constancia	6
7	March 11, 1931	Toa silt loam	San Antonio farm at km. 10.3 San Juan–Bayamón road	6
8	March 5, 1931	Toa silt loam	Tamarindo farm, at km. 26.6 Río Grande–Mameyes road	3
9	March 11, 1931	Toa silt loam	El Naranjal farm, 1 km. N. Carolina	3
10	March 5, 1931	Múcara silty clay loam	Km. 45 Luquillo–Fajardo road	3
11	March 11, 1931	Múcara silty clay loam	Km. 9.9 Guaynabo–La Muda road	6
12	March 5, 1931	Múcara silt loam	La Carmen farm on El Verde side-road at km. 26.6 Río Grande–Mameyes road	3
13	April 27, 1930	Coto clay-light texture phase	Km. 24 Quebradillas–Guajataca road	1
14	April 27, 1930	Coto clay-light texture phase	Km. 8.5 Arecibo–Lares road	1
15	April 28, 1932	Espinosa clay	Km. 68.8 Arecibo–Barceloneta	1
16	Nov. 24, 1933	Sabana Seca clay	University farm Río Piedras	About 30
17	August 20, 1933	Catalina clay, sample A	Mr. Juan Esteva's farm, Lares	23

Soil preparation: All samples in part I were sifted in the air-dried state through a 2 mm. sieve. All samples in part II were sifted in the fresh state.

Soil analysis and treatments: Moisture and soil total water holding capacity was then determined.

Soils were treated in duplicate according to the Waksman's (31) tumbler method, i.e.

"1. Nitrification of soil's own nitrogen. A definite amount of soil (100 gm.) kept in the laboratory for a definite length of time (30 days) at a definite temperature (25–28° C.), under optimum moisture conditions, will give us information on the forms of nitrogen present in the particular soil and the speed with which they are transformed into nitrates and thus made available for plant growth.

"2. Nitrification of ammonium sulphate in the soil. By using a definite amount of nitrogen 30 mgm. in 100 gm. of soil, in the form of ammonium sulphate, and standard period of incubation, we get, from the amount of nitrate formed, an index on the buffering capacity of the soil in relation to nitrification. The final reaction should always be recorded.

"3. Nitrification of ammonium sulphate in the presence of a theoretical amount of $CaCO_3$, 210 mgm. for 30 mgm. N as $(NH_4)_2SO_4$ necessary to neutralize all the acid formed from the complete oxidation of the ammonium sulphate into nitric and sulfuric acid. This gives an index of the nitrifying capacity of the soil under optimum reaction conditions and forms an excellent basis for comparing nitrification with other biological activities."

Nitrification of soil's own nitrogen in the presence of the theoretical amount of $CaCO_3$ (210 mgm. for 100 gms. dry soil), as used in the ammonium sulfate treatment, was also studied.

In all, but one of the experiments, 60 percent of the soil total water holding capacity was taken for optimum moisture condition. The moisture lost by evaporation during the incubation period of 30 days was replaced at intervals.

Nitrates and ammonia were determined after 15 and 30 days, respectively. In most of the samples those determinations were also made at the start. The usual pH determinations were made after the 30-day period. Some pH determinations were also run at the start.

The phenoldisulphonic colorimetric method was used for nitrates.

In part II, ammonia was determined by direct distillation with magnesia. In part I, ammonia was determined by the Harper's (16) modification which consists in extracting the ammonium ion in the soil with normal potassium chloride and then distilling with magnesia. The determinations for pH were made with the potentiometer using quinhydrone and a saturated calomel-potassium chloride electrode.

I. RESEARCH WORK DONE IN NEW JERSEY AGRICULTURAL EXPERIMENT STATION

The soil type, Sassafras sandy loam, sampled on June 24th, 1929, from the experimental plots of the New Jersey Agricultural Experiment Station at New Bunswick was used as a basis for comparison.

Nitrification Studies of Puerto Rico Soils at 25 per cent Moisture Content

A twenty-five percent moisture content was chosen because it represents optimum moisture conditions for the Sassafras sandy loam. This is a productive soil derived from the unconsolidated sands and clays of the coastal plain.

TABLE I

NITRIFICATION RATE AT TWENTY-FIVE PER CENT MOISTURE
UNTREATED SOIL

Soils	pH		Mgm. of Nitrogen per 100 gms. dry soil as:			
			Nitrate		Ammonia	
	Start	30 da.	Start	30 da.	Start	30 da.
Toa silt loam	5.8	6.0	5.9	7.4	1.1	3.4
Catalina clay level phase	6.7	6.6	5.7	5.7	1.4	6.8
Lares clay loam	4.7	4.6	2.4	2.7	0.9	4.9
Múcara silty clay loam	6.8	6.1	4.8	9.2	1.6	Trace
Sassafras sandy loam	5.7	5.9	6.3	6.5	Trace	Trace

SOIL + 30 mgm. OF NITROGEN AS $(NH_4)_2SO_4$ + $CaCO_3$ (210 mgm.)

Soils	pH	Mgm. of Nitrogen per 100 gms. dry soil as.							
		Nitrate				Ammonia			
	30 da.	Start	7 da.	14 da.	30 da.	Start	7 da.	14 da.	30 da.
Toa silt loam	7.1	5.9	6.7	8.3	9.4	31.1	20.3	18.7	15.1
Catalina clay level phase	7.4	5.7	6.1	6.0	5.3	31.4	32.2	33.1	33.6
Lares clay loam	5.4	2.4	2.6	2.9	2.7	30.9	31.3	31.7	30.5
Múcara silty clay loam	7.0	4.8	7.1	9.9	11.7	31.6	25.7	24.6	20.3
Sassafras sandy loam	5.4	6.3	14.6	22.2	30.0	30.5	19.8	11.0	2.4

Under the conditions of that experiment there was no nitrate accumulation from the soil's own nitrogen sources in, and from the ammonium sulfate added to, the soil types from Puerto Rico known as Catalina clay level phase and Lares clay loam, although those soils had sufficient ammonia to start the nitrification processes.

In the other soils, the rate of nitrate accumulation goes hand to hand with ammonia disappearance.

The soil from New Jersey had a higher nitrifying rate; but we must consider that the 25 percent moisture content was probably not the optimum one for the two soils from Puerto Rico.

At the start, the pH of all the soils was in the acid range. Even with the addition of the calcium carbonate, the Lares clay loam had a pH of 5.4, after the 30-day period. The New Jersey soil had that same pH, but its nitrification rate was quite active. The pH for the other soils, at the end of the 30-day period was close to the neutral point, in the alkaline side.

Influence of Lime and Inoculation upon Nitrification in Two Puerto Rico Soils

In an attempt to find the limiting factor responsible for checking nitrification in the Catalina clay level phase and Lares clay loam, these soils were treated as before; but this time, 500 milligrams of calcium carbonate were added, and treatments also included inoculation of the soils with one cubic centimeter of a 10 percent water infusion of the Sassafras sandy loam. All the soils were kept at a 25 percent moisture content.

TABLE 2

EFFECT OF LIME AND INOCULATION UPON NITRIFICATION RATE OF CATALINA CLAY LEVEL PHASE AND LARES CLAY LOAM

Treatment	pH	Mgm. of Nitrogen per 100 grams of dry soil			
		As Nitrate		As Ammonia	
	30 da.	Start	30 da.	Start	30 da.
Catalina clay level phase	6.0	5.7	6.4	1.4	7.6
Catalina clay level phase + $(NH_4)_2SO_4$ (30 mgm. N.)	6.8	5.7	7.2	31.4	37.5
Catalina clay level phase + $(NH_4)_2SO_4$ + $CaCO_3$ (500 mgm.)	7.9	5.7	5.2	31.4	34.6
Catalina clay level phase + $(NH_4)_2SO_4$. Inoculated	6.8	5.7	10.0	31.4	36.1
Catalina clay level phase + $(NH_4)_2SO_4$ + $CaCO_3$. Inoculated	7.9	5.7	4.8	31.4	31.7
Lares clay loam	5.1	2.4	2.8	.9	2.2
Lares clay loam + $(NH_4)_2SO_4$ (30 mgm. N.)	5.1	2.4	3.0	30.9	34.6
Lares clay loam + $(NH_4)_2SO_4$ + $CaCO_3$ (500 mgm.)	7.8	2.4	2.2	30.9	34.6
Lares clay loam + $(NH_4)_2SO_4$. Inoculated	5.1	2.4	2.2	30.9	36.1
Lares clay loam + $(NH_4)_2SO_4$ + $CaCO_3$. Inoculated	7.8	2.4	2.6	30.9	35.3

The Lares clay loam did not show nitrate accumulation in any of the different treatments. Reaction was not the limiting factor; since the calcium carbonate added, was enough to bring the pH towards the alkaline side.

The addition of lime did not favor nitrate accumulation in the Catalina clay level phase. Perhaps the alkalinity (pH 7.9) did not favor the activity of the nitrifying organisms. That same activity

was checked at (pH 7.4) in the previous experiment. Where ammonium sulfate was added alone there was a slight accumulation of nitrates, more so, in the inoculated soil. The reaction in those cases was (pH 6.8). Alkaline reactions or the presence of an excess of calcium ions have limiting effects on the activity of the nitrifying organisms in this soil. The next experiment will help to clear these points and indicate whether the effect of inoculation on nitrate accumulation may be considered as significant.

Nitrification Studies of Puerto Rico Soils at Optimum Moisture

To favor conditions for the activity of the nitrifying organisms the Puerto Rico soils were brought to optimum moisture at 60 percent of the total water holding capacity. This work has been reported elsewhere (**8**). (In that paper the soil type reported as Río Piedras clay should be Catalina clay level phase; the Bayamón clay loam should be Lares clay loam, and the Múcara clay should be Múcara silty clay loam.) The rate of nitrification of dried blood was also studied. The amount added was 250 milligrams containing 32 milligrams of nitrogen. The calcium carbonate added was 500 milligrams.

TABLE III

NITRIFICATION RATE OF SOILS AT OPTIMUM MOISTURE

(MGM. N PER 100 GMS. DRY SOIL)

Treatments	Catalina clay level phase					Lares clay loam				
	pH	Nitrate			Amm.	pH	Nitrate			Amm.
	30 da.	Start	15 da.	30 da.	30 da.	30 da.	Start	15 da.	30 da.	30 da.
Soil alone	6.5	5.7	5.5	5.5	8.6	5.0	2.4	1.6	1.8	7.6
Soil + $CaCO_3$ (500 mgm.)	7.2	.....	5.2	5.0	5.7	7.1	.....	1.9	2.2	9.4
Soil + dried blood (32 mgm. N)	6.6	.....	3.8	5.0	27.4	5.3	.....	3.0	3.4	24.9
Soil + $(NH_4)_2SO_4$ (30 mgm. N)	6.4	.....	5.2	5.7	35.3	4.6	.....	1.8	1.7	33.8
Soil + $(NH_4)_2SO_4$ + $CaCO_3$	6.4	.....	4.8	10.6	13.3	7.0	.....	1.8	1.5	34.6

TABLE III

NITRIFICATION RATE OF SOILS AT OPTIMUM MOISTURE

(MGM. N PER 100 GMS. DRY SOIL)

Treatments	Múcara silty clay loam					Toa silt loam				
	pH	Nitrate			Amm.	pH	Nitrate			Amm.
	30 da.	Start	15 da.	30 da.	30 da.	30 da.	Start	15 da.	30 da.	30 da.
Soil alone	6.3	4.8	8.0	12.3	2.2	6.6	5.9	6.1	7.3	6.5
Soil + $CaCO_3$ (500 mgm.)	7.4	.....	8.0	13.3	2.2	7.6	.....	7.0	10.7	1.5
Soil + dried blood (32 mgm. N)	6.5	.....	8.0	13.3	18.0	6.9	.....	7.0	8.6	18.0
Soil + $(NH_4)_2SO_4$ (30 mgm. N)	6.0	.....	6.7	12.0	25.2	6.3	.....	5.6	6.1	23.7
Soil + $(NH_4)_2SO_4$ + $CaCO_3$	7.1	.....	7.3	27.9	8.6	7.1	.....	8.0	19.4	9.4

The rate of nitrification of the soil's own nitrogen under optimum conditions was almost identical to that under the 25 percent moisture content reported in Table I. All soils were able to ammonify dried blood. The accumulation of nitrates from dried blood was only evident in the Múcara silty clay loam and Toa silt loam.

Let us compare now the nitrification rate of ammonium sulfate in the presence of lime, under these conditions, with that of the Sassafras sandy loam reported in Table I.

Soil Type	Optimum moisture %	pH		Milligrams of Nitrogen per 100 gms. dry soil as:				
				Nitrate			Ammonia	
		Start	30 da.	Start	15 da.	30 da.	Start	30 da.
Lares clay loam	32.8	4.7	7.0	2.4	1.8	1.5	30.9	34.6
Catalina clay level phase	40.0	6.7	6.4	5.7	4.8	10.6	31.4	13.3
Toa silt loam	40.0	5.8	7.1	5.9	8.0	19.4	31.1	9.4
Múcara silty clay loam	36.0	6.8	7.1	4.8	7.3	27.9	31.6	8.6
Sassafras sandy loam	25.0	5.7	5.4	6.3	22.2	30.0	30.5	2.4

Optimum moisture and pH conditions did not favor nitrification in the Lares clay loam; but were effective in showing that the Catalina clay level phase contains active nitrifying organisms able to convert ammonium sulfate into nitrates and that the organisms were not affected by excess of calcium ions.

The rate of nitrification after the 15-day period, for the Puerto Rico soils, is identical with 25 percent and optimum moisture contents. Striking difference is observed after the 30-day period. The nitrification rate was favored in all soils; but the Lares clay loam. Taking the rate of nitrification on the New Jersey soil as 100 percent, the nitrifying Puerto Rico soils, at the end of the 30-day period, compare as follows:

	Percentage
Múcara silty clay loam	99.67
Toa silt loam	64.67
Catalina clay level phase	35.33

II. RESEARCH WORK DONE IN THE INSULAR STATION OF PUERTO RICO

Nitrification studies were continued in Puerto Rico, with new samples of soils sifted in the fresh state.

Further Nitrification Studies with Lares clay loam at Optimum Moisture Conditions

As the previous experiments indicated lack of nitrate accumulation in the Lares clay loam, three fresh soil samples were taken from different places. Sample #2 was taken from the same place as that one used in New Jersey. Nitrification of ammonium dibasic phosphate was also studied. The optimum moisture for samples #1 and #2 were 45 and 36 percent, respectively.

TABLE IV (a)

NITRIFICATION RATE OF LARES CLAY LOAM

(MGM. N PER 100 GMS. DRY SOIL)

Tumbler No.	Tratments	Sample (No. 1)						Sample (No. 2)					
		pH	Nitrate			Ammonia		pH	Nitrate		Ammonia		
		30 da.	Start	15 da.	30 da.	15 da.	30 da.	30 da.	Start	15 da.	30 da.	15 da.	30 da.
1.....	Soil.....	4.6	.8	1.7	2.5	5.5	5.5	4.9	Tr.	1.3	2.3	7.6	5.6
2.....	Soil + $CaCO_3$ (210 mgm.)	5.9	.....	3.1	5.5	7.6	11.1	5.9	.....	1.6	3.1	8.8	7.6
3.....	Soil + $(NH_4)_2SO_4$ (30 mgm. N).....	4.7	.....	1.9	2.5	38.5	39.9	4.9	.....	1.8	1.5	36.4	35.8
4.....	Soil + $(NH_4)_2SO_4$ + $CaCO_3$ (210 mgm.).....	5.0	.....	4.1	8.6	37.1	33.0	6.0	.....	1.7	2.9	35.7	35.8
5.....	Soil + $(NH_4)_2SO_4$ + $CaCO_3$ (360 mgm.).....	5.3	.....	18.3	23.9	23.4	15.1	6.3	.....	1.4	5.4	38.5	33.0
6.....	Soil + $(NH_4)_2HPO_4$ (30 mgm. N).....	5.0	.....	2.2	3.4	35.8	37.3	5.1	.....	1.8	2.5	32.3	33.0
7.....	Soil + $(NH_4)_2HPO_4$ + $CaCO_3$ (210 mgm).....	4.7	.....	8.3	20.3	24.8	22.0	5.8	.....	2.2	3.8	33.0	30.4

Both ammonium salts added to sample #1 are able to nitrify in the presence of lime. The higher amount of lime (360 mgm.) favored nitrification of ammonium sulfate in both samples, but in sample #2 the nitrification rate was quite low. Please recall that this fresh sample was taken from the same place as that one used in New Jersey.

It was decided to study nitrification rate of the same ammonium sulfate up to a 60-day incubation period. The moist soils left after the 30-day period in treatments 3, 4 and 5 of sample #2 were modified, as shown below, either by adding more calcium carbonate or

potassium chloride. The amounts added are in terms of 100 grams dry soil. A and B are duplicates from the 30-day period treated differently. Sample 5A was left as the check.

TABLE IV (b)

EFFECT OF THE EXTENSION OF THE INCUBATION PERIOD AND PRESENCE OF POTASH AND MORE LIME ON THE NITRIFICATION RATE OF A POOR NITRIFYING, LARES CLAYLOAM

Tumber No.	Treatment given to sample (No. 2) after the 30 day period	pH		Mgm. N per 100 gms. dry soil as:			
				Nitrate		Ammonia	
		30 da.	60 da.	30 da.	60 da.	30 da.	60 da.
3A	Soil + $(NH_4)_2SO_4$ + $CaCO_3$ (500 mgm.)	4.9	7.1	1.5	4.4	35.8	39.5
3B	Soil + $(NH_4)_2SO_4$ + $CaCO_3$ (750 mgm.)	4.9	7.2	1.5	3.9	35.8	43.7
4A	Same as 3A	6.0	6.8	2.9	7.7	35.8	31.5
4B	Same as 3A + K_2O (30 mgm.)	6.0	6.7	2.9	6.9	35.8	31.5
5A	CHECK: Soil + $(NH_4)_2SO_4$ + $CaCO_3$ (360 mgm.)	6.3	6.1	5.4	8.7	33.0	18.7
5B	Soil + $(NH_4)_2SO_4$ + $CaCO_3$ (500 mgm.) + K_2O (100 mgm.)	6.3	6.4	5.4	13.3	33.0	14.3

Results indicate that the rate of ammonium sulfate nitrification of Lares clay loam is limited by the length of the incubation period and the absence of lime and potash.

Nitrification rate of ammonium sulfate and ammonium dibasic phosphate in Lares clay loam

Nitrification rate of ammonium sulfate and ammonium dibasic phosphate in the presence of more lime and potash was studied in sample #2 of Lares clay loam which showed slight evidence of nitrification after the 30-day period.

TABLE V

NITRIFICATION RATE OF AMMONIUM SULFATE AND AMMONIUM DIBASIC PHOSPHATE IN A POOR-NITRIFYING LARES CLAY LOAM

Treatment given to Lares clay loam Sample No. (2)	pH	Mgm. N per 100 gms. dry dry soil as:			
		Nitrate		Ammonia	
	30 da.	15 da.	30 da.	15 da.	30 da.
Soil + $(NH_4)_2SO_4$ (30 mgm. N) + $CaCO_3$ (500 mgm.)..	6.7	.5	2.2	48.6	41.3
Soil + $(NH_4)_2SO_4$ + $CaCO_3$ + K_2SO_4 (100 mgm. K_2O)..	6.4	.6	1.7	44.9	44.1
Soil + $(NH_4)_2HPO_4$ (30 mgm. N) + $CaCO_3$ (500 mgm.)...	6.3	.9	7.4	40.2	29.4
Soil + $(NH_4)_2HPO_4$ + $CaCO_3$ + K_2SO_4 (100 mgm. K_2O).	6.3	.7	7.1	39.8	37.0

Dibasic ammonium phosphate nitrifies in the presence of 500 milligrams of calcium carbonate without the influence of potash. The ammonium sulfate showed a very slight tendency to nitrify in the presence of sufficient lime and potash. It seems therefore, that the presence of the phosphate ion in the presence of sufficient lime favors the nitrification process in this soil.

Nitrification studies in another sample of Lares clay loam

Nitrification studies of ammonium sulfate and ammonium dibasic phosphate were made in a sample of Lares clay loam from a different location. The treatments given are shown in Table VI.

TABLE VI.

NITRIFICATION RATE OF ANOTHER SAMPLE OF LARES CLAY LOAM

Treatments given to Sample (No. 3). Optimum moisture-30%	pH	Mgm. N. per 100 gms. dry soil as				
		Nitrate			Ammonia	
	30 da.	Start	15 da.	30 da.	15 da.	30 da.
Soil	5.6	Tr....	1.7	2.4	9.9	8.9
Soil + $CaCO_3$ (360 mgm.)	7.1	.....	7.7	6.7	7.1	8.1
Soil + $(NH_4)_2SO_4$ (30 mgm. N)	5.5	.....	1.3	2.0	40.1	39.5
Soil + $(NH_4)_2SO_4$ + $CaCO_3$ (360 mgm.)	5.3	.....	25.0	19.3	11.5	7.2
Soil + $(NH_4)_2SO_4$ + $CaCO_3$ (500 mgm.)	6.4	.....	13.8	36.4	9 9	3.6
Soil + $(NH_4)_2HPO_4$ (30 mgm. N) + $CaCO_3$ (360 mgm.)	6.0	.....	13.8	36.4	7.8	2.9
Preceding treatment + KCl (100 mgm. K_2O)	5.7	.....	8.5	19.3	16.5	7.2

The amount of lime was a dominant factor in the nitrification of both ammonium salts. The ammonium sulfate maximum nitrification was obtained with 500 milligrams of calcium carbonate. That same maximum nitrification was obtained with the ammonium dibasic phosphate; but in the presence of less lime (360 mgm. $CaCO_3$). The potash had no effect.

NITRIFICATION STUDIES IN SIX SAMPLES OF TOA SILT LOAM

Sample Number	Location	Optimum moisture (60% of total water holding capacity
4	Central Constancia, Toa Baja	47.70
5	Central Canóvanas	35.40
6	Nevares farm, Toa Baja	40.20
7	San Antonio farm, Bayamón	43.80
8	Tamarindo farm, Río Grande	39.90
9	El Naranjal farm, Carolina	36.00

TABLE VII

TABLE VII

NITRIFICATION RATE OF TOA SILT LOAM

(MGM .N PER 100 GMS. DRY SOIL)

Treatments	Soil Number (4)							Soil Number (5)							Soil Number (6)						
	pH		Nitrate			Ammonia		pH		Nitrate			Ammonia		pH		Nitrate			Ammonia	
	Start	30 da.	Start	15 da.	30 da.	15 da.	30 da.	Start	30 da.	Start	15 da.	30 da.	15 da.	30 da.	Start	30 da.	Start	15 da.	30 da.	15 da.	30 da.
Soil	6.3	6.2	N.D*	2.5	4.5	5.2	6.6	5.8	5.2	.4	2.7	3.1	7.4	6.6	5.7	5.7	N.D*	4.0	4.9	7.4	7.4
Soil + $CaCO_3$ (210 mgm.)	.....	6.7	.....	4.3	4.7	9.5	8.1	.....	5.9	.....	2.9	3.8	6.6	5.9	.....	6.1	.....	4.8	5.9	7.4	5.9
Soil + $(NH_4)_2SO_4$ (30 mgm. N)	.....	5.5	.....	23.5	26.7	11.1	5.9	.....	4.8	.....	5.3	10.8	23.5	18.3	.....	6.5	.....	15.4	15.4	15.4	11.1
Soil + $(NH_4)_2SO_4$ + $CaCO_3$	.....	5.9	.....	28.8	26.7	6.6	7.4	.....	5.1	.....	18.2	21.6	14.0	8.1	.....	5.7	.....	34.8	23.5	4.2	4.5

TABLE VII

NITRIFICATION RATE OF TOA SILT LOAM

(MGM .N PER 100 GMS. DRY SOIL)

Treatments	Soil Number (7)							Soil Number (8)							Soil Number (9)						
	pH		Nitrate			Ammonia		pH		Nizrate			Ammonia		pH		Nitrate			Ammonia	
	Start	30 da.	Start	15 da.	30 da.	15 da.	30 da.	Start	30 da.	Start	15 da.	30 da.	15 da.	30 da.	Start	30 da.	Start	15 da.	30 da.	15 da.	30 da.
Soil	6.1	5.7	4.8	4.5	4.9	5.9	7.4	5.6	5.3	.4	4.8	7.4	5.2	5.9	7.1	6.2	.4	3.6	2.9	2.9	5.2
Soil + $CaCO_3$ (210 mgm.)	.....	6.4	.....	4.1	4.9	8.0	7.4	.....	6.3	.....	6.2	6.3	5.9	6.6	.....	6.6	.....	3.9	3.8	7.4	5.2
Soil + $(NH_4)_2SO_4$ (30 mgm. N)	.....	5.4	.....	6.6	10.3	24.4	19.9	.....	5.2	.....	6.8	8.3	39.1	25.8	.....	5.8	.....	4.8	8.7	25.8	21.4
Soil + $(NH_4)_2SO_4$ + $CaCO_3$	.....	5.5	.....	12.5	21.1	22.1	6.7	.....	5.4	.....	8.7	14.8	23.5	11.8	.....	6.0	.....	14.8	23.5	10.4	6.6

*N. D.—Not determined.

NITRIFICATION STUDIES IN THREE SAMPLES OF MÚCARA SOILS

Sample Number	Soil Type	Location	Optimum moisture (60% of total water holding capacity
10	Múcara silty clay loam	Luquillo	37.2
11	Múcara silty clay loam	Guaynabo	36.3
12	Múcara silt loam	Río Grande	39.6

TABLE VIII

NITRIFICATION RATE OF MÚCARA SOILS

(MGM. N PER 100 GMS. DRY SOIL)

Treatments	Soil Number (10)						Soil Number (11)						Soil Number (12)					
	pH	Nitrate			Ammonia		pH	Nitrate			Ammonia		pH	Nitrate			Ammonia	
	30 da.	Start	15 da.	30 da.	15 da.	30 da.	30 da.	Start	15 da.	30 da.	15 da.	30 da.	30 da.	Start	15 da.	30 da.	15 da.	30 da.
Soil	6.7	Tr...	5 9	11.8	3.6	4.5	6.1	Tr...	.5	1.3	6.7	8.1	5.5	Tr...	4.9	17.2	15.5	4.5
Soil + $CaCO_3$ (210 mgm.)	7.4	.....	6.3	13.8	4.5	2.9	6.3	.....	1.8	6.9	6.7	2.9	5.9	.....	8.2	18.9	8.1	2.9
Soil + $(NH_4)_2SO_4$ (30 mgm. N)	6.7	.....	11 1	21.7	5.2	3.6	5.4	.....	.5	1.0	35.6	35.6	5.8	.....	3.8	16.3	11.4	28.1
Soil + $(NH_4)_2SO_4$ + $CaCO_3$	7.0	.....	11.1	25.7	4.5	2.9	5.3	.....	3 7	18.5	31.1	12.6	5.8	.....	8.3	27.8	29.5	5.9

The Toa silt loam is a good nitrifying soil for ammonium sulfate in the absence or presence of lime. The addition of lime does not favor the nitrification rate of the soil's own nitrogen.

The presence of lime favored nitrification in all samples of Múcara soils. Samples #10 and #12 are good nitrifiers of the soil's own nitrogen and of the ammonium sulfate added.

NITRIFICATION STUDIES IN COTO CLAY-LIGHT TEXTURE PHASE AND ESPINOSA CLAY

Sample Number	Soil Type	Location	Optimum moisture (60% of total water-holding capacity
13	Coto clay light texture phase	Quebradillas	42.8
14	Coto clay light texture phase	Arecibo	35.4
15	Espinosa clay	Barceloneta	40.1

TABLE IX

NITRIFICATION RATE OF COTO CLAY LIGHT TEXTURE PHASE (MGM. N PER 100 GMS. DRY SOIL)

Treatments	Sample Number (13)						Sample Number (14)					
	pH		Nitrate		Ammonia		pH		Nitrate		Ammonia	
	Start	30 da.	15 da.	30 da.	15 da.	30 da.	Start	30 da.	15 da.	30 da.	15 da.	30 da.
Soil	6.0	5.6	4.1	5.8	12.1	12.8	6.1	5.7	2.6	3.6	12.9	12.1
Soil+$CaCO_3$(2!0 mgm.)	.....	5.7	5.2	6.6	12.9	12.9	.....	6.0	3.2	4.4	12.9	12.9
Soil + $(NH_4)_2SO_4$(30 mgm. N)	.....	5.4	13.0	22.2	27.7	26.3	.....	5.3	6.9	10.0	31.8	13.6
Soil + $(NH_4)_2SO_4$ + $CaCO_3$	.....	5.2	16.2	36.4	21.0	12.9	.....	5.2	29.6	40.0	13.5	12.2

TABLE X

NITRIFICATION RATE OF ESPINOSA CLAY

(MMG. N PER 100 GMS. DRY SOIL)

Treatments	pH		Nitrate			Ammonia	
	Start	30 da.	Start	15 da.	30 da.	15 da.	30 da.
Soil	6.6	5.8	5.6	6.3	8.8	13.6	13.6
Soil + $CaCO_3$ (210 mgm.)	.....	5.9	.....	6.6	8.9	14.8	14.8
Soil + $(NH_4)_2SO_4$ (30 mgm. N)	.....	5.6	.....	6.6	14.0	37.9	25.8
Soil + $(NH_4)_2SO_4$ + $CaCO_3$	.....	5.5	.....	11.1	17.4	18.3	13.6

The Coto and Espinosa soils are able to nitrify the ammonium sulfate in the absence of lime.

NITRIFICATION STUDIES IN SABANA SECA CLAY AND CATALINA CLAY

Sample Number	Soil Type	Location	Optimum moisture (60% of total water holding capacity
16	Sabana Seca Clay	Río Piedras	27.6
17	Catalina Clay Plot (A)	Lares	39.

TABLE XI

NITRIFICATION RATE OF SABANA SECA CLAY

Treatments	pH		Mgm. N per 100 gms. dry soil as:			
			Nitrate		Ammonia	
	Start	30 da.	15 da.	30 da.	15 da.	30 da.
Soil	5.4	5.3	.7	3.3	9.1	5.0
Soil + $CaCO_3$ (210 mgm.)	.....	6.5	1.2	5.4	9.8	5.0
Soil + $(NH_4)_2SO_4$ (30 mgm. N)	.....	5.3	.7	3.6	40.0	38.
Soil + $(NH_4)_2SO_4$ + $CaCO_3$	.....	5.8	.6	8.3	41.4	28.7

TABLE XII

NITRIFICATION RATE OF CATALINA CLAY

(MGM. N PER 100 GMS. DRY SOIL)

Treatments	pH		Nitrate		Ammonia	
	Start	30 da.	15 da.	30 da.	15 da.	30 da.
Soil	4.6	4.8	.9	.9	9.1	10.5
Soil + $CaCO_3$ (210 mgm.)		5.1	.8	1.3	9.8	11.1
Soil + $(NH_4)_2SO_4$ (30 mgm. N)		4.7	.7	.9	40.3	40.9
Soil + $(NH_4)_2SO_4$ + $CaCO_3$		5.1	.7	1.1	43.5	45.5

Lime helps the nitrification process of the Sabana Seca clay.

The treatments given to the Catalina clay were not effective in starting the nitrification process. The amount of lime added (210 mgm. $CaCO_3$) was not enough to correct the excess acidity of this soil.

GENERAL DISCUSSION

Let us now compare in table XIII the nitrification rate, at the end of the thirty-day period, of all the soils worked out, under similar conditions, in the preceding experiments.

Table XIII gives valuable information on the relation of soil pH to nitrate and ammonia accumulation. It also compares the nitrification and ammonification rate of the different soil types and the extent of variation of such processes within various samples of the same soil type. It offers valuable information as to whether ammonium sulfate should be used as source of nitrate for plant assimilation and whether it should prove advisable to lime the soil in order to hasten the nitrifying rate of ammonium sulfate or to use nitrate salts as a source of nitrogen.

Results indicate that ammonium sulfate should not be used as a source of nitrate for plants in the Lares clay loam, Sabana Seca clay and Catalina clay, and in some of the Múcara silty clay loam, unless special attention is given to lime broadcasting. The lime might be partially beneficial in the Lares clay loam and Catalina clay. It should also prove advisable, in those soils and in Sabana Seca clay, to apply part of the nitrogen as nitrate salts, preferably calcium nitrate.

Ammonium sulfate nitrifies quite well in the Toa silt loam, Coto clay light texture phase and Espinosa clay, even in the absence of lime. The application of ammonium sulfate as a source of nitrate

TABLE XIII

COMPARISON OF NITRIFICATION RATE OF SOME PUERTO RICO SOILS AND A NEW JERSEY SOIL AT OPTIMUM MOISTURE ON THE BASIS OF NITRIFICATION RATE AFTER THIRTY DAY PERIOD

(MGM. N. PER 100 gms. DRY SOIL)

Sample No.	Soil Type	Soil's own nitrogen				Ammonium sulfate (30 mgm. N)				Ammonium sulfate* (30 mgm. N) and $CaCO_3$ (210 mgm.)			
		pH	Nitrate	Ammonia	Total N*	pH	Nitrate	Ammonia	Total N*	pH	Nitrate	Ammonia	Total N*
1.....	Lares clay loam.....	4.6	2.5	5.5	8.0	4.7	2.5	39.9	42.4	5.0	8.6	33.0	41.6
2.....	Lares clay loam.....	4.9	2.3	5.6	7.9	4.9	1.5	35.8	37.3	6.0	2.9	35.8	38.7
Ave.....	**Lares clay loam.....**	**4.8**	**2.4**	**5.6**	**8.0**	**4.8**	**2.0**	**37.9**	**39.9**	**5.5**	**5.8**	**34.4**	**40.2**
4.....	Toa silt loam.....	6.2	4.5	6.6	11.1	5.5	26.7	5.9	32.6	5.9	26.7	7.4	34.1
5.....	Toa silt loam.....	5.2	3.1	6.6	9.7	4.8	10.8	18.3	29.1	5.1	21.6	8.1	29.7
6.....	Toa silt loam.....	5.7	4.9	7.4	12.3	6.5	15.4	11.1	26.5	5.7	23.5	4.5	28.0
7.....	Toa silt loam.....	5.7	4.9	7.4	12.3	5.4	10.3	19.9	30.2	5.5	21.1	6.7	27.8
8.....	Toa silt loam.....	5.3	7.4	5.9	13.3	5.2	8.3	25.8	34.1	5.4	14.8	11.8	26.6
9.....	Toa silt loam.....	6.2	2.9	5.2	8.1	5.8	8.7	21.4	30.1	6.0	23.5	6.6	30.1
Ave.....	**Toa silt loam.....**	**5.7**	**4.6**	**6.5**	**11.1**	**5.5**	**13.4**	**17.1**	**30.5**	**5.6**	**21.9**	**7.5**	**29.4**
10.....	Múcara silty clay loam.....	6.7	11.8	4.5	16.3	6.7	24.7	3.6	28.3	7.0	25.7	2.9	28.6
11.....	Múcara silty clay loam.....	6.1	1.3	8.1	9.4	5.4	1.0	35.6	36.6	5.3	18.5	12.6	31.1
Ave.....	**Múcara silty clay loam.....**	**6.4**	**6.6**	**6.3**	**12.9**	**6.1**	**12.9**	**19.6**	**32.5**	**6.2**	**22.1**	**7.8**	**29.6**
12.....	Múcara silt loam.....	5.5	17.2	4.5	21.7	5.8	16.3	28.1	44.4	5.8	27.8	5.9	33.7
13.....	Coto clay light texture phase	5.6	5.8	12.8	18.6	5.4	22.2	26.3	48.5	5.2	36.4	12.9	39.3
14.....	Coto clay light texture phase	5.7	3.6	12.1	15.7	5.3	40.0	13.6	53.6	5.2	40.0	12.2	52.2
Ave.....	**Coto clay light texture phase.....**	**5.7**	**4.7**	**12.5**	**17.2**	**5.4**	**31.1**	**20.0**	**51.1**	**5.2**	**38.2**	**12.6**	**50.8**
15.....	Espinosa clay.....	5.8	8.8	13.6	22.4	5.6	14.0	25.8	39.8	5.5	17.4	13.6	31.0
16.....	Sabana Seca clay.....	5.3	3.3	5.0	8.3	5.3	3.6	38.8	42.4	5.8	8.3	28.7	37.0
17.....	Catalina clay.....	4.8	.9	10.5	11.4	4.7	.9	40.9	41.8	5.1	1.1	45.5	46.6
	Sassafras sandy loam, New Jersey.....	5.9	6.5	Trace	6.5	.....	.....	.....	.....	5.4	30.0	2.4	32.4

* Total N is the sum of nitrate and ammonia.

for plant growth should prove beneficial in those soils without considering the practice of lime broadcasting.

The thirty milligrams of nitrogen added as ammonium sulfate was accounted for, as nitrate and ammonia in all the soils. The slight excess of total nitrogen found in some cases, was due to the formation of ammonia by those active soil organisms able to decompose the soil's own protein material.

Let us now compare the average results given in Table XIII on a percentage basis. The results obtained with the Toa silt loam were taken as a basis for the comparison, since that alluvial soil is the most valuable and productive in the northern coast of Puerto Rico.

TABLE XIV

COMPARISON OF NITRIFICATION RATE ON AVERAGE BASIS

Soil Types	Thirty Day Nitrifying Period					
	Soil's own nitrogen		Ammonium sulfate (30 mgm. N)		Ammonium sulfate + $CaCO_3$ (210 mgm.)	
	Nitrate (Mgm. N)	Percent	Nitrate (Mgm. N)	Percent	Nitrate (Mgm. N)	Percent
Toa silt loam	4.6	100	13.4	100	21.9	100
Lares clay loam	2.4	52	2.0	15	5.8	26
Múcara silty clay loam	6.6	143	12.9	96	22.1	101
Múcara silt loam	17.2	374	16.3	122	27.8	127
Coto clay light texture phase	4.7	102	31.1	232	38.2	175
Espinosa clay	8.8	191	14.0	104	17.4	79
Sabana Seca clay	3.3	72	3.6	27	8.3	38
Catalina clay	.9	20	.9	7	1.1	5
Sassafras sandy loam	6.5	141	.....	.....	30.0	137

SUMMARY

Experimental results have been reported on the nitrification rate of nine soil types from Northern Puerto Rico classified as: Catalina clay, Catalina clay level phase, Coto clay light texture phase, Espinosa clay, Lares clay loam, Múcara silty clay loam, Múcara silt loam, Sabana Seca clay and Toa silt loam. The various soil types are represented by twenty-one different soil samples. Some experimental data is also reported for a soil type from New Jersey classified as Sassafras sandy loam.

The main object of the work was to establish a correlation on the following bases:

1. Nitrification of soil's own nitrogen.
2. Nitrification of soil's own nitrogen as affected by the addition of lime.
3. Nitrification of ammonium sulfate in the soil.
4. Nitrification of ammonium sulfate as affected by the addition of lime.

Studies were also carried with various samples of the same soil type in order to study the nitrifying range within the soil type.

All soils were similarly treated with respect to incubation period, temperature, and time; optimum moisture conditions; and concentration of lime and ammonium sulfate added to the soil.

The soil types known as Catalina clay level phase and Lares clay loam which showed a slight response to the above general treatments were also studied with respect to the influence of more lime and inoculation upon nitrification. One soil sample of Lares clay loam which also showed a slight response to inoculation and extra lime addition was also studied with respect to the effect of the extension of the incubation period and addition of potash in the presence of various amounts of lime. The nitrification rate of ammonium sulfate and ammonium dibasic phosphate was also compared in that soil.

Table XIII summarizes the experimental results comparing the nitrification and ammonification rate of the different soil types similarly treated and the variations of such a rate within various samples of the same soil type. Those results should prove of value in the practical application of ammonium sulfate as a fertilizer and indicate if it is advisable to lime the soil in order to hasten the nitrification rate of ammonium sulfate or to apply part of the nitrogen in the form of nitrate salts.

Table XIV contains data on the comparison of the nitrification rate of the various soils treated on a percentage basis. The most productive and valuable alluvial soil type of the north coast of Puerto Rico, Toa silt loam, was taken as the unit basis.

Conclusions

1. Ammonium sulfate should not be used as a source of nitrate for plants in the Lares clay loam, Sabana Seca clay and Catalina clay, and in some fields of Múcara silty clay loam, unless special attention is given to lime broadcasting. It should be more advisable to apply ammonium dibasic phosphate instead of ammonium sulfate to Lares clay loam, after special attention is given to lime broad

casting. Although the lime might be beneficial in the Lares clay loam and Catalina clay, it should also prove advisable, in those soils, and in Sabana Seca clay, to apply part of the nitrogen as nitrate salts, preferably calcium nitrate.

2. Ammonium sulfate nitrifies quite well in the Toa silt loam, Coto clay light texture phase, and Espinosa clay, even in the absence of lime. The application of ammonium sulfate as a source of nitrate for plant growth should prove beneficial in those soils without considering of upmost importance the practice of lime broadcasting.

Literature Cited

1. **Alicante, M.** Nitrifying power of some Philippine soils. Philippine Journ. Sci. **32**(1) : 1–27, 1927.
2. **Antipov-Karataev, I. N.** (Nitrification in the soils of the Nikita orchard in the year 1925). Journ. Gvt. Bot. Gard. Nikita Yalta. Crimea **9**(2) : 54 p., 1926.
3. **Aquino, D. I. & Javier, V. I.** Nitrification studies of certain Philippine soils. Philippine Agr. **21**(5) : 309–317, 1932.
4. **Bal, D. V.** Effect of varying concentrations of ammonia on the nitrifying power of the soil. Agr. Journ. India **22**(4) : 298–300. 1927.
5. **Batham, H. N.** Nitrification in soils. Soils Sc. **20**(5) : 337–351, 1925.
6. **Bonnet, J. A.** Nitrogen transformations in the decomposition of sugar cane trash, with special bearing upon Puerto Rico soil problems. Journ. Dept. Agric. Puerto Rico **15**(2) : 113–146, 1931.
7. ———— Preliminary microbiological studies in certain soils of the San Juan area. Puerto Rico. Int. Soc. of Sugar Cane Technologists. Fourth Congress. Bul. 58, 6 p., 1932.
8. **Burgess, P. S.** Can we predict probable fertility from soil biological data? Soil Sc. **6**(6) : 449–462, 1918.
9. **Chardón, Carlos E.** Estudio preliminar sobre la amonificación y nitrificación de los terrenos de Puerto Rico. (Preliminary studies on ammonification and nitrification of the soils of Puerto Rico.) Rev. Agric. Puerto Rico. **6**(1) : 37–46, 1921.

10 a. **Fraps, G. S., & Sterges, A. J.** Causes of low nitrification capacity of certain soils. Soil Sc. **34**(5) : 353–363, 1932.

10 b. Effect of sunlight on the nitrification of ammonium salts in soils. Soil Sc. **39**(2) : 85–94, 1935.

11. **Gerretsen, F. C.** (Nitrification and denitrification in tropical soils.) Arch Suikerind **29** : 1397–530, 1921.
12. **Gopala Rao, G.** Newer aspects of nitrification I. Soil Sc. **38** (2) : 143. 159, 1934.
13. **Gopala Rao, G., & Dhar, N. R.** Photosensitized oxidation of ammonia and ammonium salts and the problem of nitrification in soils. Soil Sc. **31**(5) : 379–384, 1931.

14. **Hall, T. D.** Nitrification in some South African soils. Soil Sc. **12**(4) : 301–363, 1921.
15. ———————— Nitrification in some South African soils. Part II. Soil Sc. **18**(3) : 219–235, 1924.
16. **Harper, H. J.** The determination of ammonia in soils. Soil Sc. **18**(5) : 409–418, 1924.
17. **Hutchinson, C. M.** Report of the Imperial Agr. Bacteriologist. Sci. Rpt. Agr. Res. Inst. Pusa 1918–19 : 106–14, 1919.
18. **Itano, Arao, & Arakawa, Satiyo.** Studies on the soils in rice fields. IV. Relation of ammonification, nitrification and denitrification of the soils to the crop yield of rice. Ber. Ohara Inst. Landw. Forsch. (Japan) **5**(1) : 151–161, 1931.
19. **Joshi, N. V.** Rate of nitrification of different green manures and parts of green manures and the influence of crop residues on nitrification. Agric. Journ. India Sci. Congress, No. 1919 : 395–413, 1919.
20. **Kelley, W. P.** Ammonification and nitrification in Hawaiian soils. Hawaii Agr. Expt. Sta. Bul. 37, 52 p., 1915.
21. **Martin, F. S. & Massey, R. E.** Nitrification in Sudan soils. Wellcome Trop. Res. Lat., Chem. Sect. Publ. **29,** 29 p., 1923.
22. **Pañganiban, E. H.** A study of nitrification in Philippine soils. Philippine Agr. and Forester **4**: 81–91, 1915.
23. **Peck, S. S.** Influence of molasses on nitrification in cane soils. Hawaii Sugar Planters Assoc. Expt. Sta. Chem. Ser. Bul. **39** : 1–23, 1912.
24. ———————— Some biochemical investigations of Hawaiian soils, with special reference to fertilizing with molasses. Hawaii Sugar Planters Assoc. Expt. Sta. Agr. Chem. Ser. Bul. **34** : 1–39, 1910.
25. **Plymen, F. J. & Bal, D. V.** The relative nitrifiability of different nitrogenous organic manures in some typical soils of the central provinces and Berar. Agr. Journ. India. **17**: 551–9, 1922.
26. **Prescott, J. A.** Nitrification in Egyptian solis. Journ. Agr. Sci. **9**(3) : 216–236, 1919.
27. **Ramírez, J. H.** Nitrificación de las tierras coloradas arcillosas ácidas de Puerto Rico. (Nitrificaction of the acid red clay soils of Puerto Rico). El Mundo Azucarero **10**(9) : 263–267, 1923.
28. **Roche, Raoul.** Study of the nitrification of Egyptian soil. Nag-Hamadi. Bul. de l'Institut Egyptien **5**(1) : 107–10, 1908.
29. **Tiedjens, V. A. & Robbins, W. R.** The use of ammonia and nitrate nitrogen by certain crop plants. New Jersey Agric. Expt. Sta. Bul. 526, 46 p., 1931.
30. **Tiedjens, V. A.** Factors affecting assimilation of ammonium and nitrate nitrogen, particularly in tomato and apple. Plant Physiology : **9**(1) : 31–57, 1934.
31. **Waksman, S. A.** Microbiological analysis of soil as an index of soil fertility. V. Methods for the study of nitrification. Soil. Sc. **15**(4) : 241–260, 1923.

32. **Waksman, S. A.** Microbiological analysis of soil as an index of soil fertility; VI. Nitrification. Soil Sc. **16**(1): 55–67, 1923.

33. ———————— Principles of soil microbiology. p. 695–699. London, Balliere, Tindall & Cox. 2d ed. 1931.

34. **Walton, J. H.** Nitrification of calcium cyanamide in some Indian soils. Mem. Dept. Agr. India, Bacteriol. Ser. **2**: 35–64, 1928.

35. ———————— The influence of alkali salts on nitrification in some Indian soils. Indian Journ. Agric. Sci. **1**(4): 480–494, 1931.

36. **Watt, R. D.** Nitrification in Transvaal soils. Transvaal Dept. Agr. Ann. Rept. 1908: 274–277, 1909.

37. **Wilcox, O. W.** The work of Arrhenius on Java cane soils. A summary of the investigations made for the Java Sugar Syndicate. Comparison with the Cuban Cane Soil Survey. Facts About Sugar **23**(36): 854–859, 1928.

HEAT STERILIZATION OF MANGOES AND GUAVAS FOR FRUIT FLIES

FRANCISCO SEÍN JR., *Assistant Entomologist,*
Agricultural Experiment Station, Río Piedras, Puerto Rico.

Some mango varieties and apparently all guavas in Puerto Rico are infested by fruit flies and thus all mango and guava varieties are excluded from exportation to the United States. There is practically no demand for guava fruits, but the present limited demand for mangoes would probably increase as that fruit became better known.

Mangoes and guavas are excluded under quarantine regulation No. 58 established in 1925 on the basis of infestation by the West Indian fruit fly, *Anastrepha fraterculus* Wiedeman.

According to Greene (**1**), *A. fraterculus* Wied., does not occur in Puerto Rico. In the opinion of the writer (**2**), a variety of the South American *A. fraterculus* Wied., occurs in Puerto Rico which he has described as *A. fraterculus* Wied., var., *mombinpraeoptans.* It is the one that breeds in some of the mango varieties. The species that breeds in guavas has been described by the writer (**2**) as *A. unipuncta.* According to Greene it is *A. suspensa* Loew. In this paper the two flies will be referred to as the species breeding in mangoes and the species breeding in guavas. Occasionally, when large numbers are reared, a few adults of the species breeding in mangoes will be found to breed also in guavas. That occurred in some of the experiments, but since it is not significant was not recorded.

The purpose of this paper is to report a series of tests under Puerto Rican conditions of the method of heat sterilization that has been used in Florida against the Mediterranean fruit fly and in Texas against the Mexican orange maggot and other fruit flies. The tests were made during the months of April, May, June and July 1934.

When the Mediterranean fruit fly was present in Florida, sterilized avocados from that State were allowed to be shipped to other parts of the country. The fruit had been grown in a regulated area outside the designated infested areas and was not infested. It was submitted to sterilization as an added precaution.

In the case of the mangoes that have commercial value in Puerto Rico, the conditions are similar in that the varieties to be sterilized have not been found infested and do not appear to be susceptible to infestation.

The sterilization treatment as described by Dr. Lee A. Strong in Administrative Instructions No. 287 (approved July 24, 1930) consists simply of "heating the fruit to a temperature of 110° F or above (not to exceed 112° F) in the approximate center of the fruit and holding the temperature of 110°F or above (not to exceed 112°F) for a period of eight hours".

No specifications as to the exact method or type of equipment to be used in sterilizing the fruit are prescribed. Available information indicates that a high humidity is essential for the best results and that the temperature should be maintained with as little variation as possible above the prescribed temperature of 110° F. For keeping the fruit after the sterilization it should be cooled to a temperature around 45° F as soon as possible.

Method of Procedure

The method of procedure and the equipment described by Mackie (3) in California served as a basis for planing the experiments and for constructing the sterilizer.

The sterilization experiments with mangoes and guavas were conducted at the entomological laboratory of the Insular Experiment Station in a sterilizer constructed by Mr. Manuel L. Vicente, Chief of the Division of Agricultural Engineering of the Station. It consists of two sections, a chamber for the fruit provided with a thermostat, thermometer, and racks for wire netting trays, and another section consisting of a water tank, heating unit with three heat switch, fan, motor and switches. The circulation is closed. The air becomes warm and saturated with moisture by passing over the hot water and heats the fruit chamber entering at the upper part, circulating around the fruit and returning to the water tank through an opening at the lower part of the chamber.

To conduct the experiments during office hours, the sterilizer was kept running on "low" during the previous night. The fruit was introduced the next morning at 7:00 A. M. The switch was then turned to "high" and as soon as the temperature reached 43° C (about 110° F) changed to "low". The thermostat maintained it uniformly at 43° C for eight hours.

Mango Varieties

There are two types of mangoes in Puerto Rico, the native seedlings and the more recently introduced selected East Indian varieties. In each type there are some varieties always infested with fruit fly maggots and others which have not been found infested at all and appear to be immune. Of the varieties that have not been found infested, the only native one that has commercial value is the "mango de Mayagüez" or Mayagüez mango. Of the recently imported varieties, the Colombo Kidney has high quality and is prolific. At Ponce there is a large grove of a variety that has been given the owner's name, Mr. Girón. In the island of Vieques there are some varieties that have commercial value. At present the only commercial variety grown on a large scale is the Mayagüez mango.

The Mayagüez mango is medium sized and attractively colored. It ships and keeps well but is very fibrous. There would be a good demand for it however, among West India residents in the United States. If the demand for mangoes higher in quality increased, they would be planted on a large scale.

Only Immune Varieties Have Commercial Value

Even in local markets, mangoes infested with fruit fly maggots have no commercial value. Their exportation to United States markets would be even less practical. Infested fruit can not be rendered marketable by sterilization because, though the maggots are killed, their dead bodies remain inside the fruits where they decompose, together with the pulp tissues previously injured by their tunneling.

The female fruit fly lays her eggs in the mangoes when green and the maggots may become full grown before the fruits are half ripe. For the mangoes to ripen properly they must be picked from the tree when at least half ripe. It would be impossible therefore to pick marketable fruit of the susceptible varieties early enough to prevent infestation.

The admission of sterilized mangoes into the United States is thus rendered doubly safe (1st) by the fact that only varieties that are not infested are marketable, and (2nd) because such sound fruit would nevertheless, and in addition, have been submitted to a treatment that kills the pest if it were present in them.

The Mango Variety Used in the Experiment

The mango variety used in the experiments is the one known as "mango blanco" or white mango. It was not chosen for its commercial possibilities but on the contrary, for its susceptibility to fruit fly infestation. A high percentage of the fruits of this variety is always infested. Since a large number of trees grow on the Station grounds and produce very large crops, procuring the fruit at the right stage of maturity was rendered easy.

The Guava Varieties Used in the Experiments

The guava fruits used in the experiments were of the two common varieties that grow on pastures, waste lands and fence rows on the Island. The two varieties have round-shaped medium sized fruits. The fruits of one variety are sweet, those of the other are sour. High percentages of the fruits of both varieties are ordinarily infested with fruit fly maggots. That does not however, destroy their value, for they are not consumed fresh but used for making preserves and jellies. For cooking purposes, most of the maggots can be made to abandon the fruit by soaking in cold water overnight.

Method of Handling the Fruit

The half ripe mangoes and guavas used in the experiments were picked from the trees the previous day. Some showed the emergence holes of the maggots opening through the cuticle. Emergence holes indicate that some full grown larvae may have left the fruit and also that full grown larvae are present in the fruit. Each lot of fruit to be sterilized or kept as checks consisted partly of fruit with emergence holes. When the fruit had cooled off after sterilization, some having emergence holes were cut open to determine the condition of the larvae. The rest of the sterilized froits and the checks were kept under observation on moist sand. The appearance of new emergence holes was noted. When the fruits began to rot they were cut open and inspected. The sand on which they stood was then sifted for pupae.

The presence of fruit fly eggs in the half ripe mangoes can be determined by observation of the tips which protude out of the cuticle (**2**). The species that ordinarily infests guavas lays its eggs entirely underneath the cuticle (**2**). Their presence in the fruit was inferred by the fact that the infestation of maggots in the checks was about normal.

Preliminary Experiments with Mangoes

Ten preliminary experiments were conducted using the first infested fruits of the 1934 crop which dropped from the trees and were picked from the ground in April. All showed emergence holes when collected.

After the sterilization, some dead maggots were observed on the floor of the chamber underneath the fruit. Some maggots were also found dead with their bodies partially out of the emergence holes. Cutting the fruits open, dead fruit fly maggots of various sizes were found. None was found alive. The cut open fruits were kept under observation on sand until the pulp rotted and inspected daily but no fruit fly maggots developed in them and no pupae were recovered by sifting the sand on which the fruit stood. The number of maggots found per fruit was as follows:

Number of fruits, 5____Number of maggots per fruit: 3, 6, 5, 7, 4
Number of fruits, 7____Number of maggots per fruit: 6, 2, 5, 4, 3, 4, 7
Number of fruits, 6____Number of maggots per fruit: 3, 5, 6, 2, 8, 5
Number of fruits, 8____Number of maggots per fruit: 4, 10, 5, 6, 9, 4, 6, 5
Number of fruits, 12____Number of maggots per fruit: 3, 7, 5, 6, 9, 4, 6, 5, 4, 8, 7, 6
Number of fruits, 15____Number of maggots per fruit: 4, 10, 5, 3, 7, 4, 6, 3, 9, 5, 7, 2, 4, 7, 6
Number of fruits, 8____Number of maggots per fruit: 5, 3, 7, 12, 6, 4, 5, 3
Number of fruits, 7____Number of maggots per fruit: 6, 9, 5, 5, 4, 8, 6
Number of fruits, 10____Number of maggots per fruit: 4, 7, 2, 5, 3, 6, 4, 8, 11, 5
Number of fruits, 7____Number of maggots per fruit: 12, 6, 3, 2, 5, 4, 6.

Final Experiments

When mangoes became abundant in the months of May, June and July 20 lots of 100 apparently infested fruits each were sterilized at different intervals keeping a lot of 100 similar fruits unsterilized as checks. The results were the following:

Experiment No. 1.—Sterilized: percentage of fruits infested_____ 0
Check—percentage of fruits infested_____ 73
Experiment No. 2.—Sterilized: percentage of fruits infested_____ 0
Check—percentage of fruits infested_____ 67
Experiment No. 3.—Sterilized: percentage of fruits infested_____ 0
Check—percentage of fruits infested_____ 87
Experiment No. 4.—Sterilized: percentage of fruits infested_____ 0
Check—percentage of fruits infested_____ 70
Experiment No. 5.—Sterilized: percentage of fruits infested_____ 0
Check—percentage of fruits infested_____ 84
Experiment No. 6.—Sterilized: percentage of fruits infested_____ 0
Check—percentage of fruits infested_____ 74

Experiment No. 7.—Sterilized: percentage of fruits infested_____ 0
Check—percentage of fruits infested_____ 58
Experiment No. 8.—Sterilized: percentage of fruits infested_____ 0
Check—percentage of fruits infested_____ 72
Experiment No. 9.—Sterilized: percentage of fruits infested_____ 0
Check—percentage of fruits infested_____ 57
Experiment No. 10.—Sterilized: percentage of fruits infested_____ 0
Check—percentage of fruits infested_____ 86
Experiment No. 11.—Sterilized: percentage of fruits infested_____ 0
Check—percentage of fruits infested_____ 61
Experiment No. 12.—Sterilized: percentage of fruits infested_____ 0
Check—percentage of fruits infested_____ 45
Experiment No. 13.—Sterilized: percentage of fruits infested_____ 0
Check—percentage of fruits infested_____ 52
Experiment No. 14.—Sterilized: percentage of fruits infested_____ 0
Check—percentage of fruits infested_____ 80
Experiment No. 15.—Sterilized: percentage of fruits infested_____ 0
Check—percentage of fruits infested_____ 71
Experiment No. 16.—Sterilized: percentage of fruits infested_____ 0
Check—percentage of fruits infested_____ 65
Experiment No. 17.—Sterilized: percentage of fruits infested_____ 0
Check—percentage of fruits infested_____ 42
Experiment No. 18.—Sterilized: percentage of fruits infested_____ 0
Check—percentage of fruits infested_____ 78
Experiment No. 19.—Sterilized: percentage of fruits infested_____ 0
Check—percentage of fruits infested_____ 70
Experiment No. 20.—Sterilized: percentage of fruits infested_____ 0
Check—percentage of fruits infested_____ 63

Total sterilized fruits, 2,000, total infested, 0.
Total check fruits, 2,000, total infested, 1,357, percentage infested, 67.8.

No pupae were recovered from the sand on which the sterilized fruits stood. From the sand on which the check fruits stood 4,032 pupae were recovered and 3,200 adults emerged from them. The adults were identified by the writer as the species breeding in mangoes.

The sterilized fruits which were not infested, ripened normally and showed no difference in flavor, texture, or keeping qualities from the non-infested check fruits.

Experiments with Guavas

Ten lots of 50-half ripe apparently infested guava fruits each were sterilized keeping ten lots of 50 similar fruits unsterilized as checks.

Experiment No. 1.—Sterilized: number of fruits infested_____ 0
Check—number of fruits infested_____ 43
Experiment No. 2.—Sterilized: number of fruits infested_____ 0
Check—number of fruits infested_____ 32

Experiment No. 3.—Sterilized: number of fruits infested..... 0
Check—number of fruits infested..... 36
Experiment No. 4.—Sterilized: number of fruits infested..... 0
Check—number of fruits infested..... 41
Experiment No. 5.—Sterilized: number of fruits infested..... 0
Check—number of fruits infested..... 35
Experiment No. 6.—Sterilized: number of fruits infested..... 0
Check—number of fruits infested..... 46
Experiment No. 7.—Sterilized: number of fruits infested..... 0
Check—number of fruits infested..... 30
Experiment No. 8.—Sterilized: number of fruits infested..... 0
Check—number of fruits infested..... 40
Experiment No. 9.—Sterilized: number of fruits infested..... 0
Check—number of fruits infested..... 26
Experiment No. 10.—Sterilized: number of fruits infested..... 0
Check—number of fruits infested..... 32
Total sterilized, 500 fruits, total infested, 0.
Total checks, 500 fruits, total infested, 361, percentage infested, 72.2.

No new emergence holes were observed in the guava fruits that were sterilized, nor were maggots found in them. No pupae were recovered from the sand on which the fruit stood. In the check fruits new emergence holes and living maggots were observed and 983 pupae were recovered from the sand on which the fruit stood. From the pupae, 840 adults emerged which were identified by the writer as the species that breeds in guavas.

Sterilization of Pupae From Mangoes

Although the fruit fly infesting mangoes in Puerto Rico does not pupate inside the fruit, nevertheless, to determine whether the pupae would be affected in the same manner as the eggs and the maggots by the sterilization treatment, 500 pupae were placed among mango fruits and sterilized at 43° C for 8 hours.

The sterilized pupae as well as 500 more similar pupae which served as checks were kept in moist sand. No flies emerged from the sterilized pupae. From the checks, 423 flies emerged. They were identified by the writer as the species that breeds in mangoes.

Sterilization of Pupae From Guavas

The species breeding in guavas, is the one that occasionally infests citrus. In guavas it does not pupate inside the fruit but in citrus it occasionally does. One hundred pupae of this species were sterilized having been placed among guava fruits. No flies emerged from them. From 100 similar pupae kept as check, 76

adults emerged which were identified by the writer as the species that breeds in guavas.

Conclusions

The tests prove that an exposure of eight hours at 43° C in an appropriate sterilizer is sufficient to kill the eggs, maggots and pupae of the fruit flies that infest mangoes and guavas in Puerto Rico. The treatment does not alter the flavor, texture or keeping qualities of the fruit that is not infested with fruit flies.

Shortening the Sterilization Period For Mangoes

To determine whether a period shorter than eight hours at 43° C might be sufficient, five experiments were conducted using 50 half-ripe "white mango" fruits in each experiment and keeping an equal number unsterilized as checks. A total of 250 fruits were sterilized for four hours and an equal number kept as checks. None of the sterilized fruits showed new exit holes or was found to contain living fruit fly maggots. No pupae were recovered from the sand on which the sterilized fruit stood. Out of the checks, 112 fruits showed new exit holes and contained living fruit fly maggots. From the sand under the checks 305 pupae were recovered from which 235 adults emerged. They were identified by the writer as the species that breeds in mangoes.

The tests show that four hours at 43° C are sufficient to kill the fruit fly eggs or maggots present in mango fruits. Thus an eight-hour period provides a very large margin of safety.

Shortening the Sterilization Period For Guavas

Five more experiments similar to the previous were conducted using 50 half-ripe guavas in each and keeping 250 similar fruits unsterilized as checks. None of the fruits treated for four hours showed fresh exit holes or was found to contain living fruit fly maggots. No pupae were recovered from the sand underneath them. Out of the 250 fruits kept as checks 186 showed new exit holes and contained living fruit fly larvae. From the sand under the checks 460 pupae were recovered from which 340 adults emerged. The adults were identified as the species breeding in guavas.

The tests show that with the species breeding in guavas, the sterilization period can also be shortened without reducing its effectiveness and that therefore the eight-hour period offers a large margin of safety.

Sterilization of Mangoes Wrapped in Paper

To determine whether eight hours at 43° C would be sufficient to kill the eggs or maggots of the fruit fly in mangoes that have been wrapped in paper similar to that used for packing citrus fruits, 50 fruits were used in each of five experiments, keeping 50 similar fruits as checks. None of 250 sterilized fruits showed new exit holes or was found to contain living fruit fly maggots. No pupae were recovered from the sand under them. Out of 250 check fruits 92 showed new exit holes and were found to contain fruit fly maggots. From the sand under them 340 pupae were recovered and 194 adults emerged. The adults were identified by the writer as the species that breeds in mangoes.

The results of the experiments show that mangoes can be successfully sterilized after having been wrapped in paper.

Sterilization of Mangoes Packed in Crates

To determine whether 8 hours at 43°C would be sufficient to kill the eggs and maggots of the fruit fly in mangoes packed in crates, five experiments were conducted. Thirty half-ripe "white mango" fruits were used in each experiment keeping 30 similar fruits for checks. Three kinds of crates were used: (1) all sides ½ inch white pine wood, (2) bottom and two sides ½ inch white pine wood other two sides and top wire netting and (3) all sides ½ inch white pine wood with an opening 2 inches square covered with wire netting on each side except top and bottom. Ten mangoes were placed in each crate and sterilized. The experiment was repeated five times. The total number of fruits sterilized was 150. No new emergence holes, nor living fruit fly maggots were found in them and no pupae in the sand underneath them. Out of 150 check fruits, 93 showed new emergence holes or were found to contain living fruit fly maggots and 240 pupae were recovered from the sand under the fruits. From the pupae 146 adults emerged which were identified by the writer as the species that breeds in mangoes.

These experiments show that mangoes packed in crates similar to those used commercially for fruits and vegetables can be sterilized to kill eggs or maggots of the fruit fly as readily as if the mangoes were placed on trays.

Sterilization of Mangoes Wrapped and Crated

To determine whether mangoes wrapped in paper, packed in crates and sterilized for 8 hours at 43°C would have the eggs and

maggots of the fruit fly killed, 5 experiments were conducted which were a repetition of the previous one excepting that the fruits before crating were wrapped in tissue paper sheets similar to those used for citrus. No new emergence holes nor fruit fly maggots were found in any of the 150 sterilized fruits and no pupae in the sand underneath them, but of 150 fruits kept as checks, 103 showed new emergence holes or were found to contain living fruit fly maggots and 327 pupae were recovered from the sand underneath them.

These experiments show that mangoes wrapped in paper and packed in crates similar to those used for other fruits and vegetables could be sterilized successfully.

Summary

1. Mango fruits infested with fruit fly maggots have no commercial value and could not profitably be shipped to the United States.

2. Sterilization at a temperature of 43°C for eight hours in a circulating atmosphere saturated with moisture kills the eggs, maggots and pupae of the fruit flies that infest mangoes and guavas in Puerto Rico without unfavorably affecting the flavor, appearance or keeping qualities of the fruit if it is afterwards placed in refrigeration.

3. Sterilization does not render marketable mangoes which have already been injured by fruit fly maggots. Such fruit shows the injury and the dead maggots on being cut open, and decays rapidly.

4. Sterilized mangoes from Puerto Rico could safely be allowed into the United States under permit because (1st) only varieties that are not infested would be shipped and (2nd) the fruits would in addition have been submitted to a treatment which renders them free from the pest if they contained it.

5. The varieties that could be sterilized are the Mayagüez mango, the Girón, the Colombo kidney and some of the better ones from Vieques.

6. Since the period of sterilization of eight hours can be reduced to four hours without rendering the treatment less effective in destroying the insect, the margin of safety is very large.

7. Mangoes can be rendered free from fruit fly infestation by sterilization for 8 hours at 43°C piled on trays, crated or wrapped in paper and crated.

Literature Cited

1. **Greene, C. T.** A revision of the genus *Anastrepha* based on a study of the wings and the length of the ovipositor sheath (Diptera: Trypetidae). Proc. Ent. Soc. Wash. **36**(6): 127–179, Illus. July 9, 1934.
2. **Sein, Francisco Jr.** *Anastrepha* (Trypetidae, Diptera) fruit flies in Puerto Rico. Jour. Dept. Agric. Puerto Rico, **17**(3): 83–196, Illus. Nov. 14, 1933.
3. **Mackie, D. B.** Heat treatment of California fruits from the standpoint of compatibility of the Florida process. Monthly Bul. Dept. Agr. Calif. **20**(3): 211–218, Illus. 1931.

THE FIRST RECORDS OF THE MOSAIC DISEASE OF SUGAR-CANE IN PUERTO RICO

By GEORGE N. WOLCOTT, *Entomologist.*

The purpose of the present note is to place on record the first observations on the occurrence of mosaic disease of sugar-cane in Puerto Rico, and to supplement the statement of Mr. J. A. Stevenson, Plant Pathologist of the Insular Experiment Station at the time of its discovery here, as to its early distribution. In his final, and presumably definitive, paper on "The Mottling or Yellow Stripe Disease of Sugar-Cane" (Jour. Dept. Agr. P. R., Vol. 3, No. 3, July, 1919), he writes:

> "At the end of the first season's studies as noted in the 1915–16 report, it had attacked the cane in the region bordered by Aguadilla to the west and a line from Utuado to Arecibo along the valley of the Arecibo River on the east. Lack of time did not permit the working out of more exact boundaries, particularly along the south and west."

During the grinding seasons of 1914–15 and 1915–16, the writer was devoting most of his time to making studies on the status of insects of sugar-cane in Puerto Rico, giving special attention to infestations of *Diatraea saccharælis* F., in mature stalks as observed when they were being harvested. Counts of infestations by the moth-borer in units of a hundred stalks were made, either in fields where the cane was being cut or at loading stations near-by, in all sections of the Island. On the status card, not only were spaces available for recording observations on what presumably would be all pertinent factors affecting the abundance or scarcity of the moth-borer, and on the number noted of the other principal insects attacking stalks, but also "Phys. Dead-heart", "Root Disease" and "Other Injury". The only purpose of the entomologist in making observations on diseases was as they might affect the insects, and whenever diseased material was sufficiently abundant or interesting to justify bringing it back to Río Piedras, it was naturally turned over to the Plant Pathologist for determination.

On March 26, 1915, at Añasco, Tablon Pablo, concerning the field southwest of the long railroad bridge, (Status No. 148), containing mixed Rayada cane in first ratoon, the following was written:

"The cane appeared to be dried up: small and shrunken between the joints. The lower part of the field was overflowed by the river and the cane was covered with silt. It appeared to be the worst cane."

On the same day, cane at the loading station at Añasco, coming from Colonia Pacha in the hills, was noted to be "suffering from a stem disease like a leaf spot." Material was brought in to Mr. Stevenson, who examined it carefully at the time and stated that the lesions bore no traces of fungi or bactria, and that he had no idea of what might be the cause of the disease and would not even venture to make a tentative guess, without seeing young material, in an earlier stage of the disease. Travel was not so simple at that time, and as the entomologist had made a sufficient number of insect status examinations at that point for his purposes, the locality was not again revisited for obtaining the required material. Yet the written description of the cane, and his remembrance of the appearance of the lesions indicate that, without a doubt, it was severely cankered with mosaic disease, and definitely establishes a record for Añasco at least a year previous to any given by Stevenson, besides indicating by the condition of the cane that the disease must have been present here for at least two years previously, and possibly several more.

The disease was also present considerably to the south of Añasco, for when accompanied by Mr. R. H. Van Zwalumenburg on April 6, 1915, the status No. 154 at Colonia Carlo Bravo, between hills and the railroad by the road crossing to Mayagüez, bears the notation: "Leaf-spot stem disease."

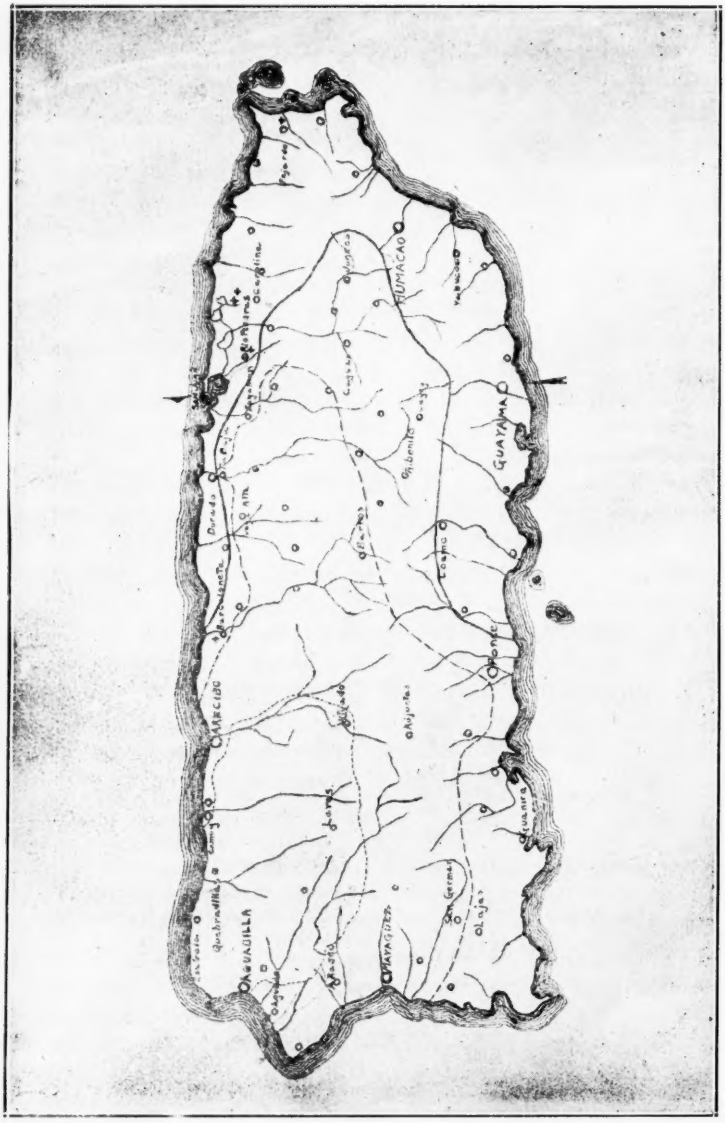
The following year in April, 1916, status observations at Quebradillas, Camuy and Aguadilla record:

"The drying-out disease abundant—elongate lesions between the joints."
"A little of the drying-out disease." (three records)
"Drying-out disease abundant."
"It was cane in a field near to this that suffered so severely from drying-up disease that two Centrals refused it. The buyer said he thought it was due to lack of air drainage. Rainfall had been ample." (Camuy.)

These records only confirm Stevenson's statements as to where he first found the disease. Other records made by the entomologist in this year, however, greatly extend the known range of the disease. Status Nos. 166–7–8, all at Filial Amor, record:

"Drying-out disease abundant—bad." (at Colonia Rosario.)
"Drying-out disease abundant." (at Colonia Francisco Román.)
"More of drying-out disease." (at Colonia Emisa.)

Filial Amor is the railroad junction northwest of San Germán, thus these records for 1916 indicate that even this early the disease must have existed far south of the area mapped by Stevenson, and that the original focus of infection may not have been in the hills back of Camuy, but possibly much farther south.



Reproduction of Stevenson's Map, Showing Supposed Early Distribution and Known Later Spread of Mosaic Disease of Sugar-Cane in Puerto Rico. (After Stevenson).

ROOT DISEASES OF SUGAR CANE IN PUERTO RICO

PART I.—NORMAL STRUCTURE OF ROOTS

By MELVILLE T. COOK, *Plant Pathologist,*
Agricultural Experiment Station, Río Piedras, Puerto Rico.

Before starting the studies on the diseases of roots of sugar cane, it appears to be desirable to give a brief review of our knowledge of normal healthy roots. This has been done in other publications and there is nothing new in this brief discussion but it appears desirable in order to make comparisons. When a cutting is planted, it produces two kinds o roots. Very small roots develop just above the node which are known by several names, such as primary, adventitious, etc. Much larger roots are formed at the bases of the buds soon after the shoots start to elongate. They are known as secondary or true roots, etc.

The primary or adventitious roots usually die early but under some conditions presist and form dense mossy like growths. A very large percentage of the large or true roots die early but many of them grow to twelve inchs or more in length. Some few of them attain a much greater length, sometimes as much as six feet. They branch to some extent by the formation of small lateral roots. The number of these small lateral roots is extremely variable. We do not know all the factors that may influence the number of small lateral roots but injuries by insect larvae, fungi and other agencies at or near the apex are important. The writer is inclined to believe that the environmental factors, such as character of the soil, water supply, etc., are extremely important. When a root attains a length of about six or eight inches the cortex usually dies except for three or four inches at the tip. Many people in digging the roots of sugar cane believe these roots are dead, but an examination of the axis cylinder will show that they are alive. The absorption is restricted entirely to the small amount of living cortex near the tip. The dead cortex usually contains many fungi and bacteria, usually saprophytic forms.

The structure of the large roots is practically the same as for the roots of most plants. An axis cylinder, a cortex, an epidermis and a root cap. All of which are shown in longitudinal section (figure 1). The tracheary tubes begin to form early and just back of the root tip (figure 1). They originate from several single rows of

cells in the axis cylinder and are arranged so that they form a circle in cross section (figures 1, 2 and 12). They are small at first (figure 7) but increase in size (figures 8 and 9). This relationship to the surrounding cells is shown in figures 5 and 10. They are cells of the axis cylinder which differentiate, enlarge and exert a pressure on the surrounding cells (figures 5 and 10). The density of the protoplasm in these cells is variable (figures 5, 10 and 11). The general appearance of these cells is that of great activity until they have reached their full growth when the contents undergoes degeneration (figure 6). Eventually the cell walls between the cells of a row disappear and the tracheary tube is complete.

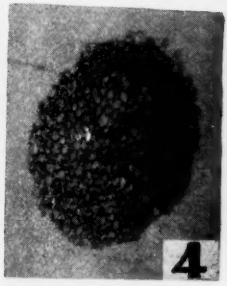
The demarkation between the axis cylinder and the cortex and between the cortex and epidermis appear early and are well defined (figures 12, 13 and 14). The axis cylinder may presist for a long period, as previously stated in this paper, but in other cases the entire root dies very early.

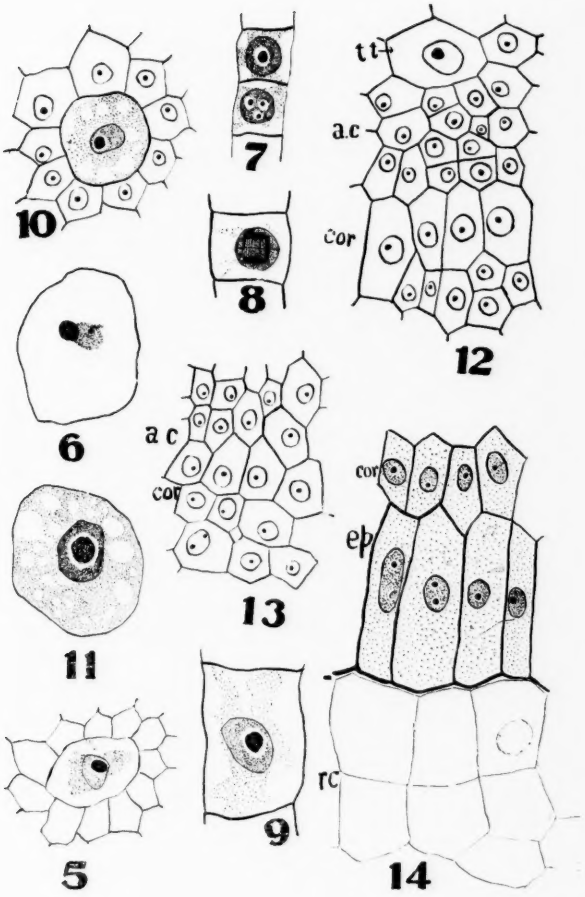
The root cap is the same as in the roots of most plants. It projects beyond the apex of the root. The cells at point of origin are well supplied with protoplasm (figure 4) while those most remote contain little or no protoplasm (figure 1). The root cap extends up the sides of the root tip for a short distance and the demarkation between epidermis and cap is very distinct (figures 1, 2, 3 and 14).

The injuries to the root originate in the active, healthy cortex. Fungi and bacteria may be found in the dead cortex of the old roots but their presence does not indicate that they are injurious. Most of them appear to be saprophytic. Of course some of the organisms found in the dead cortex may cause some injury. The healthy cortex is attacked by fungi, bacteria, nematodes, the larva of insects and possibly other forms of life. The results of the writers studies on these forms of life will be published from time to time.

Studies on *Marasmius sacchari,* which has been referred to so often and from so many different countries as a cause of root diseases, have been made by the writer and the results published in the proceedings of the Fourth Congress of the International Society of Sugar-Cane Technologists held in San Juan, Puerto Rico, March 1932, under the title of "The Parasitism of Marasmius sacchari Wakker" and in The Journal of the Departament of Agriculture of Puerto Rico, Volume XVI, No. 2, pages 213–226, 1932, under the title of "*Marasmius sacchari;* a Parasite of Sugar Cane". The latter is the more complete. It should have been included in this series on Diseases of Roots of Sugar Cane.

PLATE IX.




10
7
8
tt
ac
cor
12
6
ac
cor
13
11
cor
ep
5
9
rc
14

Explanation of Plates

Figure 1.—Longitudinal section of tip of root showing axis cylinder with one row of cells that are to form a tracheary tube; cortex, epidermis and root cap.

Figure 2.—Cross section of tip of root showing axis cylinder and a circle of large cells that are to for tracheary tubes; cortex, epidermis and a small amount of the root cap around the margin.

Figure 3.—Cross section of root tip lower down and a little above apex showing axis cylinder, cortex, epidermis and root cap around the margin.

Figure 4.—Cross section through the root cap.

Figures 5, 10 and 11.—Cross sections of cells that are to form tracheary tubes. Note difference in protoplasmic contents.

Figure 6.—Same in state of disintegration.

Figures 7, 8 and 9.—Longitudinal sections of cells that are to form tracheary tubes.

Figure 12.—Cross section of root tip showing cortex, axis cylinder and one cell of a future tracheary tube.

Figure 13.—Cross section of a root tip showing primitive axis cylinder and cortex.

Figure 14.—Cross section of root tip showing parts of cortex, epidermis and root cap.

a.c.—axis cylinder. Cor-cortex. ep = epidermis. r.c. = root cap. tt = tracheary tube.

The photographs for figures 1 to 4 were made by Dr. Harold T. Cook.

PART II.—A NEW PARASITIC FUNGUS IN THE ROOTS OF SUGAR CANE

This fungus was found on the roots of sugar cane growing in pots, while the writer was studying another disease. The writer did not make a microscopic examination of the roots at the time but put a large number in a killing fluid for later study. Therefore, the disease was not recognized as new until after the roots had been sectioned and stained. Upon the discovery that the fungus was new to science, the writer made a search for additional material in order to study living material but without success. Therefore, these studies are based entirely on material that had been sectioned and stained. However, there was a sufficient amount of material for study.

The material was killed in weak Flemming and most of it was stained with Haematoxylon and iron-alum.

The *symptoms* of the disease visible to the unaided eye are of little or no importance for diagnostic work. They appear as small reddish spots which cannot be distinguished from spots due to other causes. The fungus is restricted to the younger parts of the roots. It is doubtful if it ever attacks a root more than 1½ inch back of the tip.

The *importance* of the disease cannot be determined until a more thorough study can be made. The fungus was found in great abundance in some roots and had completely destroyed the contents of many cells. It was found on four varieties, growing in pots and well supplied with water. They are Uba, M 28, PR–801, POJ–2878, SC–12(4) and BH–10(12). It was most severe on PR–801. It has not been found in the field. The nature of the disease is such that a severe outbreak might prove very destructive, The taxanomic relations of the organism are such as to indicate that it will thrive best in soil well supplied with water.

The *fungus* is found in the epidermal cells and cortex but has not been found in the axis-cylinder. It sometimes occurs in masses which have been estimated to contain between 150 and 200 cells, but it also occurs in single cells throughout the cortex. There is no enlargement of the host cells. The fungus appears as a plasmodial mass which almost completely occupies the cell (Figs. 1–3). In most cases there is a single mass, while in a few others there may be several of these masses. (Figs. 4–6) This mass may be vacuolar or densely granular. (Figs. 1–2). In the young stages the nuclei are

not visible, (Figs. 1–2) but they develop in great numbers with age (Figs. 3–4). In the great majority of cases the cell wall is thin but always distinct (Figs. 1–2) but in some cases it is very thick (Fig. 3). These thick walled cells appear to be resting spores.

The plasmodium produces a germ tube which punctures the cell walls and penetrates one to four cells, possibly more, although four was the largest number observed. (Figs. 5–23). The writer never observed the germination of more than one plasmodium in a cell. There was never more than one tube to a plasmodium and branching was observed in one case only (Fig. 13). Most of the germinating plasmodia were large but two small ones were also observed (Figs. 12 and 14 ×). Bensaude (**2**) reported a plug of dense protoplasm at the tip of the tube. No such plug was observed in any of the writer's preparations. The formation of the tubes and zoospores have a superficial resemblance to those of *Diplophlyctis intestina* as described by Karling (**4**).

The tubes usually grew more or less in a direct line, but there were some exception. In one case the tube grew into the form of a letter U (Fig. 15). In the great mayority of cases the tube grew in the direction of the epidermis. All the exceptions were a considerable distance from the surface. The writer is unable to say whether this indicates some type of tropism or not.

The formation of nuclei and spores was not correlated with the formation of the germ tube. Sometimes the germ tube is without any evidence of nuclei (Figs. 9, 10, 11, 12, 14 and 16) while in others the nuclei are very distinct and numerous (Figs. 7, 8, 13, 15, 17 and 20) and in still others the spores are formed before the tubes have made much progress (Figs. 18 and 19). The tubes penetrated the cell walls in all cases observed except one in which it went through the wall of its own cell and then between the walls of other cells (Fig. 16). A few cases were observed in which the germ tube was unable to penetrate the cell wall but pushed it forward (Fig. 20).

The spores are numerous, more or less spherical and uninuclear with very thin but distinct walls. In some cases the germ tube passes through the epidermis and the spores are emptied into the soil (Figs. 14 and 23) while in other cases they are emptied into another host cell.

The spores that empty into the soil, presumably penetrate the epidermal cells of the same or other roots. Bensaude (**2**) reports a discharge of spores directly into the soil. Actual penetration was not observed but many cases were observed in which the epidermal cells were infected (Figs. 41 to 42). Ciliated stages of the spores

were not observed but many preparations showed amoeboid characters. It is possible that a study of fresh material would have shown ciliated spores.

The behavior of the organism in the host was observed in many preparations. They were amoeboid and frequently united as shown by the number of nuclei (Figs. 30–34 and 41). In some cases they tended to become filamentous (figs. 35 and 37) and many of them penetrated cell walls (Figs. 38 and 39). They did not grow and unite with equal rapidity (Figs. 29–39). Sometimes large plasmodial bodies were found in the same cell with spores (Figs. 29–30).

A careful study of the available material leads the writer to believe that many of the sporangial bodies mature their spores without the formation of a germ tube. The formation of tubes was not observed in any of the thick walled sporangia (Fig. 3) although a careful search was made of the adjoining sections of many of them. Yet these thick-walled sporangial bodies produced an abundance of spores.

The thick-walled spores (or sporangia) were apparently resting. Roughened, thick-walled spores (or sporangia), such as are described and figured by Woronin (**6**) and Bensaude (**2**) were observed (Fig. **48**) but the writer is inclined to believe that this roughness is due to plasmolosis and shrinkage.

The possibility of a second species attracted the attention of the writer. A few sporangia were observed in which the spores were much smaller than the others (Fig. 21, 22) and many cases were observed in which small spores were germinating in the host cells (Fig. 36). These small spored sporangia were very few, but the germination and behavior of the spores appeared to be the same as in the large spored forms. The large and small spored forms were not observed to unite or have any relationship whatever.

Double infection of the large host cells by the large-spored forms was observed in many cases (Figs. 43–46. The writer's opinion is based on the fact that young sporangia and spores in various stages of development were observed in many host cells.

Abnormal sporangia were of frequence occurrence. In these cases the sporangia did not develop the spherical body but developed a worm-like structure which penetrated the walls of the host cells and produced spores in the same manner as the normal, spherical sporangia (Fig. 24–28).

Discussion

The life history of this organism is practically the same as Olpidium, with slight variations, except that the writer has not seen flagella on the spores. It is possible that flagella are present and that they can be seen in living material. Therefore, the writer will place this species tentatively in the genus Olpidium. However, it is very evident that the species is new.

Olpidium sacchari n. sp. Sporangia usually solitary in cells of the host, spherical, multinucleate, germinating by a single tube. Zoospores numerous, uninucleate, amoeboid, uniting to form plasmodia; sporangia 2.66—3.5μμ resting spores spherical and thick walled. Resting spores spherical with thick, smooth walls.

Sporangium saepius in cellulis hospitis solitarium sphericum multinucleatum unico tubo germinante; zoosporae numerosae uninucleatas amoeboideae in plasmodia conjugantes; sparangia 2.66—3.5uu sporae immobiliae sphericae parietibus crassis.

Literature

1. **Barrett, J. T.**. Development and sexuality of some species of Olpidiosis (Cornu) Fisher. Ann. Bot. **26**: 209–238. 1912.

2. **Bensaude, Matilde,** A species of Olpidium parasitic in the roots of tomato tobacco and cabbage. Phytopathology **13**(10): 451–454. 1923.

3. **Griggs, R. F.** Monochytrichum, a new genus of the Chytridiales, its life-history and cytology. Ohio Naturalist **10**(3): 44–54. 1910.

4. **Karling, J. S.** Studies in Chydriales–II. Contribution to the life history and occurrence of Diplophlyctis intestina (Schenk) Schreeter in cells of American Characeae. Amer. Journ. Bot. **15** (3): 204–214. 1928.

5. **Kusano, S.** On the life history and cytology of a new Olpidium with special reference to the copulation of motile isogametes. Journ. Coll. Agri. Tokyo. **4**: 141–199. 1912. (Rev. in Trans. Brit. Myc. Soc. **4**: 291–1913.

6. **Woronin, M.** Plasmodiophora Brassicae, Urheber der Kohlpflanhernie. Jahrb. Wiss. Botanik. **11**: 548–574. 1878.

Explanation of Plates

Figures 1 to 6.—Plasmodia in host cells. In figure 1 the protoplasm is vaculolate; in 2 it is dense and shows first evidence of formation of nuclei; in 3 the nuclei are prominent and the wall thick. Figures 1 and 2 will produce germ tubes but figure 3 will

not. Figures 4, 5 and 6 show two or more sporangia in single host cells. In figure 6 one sporangium is forming a germ tube.

Figures 7, 8 and 9 show germ tubes and nuclei in various stages of formation.

Figures 10 to 14 show sporangia of different sizes and germ tubes. Figures 10 and 13 show two undeveloped sporangia; 11 and 14 show tips of tubes of sporangia not shown in the sections; the sporangia in figures 12 and 14 are very small; the tube in figure 13 is branched. This is unusual. e = epidermal cells.

Figure 15.—The germ tube is curved. This is unusual.

Figure 16.—A germ tube passing between the cells instead of through them. This is unusual.

Figures 17, 18 and 19.—Zoospores in various stages of formation.

Figure 20.—A germ tube has pushed the wall of the host cell forward instead of penetrating it. This is unusual.

Figures 21 and 22.—The zoospores are very small. Note the size as compared with 19.—e = epidermal cells.

Figure 23.—An unusually long tube. e = equals epidermal cells.

Figures 24 to 28.—Abnormal sporangia.

Figure 29.—A host cell containing a large number of zoospores that have not been injected into the soil. Also one large sporangium.

Figures 30 to 34.—Host cells in which the zoospores are uniting to form sporangia.

Figures 35 to 39.—Host cells in which the sporangia are amoeboid. In figures 38 and 39 the sporangia have penetrated the walls of the host cells.

Figure 40.—Two large sporangia in adjacent cells have united and are forming a germ tube.

Figures 41, 42 and 47.—Plasmodia in epidermal cells of the host. e = equals epidermal cells. Apparently the zoospores penetrated from the soil.

Figures 43 to 46.—Cells of the host in which there are sporangia of different ages, probably due to infections at different times.

Figure 48.—An irregular shapped sporangium probably due to plasmolysis.

PLATE XI.

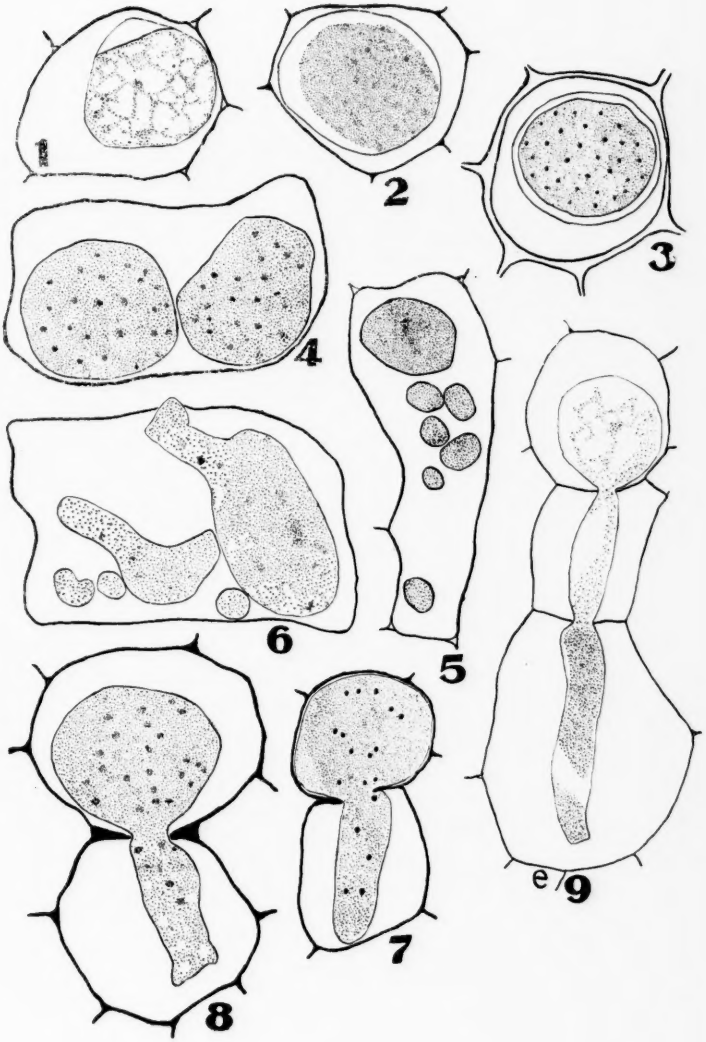


PLATE XII.

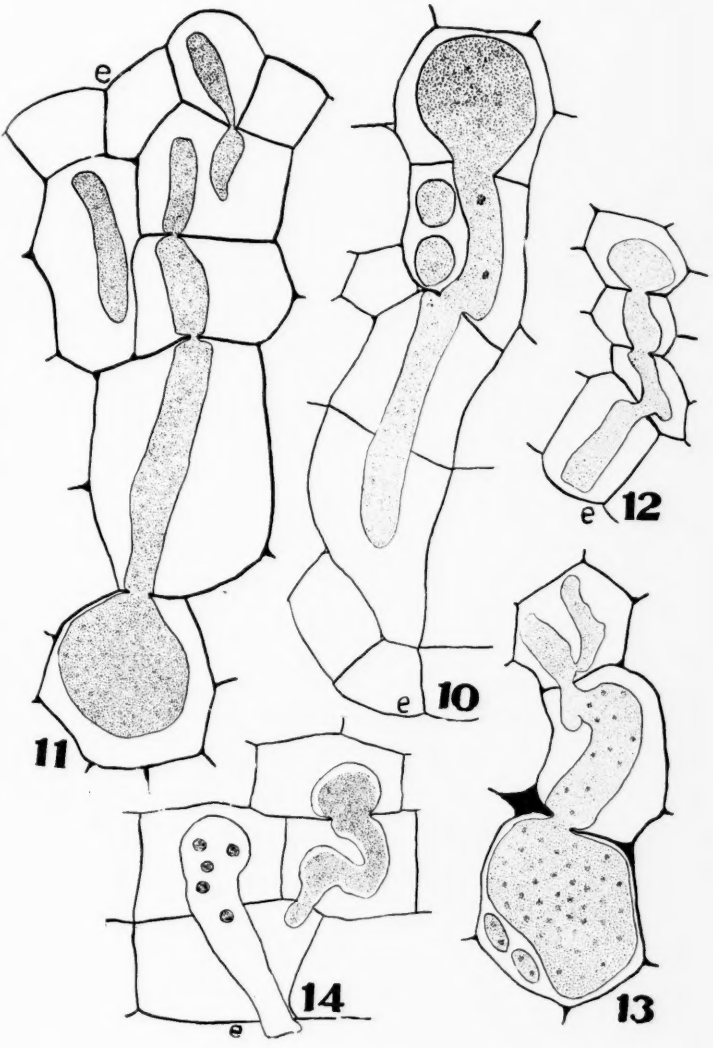


PLATE XIII.

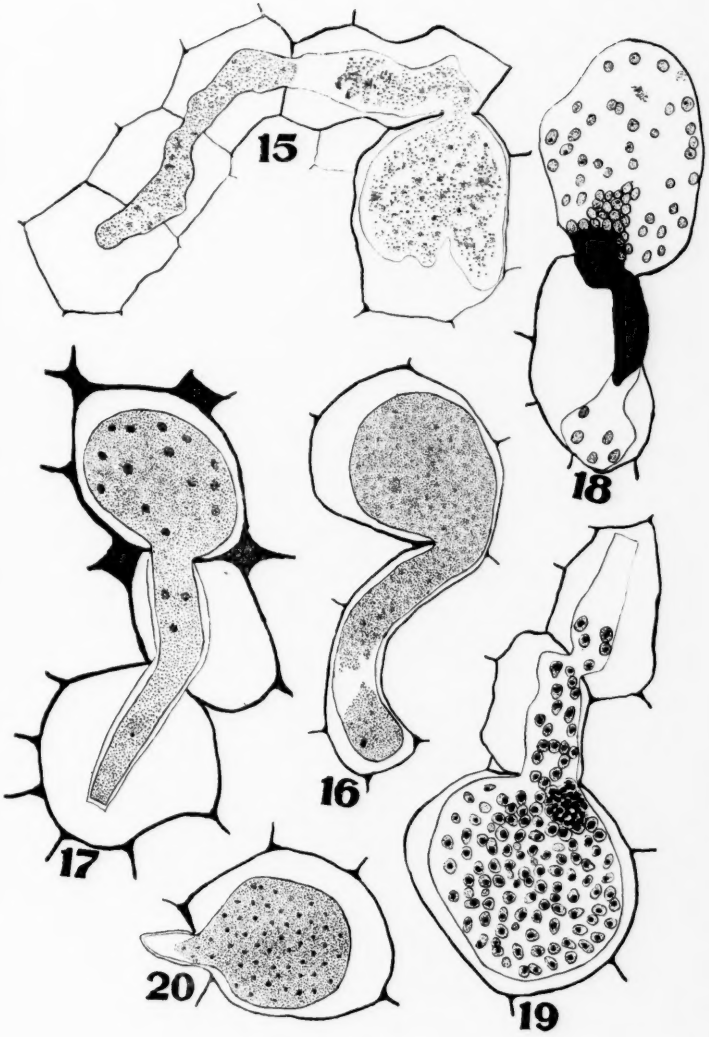


PLATE XIV.

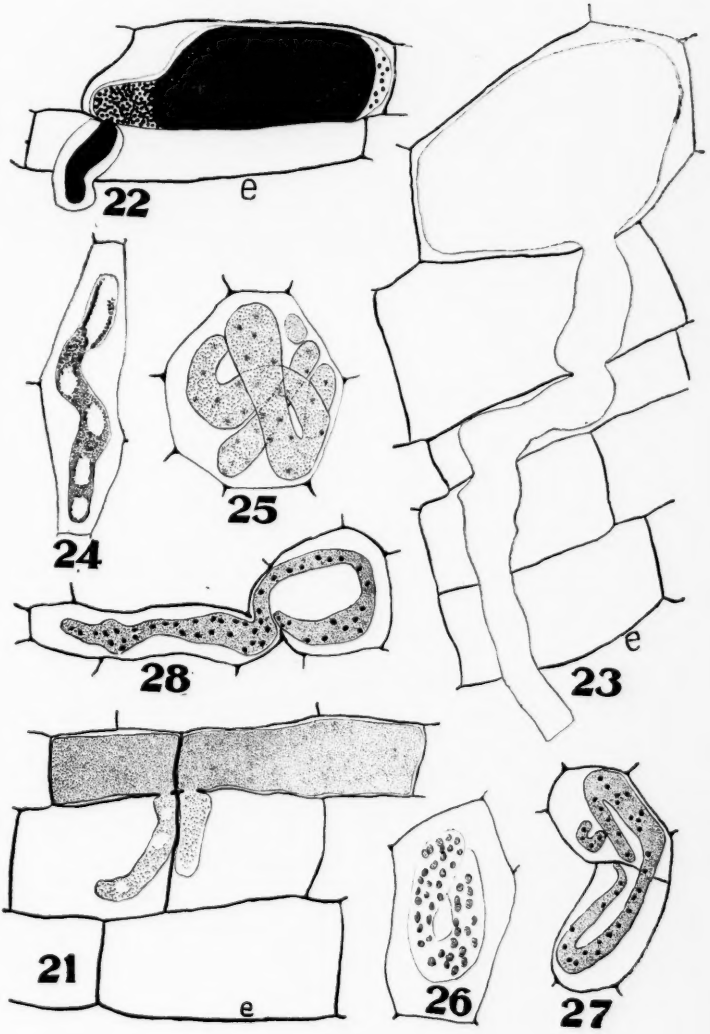


PLATE XV.

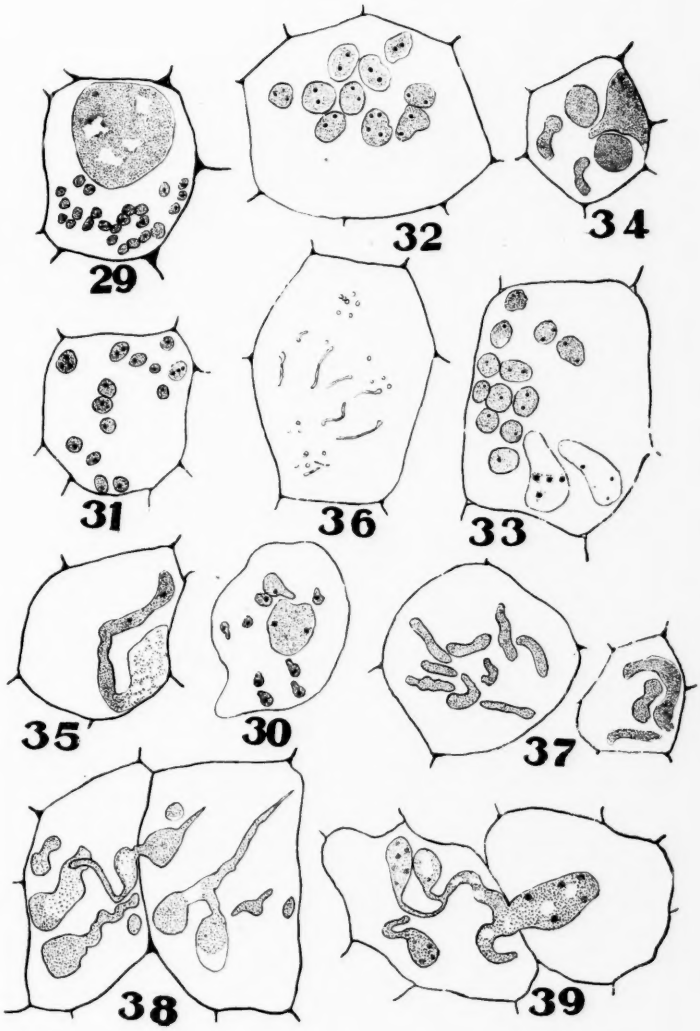
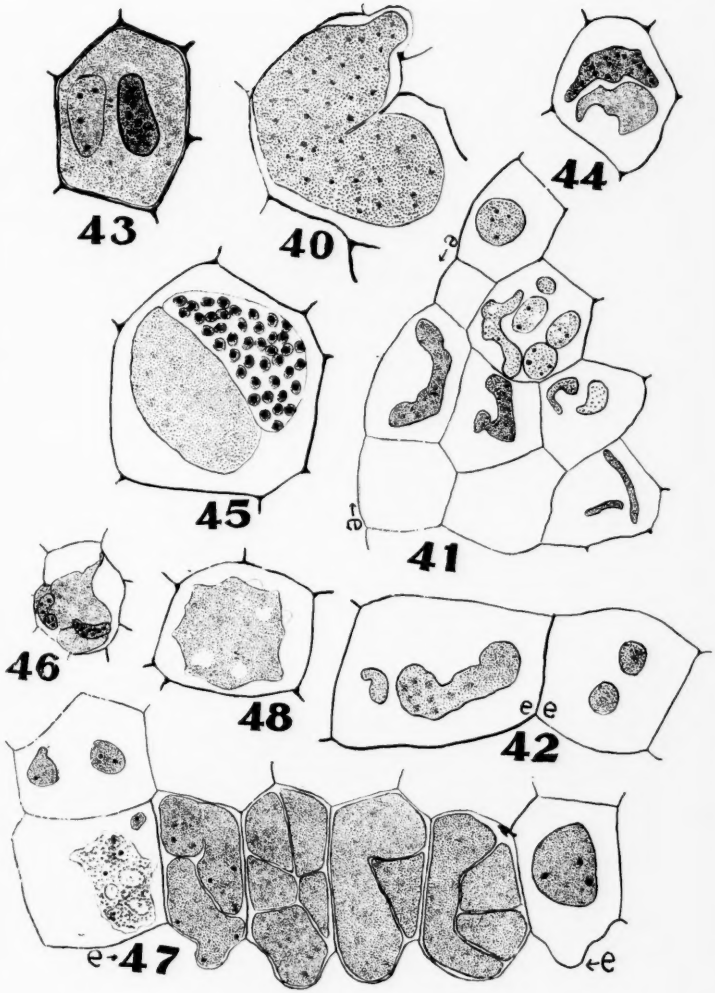


PLATE XVI.



FIRST SUPPLEMENT

TO

PARTIAL BIBLIOGRAPHY OF VIRUS DISEASES OF PLANTS *

By José I. Otero, *Librarian*, and Melville T. Cook, *Plant Pathologist*,
Agricultural Experiment Station, Río Piedras, Puerto Rico.

Introduction

This first supplement of "Partial Bibliography of Virus Diseases of Plants" is presented to the students of virus diseases of plants, hoping that it will make the original more useful.

After the publication of the above-mentioned work, we have been favored by a great number of investigators on virus with advices, corrections and contribution of papers for which we very sincerely thank all of them. We appreciate the many encouraging letters that we have received from many students interested in virus disease investigations.

It is materially impossible to locate and collect all the literature with our limited library facilities. There may be errors in this supplement as well as in the original; therefore, we will appreciate the cooperation of workers who may be interested in succeeding supplements. Please call our attention to errors, omissions, etc., and send us new papers for citation.

Address correspondence to the Junior author.

This supplement contains:

1. First Supplement to Partial bibliography of virus diseases of plants.
2. Index to this Supplement.
3. 1st Appendix.—Index to Partial bibliography of virus diseases of plants.
4. 2nd Appendix.—Authors' Index of Partial bibliography of virus diseases of plants.
5. 3rd Appendix.—Errata to Partial bibliography of virus diseases of plants.

* Journ. Agric. Univ. Puerto Rico 18(1–2): 1–410, 1934.

Afzal, Husain M.

A note on a growth abnormality of Punjab—American cottons. Indian Journ. Agric. Sci. **3**(5): 933, 1933.

Report on a disorder of cotton which was first noticed in 1930 in the Canal Colonies of the Punjab and which reappeared with great severity in 1932. The varieties affected are American. The author states that the disease has great similarity to "stenosis" reported by O. F. Cook from Haitian cotton. He attributes the disease to a virus and gives description.

Ainsworth, G. C.

Virus disease investigations. Expt. & Res. Stat. Nursery & Mark. Gard. Indus. Devel. Soc. Cheshunt. Ann. Rpt. **18**: 39–45, 1933.

----------, **Berkeley, G[raven] H[ugh] & Caldwell, John**

A comparison of English and Canadian tomato virus diseases. Ann. Appl. Biol. **21**(4): 566–580. 1934.

After a comparative study of Canadian and English tomato diseases it was found that the commoner tomato virus diseases occur in both countries. Tomato streak caused by the same single virus occurs in both countries, but streak due to a mixed virus infection appears to be more common in Canada. Several virus diseases with their causal viruses are described.

A comparison of certain English and Canadian potato viruses found infecting tomatoes. Ann. Appl. Biol. **21**(4): 581–587, 1934.

The author compared potato viruses isolated from Canadian tomato material. The different viruses which corresponded to the mild and virulent forms of the latent or "healthy potato" virus were all considered to be strains of potato virus which differed primarily in virulence. It was found that immunity against a virulent strain of the virus was conferred on tomato plants by inoculation with a mild strain.

Virus disease investigations. Cheshunt Exper. & Ros. Stat. Herf. 19 Ann. Rpt. **1933**: 54–64, 1934.

This rather extensive report includes interesting data on several virus diseases of economic importance as well as on uncultivated host plants.

Mosaic disease of the cucumber. Ann. Appl. Biol. **22**(1): 55–67. 1935.

Cucumber virus No. 1 is closely allied with Johnson's cucumber virus No. 1. It is mottle yellow. No. 3 is a green mottle mosaic. No. 4 is associated with No. 3. The author describes these viruses and the symptoms produced by them.

Allen, H. F.

Causes and effect of dahlia mosaic. Midwest Dahlia News **2**(3) : 6–8, 1934.

Altson, R[alph] A[bbey]

Report on a suspected outbreak of infectious mosaic disease among certain canes in the Colony. I & II. Journ. Bd. Agric. British Guiana. **18**(3) : 216–225, 1925.

The author reports in Part I an infection of mosaic (infectious chlorosis) in a plantation of sugar cane of the variety BH–10(12) brought from Barbados 1920 which, as preventing measure, he recommended to be eradicated. The infection was as high as 80 per cent prior to its eradication. In one instance the infection was observed on Ba. 6032. The author states that in no instance has there been any indication of spread to neighboring varieties of canes or to weed grasses. He gives a brief general account as to the nature of the disease and recommendations to prevent its spread. In Part II he reports that the above disease is not infectious and that it is not mosaic, giving his evidence and conclusions.

The spread of cane mosaic in the West Indies. Int. Sugar Journ. **27**(318) : 293, 1925.

Review of a visit by the author to Jamaica, Costa Rica and Trinidad. Mosaic of sugar cane is the most serious problem. In some cases 100 per cent of the canes are infected. He gives yield and cost data. He reports also that up to this date sugar-cane mosaic disease has not appeared in British Guiana.

Ando, H.

(On dwarf disease of rice plant.) Journ. Jap. Agr. Soc. No. **347** : 1–3, 1910.

Anonymous.

(Dwarfed rice plant, its causes and control measures). Japanese Dept. Education. Official Gazette No. **2192** : 231. 1890.

(Results of Agricultural Experiments). Nara Agric. Expt. Sta. Ann. Rpt. **3** : 51–61, 1897.

Experiments with dwarf disease of rice plant.

(Experiments with dwarf disease of rice plant. Results of Agricultural Expriments). Hyogo Agric. Expt. Sta. Rpt. **1** : 68–82, 1895; Rpt. **3** : 16–18, 1896; Rpt. **5** : 21–22, 1897; Rpt. **7** : 139–141, 1898; Rpt. **9** : 121–123, 1899.

(Experiments with dwarf disease of rice plant). Hiroshima Agric. Expt. Sta. Bull. **7**: 21–39, 1903.

(Experiments with the leafhopper in Results of Experiments with insect pests.) Shiga Agri. Expt. Sta. Reports **1**: 111-169, 1899; **2**: 1–26, 1900; **3**: 25–55, 1901; **4**: 19–65, 1902; **5**: 1–37, & (1)—(36), 1904; **6**: 1–43, 1906; **7** & **8**: 1–43 & (1)—(50), 1908.

(Experiments with dwarf disease. Results of Agricultural Experiments). Okayama Agric. Expt. Sta. Rpt. **3**: 62–70, 1902; **5**: 57–64, 1903; **7**: 51–**53**, 1904; **11**: 131–138, 1906; **13**: 169–171, 1907; **15**: 109–111, 1908; **19**: 69, 1909.

(On dwarf disease of rice plant). Kyoto Agric. Expt. Sta. Sp. Bull. **11**, 12 p., 1910. (Rice of the Prefecture of Okayama Prefecture. 300 p., 1910.

Rice disease on p. 189–193.

(Experiments with dwarf disease of rice plant.) Shiga Agric. Expt. Sta. Spec. Bull. p. 56–65, 1906. No. **2**: 35–51, 1910.

(Dwarf disease of rice plant). Miyazaki Agric. Expt. Sta. Sp. Bull. **5**, 26 p., 1915.

Miyazaki Agric. Expt. Sta. Ann. Rpts. **1913**: 32, 1914; **1914**: 29–30, 1915.

Shiga Agricultural Experiment Station. Ann. Rpts. **1907**: 55–59; 1908; **1908**: 52–56, 1909; **1909**: 52–54, 1910; **1910**: 54–56, 1911; **1911**: 58–60, 1912; **1912**: 75–78, 1913; **1913**: 128–130, 1914; **1914**: 29–30, 1915; **1915**: 167–168, 1916.

Notes on dwarf disease of rice plant.

Kagoshima Agric. Expt. Sta. Ann. Rpts. **1912**: 46, 1913; **1913**: 48, 1914; **1914**: 39–43, 1915; **1915**: 94, 1916.

Nagano Agric. Expt. Sta. Ann. Rpt. **1915**: 130–134, 1916.

"Matizado" da canna en Porto Rico (Sugar-cane mosaic in Porto Rico). Biol. Min. Agric. Ind. Com. Brasil **8**(11): **91**, 1919.

Yamanashi Agric. Expt. Sta. Ann. Rpts. **1916**: 88–91, 1917; **1917**: 87–89, 146–147, 1918; **1918**: 48, 69, 1919; **1919**: 39, 1920.

The degeneration of the potato. Gard. Chron. **76**: 1978, 1924.

Popular.

(Imperial Agric. Expt. Sta. Ann. Rpts. **1906**: 53–54; **1909**: 59; **1912**: 29, 1914; **1913–1915**: 40–41, 1917; **1916**: 28; 1918; **1922**: 68–70, 1924; **1923**: 33, 1925).

Historico do mosaico da canna. (History of cane mosaic). Bol. Agric. Bahia, Brasil **1926**(4–9): 25, 1926.

O mosaico da canna de assucar (Sugar-cane mosaic). Circ. Inst. Biol. Def. Brasil, 1926.

Bureau of Sugar Experiment Station. Fiji Disease. Queensland Agric. Journ. **26**(4): 280–281, 1926.

O mosaico da canna (Cane mosaic). Bol. Agric. Zoot. Vet., Minas Geraes, Brasil, **1**(6): 121, 1928.

(A survey on the distribution of plant diseases and insect pests.) Japanese Dept. Agric. & For. Bur. Agric. **2**, 341 p., 1929.

Notes on dwarf disease of rice on page 16–19.

Department of Botany and Plant Pathology. Oregon Agric. Expt. Sta. Director's Bienn. Rpt. **1926–28**: 97–101, 1930.

This report includes notes on several virus diseases.

O mosaico da canna (Sugar-cane mosaic). O Campo, Brasil **1**(1): 135, 1930.

O mosaico da canna (Sugar-cane mosaic). Mem. Inst. Oswaldo Cruz, Brasil, 1930.

O mosaico nos cannavieaes de Pernambuco (Mosaic in the fields of Pernambuco). Rural Brasil **3**(12) : 417, 1930.

O mosaico da canna de assucar (Sugar-cane mosaic). Fazenda Fluminense, Brasil **1**(6) : 18, 1930.

Mosaic and allied diseases of the potato. Ministr. Agric. & Fish. London Adv. Leaflet. **139,** 4 p., 1932.

Insect transmission of spike-disease. Nature **132**: 592–593, 1933.

Contribution á la connaissance de la maladie de l'enroulement des feuilles de la pomme de terre. (Contribution to the knowldge of the leaf roll disease of potato). Progr. Agric. & Vitic. (Montpellier) **100**: 507–509, 1933.

Results from imported canes. South African Sugar Journ. **17**(11) : 573–575, 577, 1933

A practice against recognized scientific procedure has been introduced on a private farm in South Africa. Corn was planted between sugar-cane rows, and it was found that streak infection was reduced. It is claimed that the vector of streak *Cicadulina mbila* prefers to feed on corn rather than on sugar cane.

Spotted wilt of tomatoes. Gard. Chron. **93**: 327, 1933.

Forty-six Annual Report of the Kentucky Agricultural Experiment Station for the year 1933. Part I, 69 p., 1934.

This report contains the description of an experiment on tobacco mosaic (green and yellow) transmission by trash left in the soil.

Mayagüez 28. Puerto Rico Agric. Expt. Sta. Ann. Rept. **1933**: 7–10, 1934.

This variety proved to be highly resistant to mosaic. In a survey of several fields, very seldom more than 3 or 4 per cent infection was found on Mayagüez 28 in plots adjoining heavily infested fields.

Legislative and administrative measures. Madagascar and Dependencies. Internt. Bull. Plant Proct. **8**(3) : 56, 1934.

Comoso Islands and Madagascar, have been declared infected by sugar-cane mosaic and importation of canes from them prohibited.

Experiment Station notes. South African Sugar Journ. **18**(1) : 11–13, 1934.

Brief notes describing the disease and a warning to farmers in varietal susceptibility.

Tobacco growers. Important notice. The "Kromnek" disease of tobacco. Rhodesia Agric. Journ. **21**(1) : 9–10, 1934.

Brief notes describing the disease and a warning to farmers in order to prevent an epidemic.

Archibald, E. S.

Report of the acting Dominion botanist. Dom. Expt. Farm. Ann. Rpt. **1920** : 58–64, 1921.

Notes on bean mosaic on page 62.

Artschwager, Ernst F[riedrich]

Anatomy of the potato plant, with special reference to the ontogeny of the vascular system. Journ. Agric. Res. **14**(6) : 221–252, 1918.

The aim of this study was to make more accurate the knowledge regarding the normal structure of the potato plant. The present work was undertaken in order to make possible a more rapid progress in the study of several important potato diseases such as leaf roll.

Ashby, S[ydney] F[rancis]

Transmission of two diseases caused by infective viruses. Trop. Agric. (Trinidad) **3** : (5)98, 1926.

In this paper the author makes a brief review of the work done on bunchy top of banana occurring in Australia and curl or rosette of peanuts in Tanganyika Territory.

Atanasoff, D[imitr]

(A new virus disease). Yearbook Univ. Sofía, Fac. Agric. **11** : 49–70, 1932.

(Bitter pit of pome fruits is a virus disease). I Contribution. Univ. Sofía. Yearbook. Fac. Agric. **13** : 1–8, 1934.

The author states that an extensive survey of apple orchards and nurseries made during the summer of 1933 showed that bitter pit of apple, pear and quince is a widely spread malady in Bulgaria. Based on observations and graft experiments the author believes himself justified in concluding that bitter pit of pome fruits is caused by a virus or a group of viruses.

Old and new virus diseases of trees and shrubs. Phytopath. Zeitschr. **8**: 197–223, 1935.

Virus disease of citrus. Yearbook Univ. Sofía. Fac. Agri. 1934–35, **13**: 1–42. 1935.

This paper is a discussion of an infectious chlorosis which the author believes to be the same as "mal secco" and blight, or withertip (not same as *Gloeosporium limitticolum*), crinkly leaf, spot mosaic, zonate chlorosis, or ring blotch. The author believes all these diseases to be due to viruses. He also suspects that Reichert's little leaf, some of Shamel's bud selections, leprosis, corticosis, brown spot, peteca, endoxerosis and membranosis may be due to viruses.

Mosaic disease of drupaceous fruit trees. Yearbook Univ. Soíía, Fac. Agric., 1934–35, **13**: 9–42, 1935.

This paper is a discussion of chlorosis, variegations, mosaics and similar symptoms on these fruits which have been reported from various parts of the world but rarely attributed to viruses. The author reports cross-inoculation experiments and says: "The mosaic disease of the various stone fruits is intertransmissible and can easily be communicated from diseased to healthy trees by budding. Under natural conditions it is spread by the plum aphid (*Anuraphis padi*)".

Austin, M. D., & Martin, H.

The incorporation of contact insecticides with protective fungicides. Potato field trials 1930–1932. J. S. E. Agric. Coll. Wye. No. **32**, p. 49–58, 1922.

Report of trials in which nicotine or pyrethrum was incorporated in the blight spray with the purpose of checking aphids on potatoes. The aphids are usually carriers of virus diseases. The results were successful.

Badami, V. K., & Venkata Rao, M. G.

A preliminary report on the varieties of *Santalum album* in Mysore. Mysore Sandal Spike Investigations Committee. Bull. **1**, 1930.

Bailey, M. A.

Leaf curl disease of cotton in the Sudan. Empire Cotton Growing Rev. **11**(4) : 280–288, 1934.

Data in regard to spread and varietal resistance of leaf curl disease of cotton.

Baker, R. E. D.

Maize stripe disease. Trop. Agric. (Trinidad). **10**(8) : 221, 1933.

Record of a disease of sorghum which resembles that on corn. It was experimentally transmitted by the leafhopper *Peregrinus maydis* from maize to sorghum with the production of symptoms resembling those on sorghum in the field. It was concluded that the two diseases are identical.

———————

Stripe disease of maize. Trop. Agri. (Trinidad). **10**(12) : 352, 1933.

A brief record of experiments which demonstrated that this disease is due to a virus and that it is not a stigmonose disease.

Ball, E[lmer] D[arwin]

The leafhopper of the sugar beet and their relation to the "curly top" condition. U. S. Dept. Agri. Bur. Ent. Bull. **66** : 33–52, 1909.

Baribeau, [Charles Henri] Bernard

A disease of the potato. Spindling tuber. Quebec Soc. Proct. Plants. 23d & 24th. Ann. Rept. **1930–32** : 199–200, 1932.

Popular.

Barton-Wright, E[ustace], & McBain, Alan

Possible chemical nature of tobacco mosaic virus. Nature 132 (3348) : 1003–1004, 1933. (Trop. Agric. (Trinidad) **11**(4) : 101–102, 1934.)

Annotated in the bibliography page 41.

———————, **Cockerham, G., & McBain, Alan**

Rept. Director of Res. Scottish Soc. Res. in Plant Breeding Ann. Gen. Meeting 26th July, p. 15–17, 1934.

Report of the results obtained so far in regard to this type of work done by the authors.

Baudys, E[duard]

Fytopatologické poznámky VIII (Zarok 1932). Phytopathological notes VIII (for 1932). Ochrana Rostlin **13**(3–4) : 90–102, 1933.

These notes are mainly on virus diseases of several plants, among which he mentions a case of potato mosaic on tomato as a record in south Moravia; he also reports the occurrence of mosaic on red clover and dahlias. This prevalence of mosaics is attributed in part to drought conditions and lack of proper fertilization.

Bawden, F. C.

Studies on a virus causing foliar necrosis of the potato. Proc. Roy. Soc. London. **B. 116**(799) : 375–395, 1934.

The author describes a virus which he designates as "D". It is the cause of a "foliar necrosis" and in some varieties a "top necrosis". However, "top necrosis" may be due to other viruses. The virus also attacks white burley tobacco, tomato, *Nicotiana glutinosa*, and *Datura stramonium*. He failed to secure successful transmission by insects.

Beauverie, J[ean Jules]

Action du parasite sur la résistance du chondriome-plastidome sa fragilisation et l'altération de a structure cellulaire. Proc. Int. Cong. Plant Sci. Ithaca 1926. **2**: 1299–1311, 1929.

A study of cell contents. Does not discuss virus diseases but is of interest to students of the subject.

Beale, Helen Purdy see **Purdy Beale, Helen**

Bechhold, H., & Erbe, F.

Die Biologie der Kartoffel XVI. Mitteilung. Studie über die Kolloidstruktur der Kartoffelknolle. (The biology of the potato. XVI. Studies on the colloid structure of the potato tuber). Unterschiede zwischen Vital-und Abbauknollen. Arb. Biol. Reichsanst. Land u. Forstw. Berlin **20**: 111–139. 1932.

---------- **& Schlesinger, M.**

Grösse von Virus der Mosaikkrankheit der Tabakpflanze. (The size of the virus of the mosaic disease of the tobacco plant). Phytopath. Zeitschr. **6**(6) : 627–631, 1933.

After detailed and prolonged experiments the authors reach the conclusion that the size of particles of tobacco mosaic are about 50 μμ in diameter. They compared it with several animal viruses.

Enzyme oder Lebewesen? (Enzyme or living entity?.) Kolloid Ztschr. **66**(3) : 329–340, **67**(1) : 66–79, 1934.

The author discusses the enzymatic conception of the viruses. He is against it and in favor of the "living entity" theory.

Beckwith, C[harles] S[teward]

False blossom. Amer. Cranberry Grow. Assoc. Proc. Ann. Meet. **65**: 25–27, 1935.

Bell, A[rthur] F[rank]

Report of the Division of Entomology and Pathology. Part of 34th Ann. Rpt. of the Bureau of Sugar Experiment Stations (Queensland, Australia) pp. 51–72, 1934.

A part of this report is devoted to diseases of sugar cane including tests to determine the effect of chlorotic streak on yields and a progress report on the dwarf disease and the Fiji disease.

Bennett, C[arlyle] W[ilson]

Plant-tissue relation of the sugar-beet curly-top virus. Journ. Res. Agric. **48**(8): 665–701, 1934.

This rather extensive study gives the author's observations in experimental work.

Properties of the sugar-beet curly-top virus. Phytopathology (title) **24**:(10): 1135, 1934.

Berkley, G[arven] H[ugh]

Recent advances in the study of tomato streak and mosaic. Canadian Hort. **58**: 58–59, 1935.

Occurrence of "spotted wilt" of tomato in Ontario. Scient. Agric. **15**(6): 387–392, 1935.

Account of this disease reported to have occurred in 1931 and studied in 1934. Symptoms are described and control measures suggested.

Bewley, W[illiam] F[leming] & Corbett, W.

Mosaic disease of the tomato. Expt. & Res. Sta. Nursery & Market Gard. Indust. Devel. Soc. Turner's Hill, Cheshunt, Hort. Ann. Rpt. **16**: 56–62, 1930.

In this report the authors describe the disease and its behaviour. Also the cultural practices by which the growing tip is removed inducing secondary infections. Two types of the disease are common—ordinary and aucuba mosaic. The last mentioned was transmitted to tobacco which shows symptoms very similar to those of Doli tobacco affected by the one known as Rotterdam B. disease.

Tomatoes: cultivation, diseases, and pests. Min. Agric. & Fish, Bull. **77**, 71 p., 1934.

Virus diseases of tomatoes are discussed in this bulletin.

Birkeland, Jorgen M.

Photodynamic action of methylene blue on plant viruses. Science n.s. **80**(2077) : 357–358, 1934.

Brief article in which the author reviews the work done on the subject in regard to animal viruses. Reports very briefly on an experiment with plant viruses of different types. In conclusion he states that from these experiments it would seem that in general, plant viruses are more resistant to the photodynamic action of dyes than are animal viruses or bacteriophages.

Bitancourt, A[gegislan]

O mosaico e "streak disease" (Mosaic and streak disease). Characas o Quintaes (Brasil) **34**(5)453, 1926.

Protozoarios do mosaico (Protozoa of mosaic). Rev. Agric. Piracicaba, Brasil **3**(1) : 37, 1928.

A criticism of the protozoan theory of sugar-cane mosaic as given by Averna-Saccá.

---, **& Grillo, H. V. S.**

A chlorose zonada uma nova doenca dos citrus. (A zonal chlorosis, a new citrus disease.) Arch. Inst. Biol. Sao Paulo **5** : 245–250, 1934.

Black, L. M.

The potato yellow dwarf disease. Amer. Potato Journ. **11**(6) : 148–152, 1934.

Potato yellow dwarf is transmissible by stem grafting. The clover leaf-hopper, *Agallia sanguinolenta* Prov. is the only vector known. The incubation period in the plant is variable; whether transmission was by grafting or by insect inoculation the incubation averaged 40 days. The virus overwinters in two or more ways: (1) in the potato tuber and (2) in the clover leafhopper or in some host other than the potato.

A mosaic on cabbage in Wisconsin. Phytopathology (Abstract) **25**(1) : 6, 1935.

Blattny, C[tibor Eugen Marie Karel]

Jde u mosaiky Revy Vinné o jediny virus? (Is only one virus involved in vine mosaic?) Ochrana Rostlin **13**(3–4) : 104–115, 1933.

Report of three year grafting experiments to study the nature of vine mosaic in Czechoslovakia. The results are given in tabular form,

they lead the author to believe that there are two different viruses in the mosaic of the vine occurring in his country.

The experiments indicate that this virus disease is distinct from "roncet".

Vertikalni rozsireni virovych chorob. (Vertical spread of virus diseases). Ochrana Rostlin **13**(3–4) : 145, 1933.

The author in a short note reports the spread of virus diseases up to an altitude of about 1,550 meters above sea level. Among those mentioned are potatoes, *Urtica dioica* and *Berberis vulgaris*.

Virové choroby Pelargonii (Virus disease of Pelargonium). Ochrana Rostlin **13**(3–4) : 145, 1933.

A short note reporting at least two virus diseases of pelargoniums in addition to leaf curl, existing in Czechoslovakia. Description of the diseases is given.

Mosaika na Celeru (*Apium graveolens*). (Mosaic of celery. (*Apium graveolens.*) Ochrana Rostlin **13**(3–4) : 145–146, 1933.

Brief note reporting for the first time celery mosaic in Czechoslovakia. The outbreak appeared to be correlated with the prevalence on the crop of the insect *Chlorita flavescens*. Celeriac planted in close proximity to diseased celery plants did not contract the disease.

Bodine, E. W.

Occurrence of peach mosaic in Colorado. Plant Disease Reporter **18**(10) : 123, 1934.

A severe outbreak of peach mosaic, first recorded from Texas, is reported from Mesa County, Colorado, where some of the orchards contain up to 100 per cent infected trees. Eradication work is in progress.

Bohme, R. W.

Das Vorkommen von Virosen auf dem Dahlemer Versuchsfelde. (The occurrence of viruses on the Dahlem experimental field). Arb. Biol. Reichsanst. für Land-und Forstwirtsch; **21**(1) : 1–58, 1934.

Continuation of previous experimental work. In this paper the author gives a very comprehensive account of his observations on the nature of the virus diseases of potatoes.

Bokura, U.

(Dwarf disase of rice plant). Journ. Japanese Agric. Soc. **593** : 56–59, 1930.

Bolley, H. L.

Methods of detecting mosaic being studied in North Dakota. Potato News Bull. **1**(10) : 220–221, 1930.

Popular.

Boning-Seubert, E.

Die Mosaikkrankheit der Gurken. (The mosaic disease of cucumber). Prakt. Blätter für Pflanzenbau & Pflanzenschutz **11**(9–10) : 291–321, 1933–34.

A popular account based on American investigations and literature.

Booberg, K. G.

Over het gebruik van gelestrepenziek plant material. (On the use of yollow stripe-diseased planting materia). Arch. voor Suikerind. Nederl.—Indie, Deel I. **42**(10)) : 319–331, 1934.

This article is a study of statistical data on the transmission of sugar-cane mosaic disease, well tabulated and discussed. The author makes distinction on "liability" to infection and "susceptibility" in connection with mosaic. He states that, due to the use of resistant POJ–2878 variety, sugar-cane mosaic became a minor problem, but on account of the recent planting of very susceptible varieties it has become prominent once more.

Bouffil, F.

Contribution á l'étude de deux maladies de l'arachide. I. II. (Contribution to the study of two peanut diseases. I. II.) Bull. Mens. l'Agence Econ. l'Afrique Occid. France **14**: 3–6, 1933.

Bouriquet, G.

Les maladies du Tabac á Madagascar. (Tobacco diseases in Madagascar.) Ann. Cryptog. Exot. **7**(2) : 97–112. 1934.

This report includes some tobacco virus diseases.

Boysen-Jensen, P.

Die Stoffproduktion der Pflanze (Blattrollkrankheit). Jena, 108 p., 1932.

Brandes, E[lmer] W[alker] & Coons, G[eorge] H[erbert]

Beet crop problems: science helps find the answers. Facts About Sugar **29**(3) : 83-85, (4) : 117–121, 1934.

In this account sugar-beet curly top disease is discussed.

Brehmer, G[ustav] von

Über die Viruskrankheiten verschiedener Kulturpflanzn. (Virus

diseases of different cultivated plants). Rapp. 2 Congr. Intern. Pathol. Comp. Paris **1**: 360–362, 1931.

Sur les maladies á virus de diverses plants cultivées. (On virus diseases of different cultivated plants.) Compt. Rend. 2 Congr. Int. Pathol. Comp. Paris **2**: 433–434, 1931.

Brierley, Philip

Dahlia mosaic and its relation to stunt. Canada, Flor. **29**: 4 9, 16, 1934.

Streak, a virus disease of roses. Phytopathology (Abstract) **25**(1): 7, 1935.

Symptoms of rose mosaic. Phytopathology (Abstract) **25**(1): 8, 1935.

Brierley, W[illiam] B[roadhurst]

Discussion on "Ultra-Microscopic Viruses". Proc. Roy. Soc. **B 104**: 558–559, 1929.

The author states that "the plant pathologist does not attach anything like as much importance to ultra-microscopic characters as the animal pathologist". He emphasizes the importance of the studies on insect transmission and the differences in strains of viruses.

Brown B[enjamin] A[rthur]

Mosaic versus leafroll. Amer. Potato Journ. **3**(4): 121–122, 1926.

Popular.

Bushnell, John

Do potato varieties degenerate in warm climates? Amer. Potato Journ. **5**(8): 245–246, 1928.

Answered in the affirmative although the author does not deny that virus diseases are important factors.

Butler, E[dwin] J[ohn]

Views on the "spike" disease in sandalwood. Reported by M. Muthannab. 6 p. 1904.

A review of the subject.

Some relations between vegetable and human pathology. Trans. Roy. Soc. Trop. Med. Hyg. **15**: 203–211, 1922.

Includes a comparison of plant and animal virus diseases.

Resolution passed by the Fifth International Botanical Congress, Cambridge, England, August 1930. Phytopath. Zeitschr. **4**(2) : 225–226, 1931.

This resolution was passed with tentative suggestions for activities of the International Committee on the naming and description of plant virus diseases.

Butler, O[rmond Rourke]

How often should the potato grower renew his stock? New Hampshire Agric. Expt. Sta. Circ. **45**, 8 p., 1934.

Popular suggestions to farmers giving information in regard to potato leafroll and mosaic diseases.

Caldwell, John

The movement of the virus agent in the plant. Deux Cong. Int. Path. Comp. Paris 1931. **II** Comptes Rend. et Comm. p. 480, 1931.

A brief note.

Possible chemical nature of tobacco mosaic virus. Nature **133** (3353) : 177, 1934.

The author agrees with Barton-Wright and McBain findings of tobacco-mosaic virus in the crystalline part of the mixed phosphate element precipitated from the juice of inoculated *Nicotiana glutinosa* plants, but the amount declines progressively with each successive washing. He gives many data of his observations and conclusions. The virus in the crystals is nothing more than an impurity and the absence of any specific relation between the crystals and the virus is readily demonstrated.

The control of virus diseases of the tomato. Journ. Min. Agric. **41**(8) : 743–749, 1934.

Popular account in regard to virus diseases of tomato in greenhouses. Gives brief descriptions of tomato mosaic, streak, spotted wilt, aucuba mosaic and "double virus" streak. Gives some data on transmission and recommendations for control.

On the interactions of two strains of a plant virus: experiments on induced immunity in plants. Proc. Roy. Soc. **B117** : 120–139, 1935.

Report of experiments with yellow mosaic of tomatoes.

Spurious cucumber "mosaic" due to copper poisoning. Journ. Min. Agric. Gt. Brit. **42**: 97–98, 1935.

Popular.

The physiology of virus diseases in plants. VII. Experiments on the purification of the virus of yellow mosaic of tomato. Ann. Appl. Biol. **22**(1): 68–85. 1935.

This paper gives the results of experiments. The author did not find any evidence that the virus could be recovered in a crystalline form. Viruliferous material always contained traces of organic nitrogen. The virus is active over a wide range of pH from 2.0 to 10.5.

Calinisan, Melanio R.

A suspected "mosaic" of Abacá in the Philippines. The Philippines Journ. Agric. **5**(4): 255–256, 1934.

A record of a new disease.

Campbell, A. H.

Virus diseases of Dahlias. The Dahlia Yearbook **1934**: 14–23, 1934.

Descriptions of dahlia mosaic, streak and spotted wilt. The paper gives practical directions for their control.

Carne, W[alter] M[illard,] & Martin, D.

Apple investigations in Tasmania: miscellaneous notes. Journ. Australian Council Sci. & Indus. Res. **7**(4): 203–214, 1934.

1. The virus theory of bitter pit. 2. Crinkle in apples. 3. Watercore breakdown. 4. The correlation of refractive index and freezing point depression.

Carpenter, C[larence] W[illard]

Pathology: Ann. Rpt. Comm. in charge of the Expt. Sta. for the year ending September 30th, 1933. Proc. H. S. P. A. 53 Ann. Meeting 1934, p. 24–35, 1934.

In an experiment the author kept mosaic leaves of sugar cane of variety Ba–11569 for 10 minutes at a temperature ranging from 52° to 56° C. and attempts were then made to transmit the disease from these leaves to healthy ones of the same variety using Seín's needle prick method; the results of these inoculations indicated that the thermal death point of the virus lies btween 53° and 54° C. Knife transmission of mosaic has been conducted with susceptible varieties. Chlorotic streak developed rapidly on sugar-cane fields after floods during storms. Studies of the phenomenon are in progress.

Carter, Walter, & Crawford, R[aymond] F[rank]

Eutettix tenellus (Baker) as a factor in the production of nicotine for insecticidal purposes. Journ. Econ. Ent. **22**(1): 158–160. 1929.

Report of observations made by the writers including their observations in regard to curly-top of beets.

Castolla, F. de

Court-noué—a mysterious vine disease. Journ. Dept. Agric. Victoria **32**(6): 298–301, 1934.

The author states that recovery from this disease is more common in Australia than in Europe. Discusses the different theories about the real nature of this disease with special reference to the recent paper by Viala and Marsaics attributing the majority of the cases to the fungus, *Pumilus medullae.*

Cation, Donald.

Peach mosaic. Phytopathology **24**(12): 1380, 1381, 1934.

The disease was transmitted by budding although the buds died. The symptoms appear during low temperatures of 40° to 50° F. but not at 75° F.

Cayley, Dorothy M.

Panachure infectieuse (breaking) des tulipes. (Infectious variegation (breaking) of tulips.) Deux. Cong. Int. Path. Comp. Paris 1931. **II** Compt. Rend. et Comm. p. 446–447, 1931.

A brief review.

Chamberlain, E. E.

Tomato mosaic. Its appearance, cause, and preventive treatment. New Zealand Journ. Agric. **48**(6): 344–351, 1934.

Report on the occurrence of tomato mosaic (ordinary and aucuba) in New Zealand. General discussion on the subject of a semi-popular character.

A virus disase of strawberries in New Zealand. New Zealand Journ. Agric. **49**(4): 226–231, 1934.

It is stated that a virus disease is one of the causes of degeneration of strawberries in New Zealand. The author gives description of the symptoms of the disease. The aphis *Capitophorus fragariae* Theo is the vector.

Narrow-leaf a virus disease of tomatoes. New Zealand Journ. Agric. **49**(5) : 257–263, 1934.

A preliminary paper.

Chandler, W. H., Hoadland, D. R., & Hibbard, P. L.

Little-leaf or rosette of fruit trees. II Effect of zinc and other treatments. Proc. Amer. Soc. Hort. Sci. **29** : 255–263, 1933.

Chester, Kenneth, S.

Specific quantitative neutralization of the viruses of tobacco mosaic, tobacco ring spot, and cucumber mosaic by immune sera. Phytopathology **24**(11) : 1180–1202, 1934.

Brief review of other workers on the subject and the literature so far. A detailed account of his studies and observations.

Serological evidence in the study of the relationships of certain plant viruses. Phytopathology (Abstract) **25**(1) : 10, 1935.

Chittenden, F. J.

Mosaic disease of narcissi. Daffodil Year Book, Roy. Hort. Soc. **1933** : 72–73, 1934.

Christoff, Alexander [Cristow.]

Mosaikkrankheit oder Virus-Chlorose bei Apfeln. Eine neue Virus-Krankheit. (Mosaic disease or virus chlorosis in apples. A new virus disease.) Phytopath. Zeitsch. **7**(6) : 521–536, 1934.

Reports a new virus disease of apples and other fruit trees, including pear, quince, apricot, peach and plum. Of occurrence and observations made since 1930 by the author in Bulgarian nurseries. The author also states that he has observed the occurrence of mosaic on a large number of other Rosaceous hosts besides those mentioned above; almond, cherry (*Prunus cerasus, P. avium*), and wild rose. Control measures are suggested.

Clinch, Phyllis, & Loughnane, James B.

A study of the crinkle disease of potatoes and its constituent or associated viruses. Sci. Proc. Roy. Dublin Soc. n. g. **20** (27–40) : 567–596, 1933.

Descriptions of several virus diseases. A thorough account of the authors' observations based on experiments in intertransmission of these virus diseases artificially and by means of the aphid *Myzus persicae.*

Clinton, G[eorge] P[erkins] & McCormick, Florence, A.
Tobacco mosaic. Connecticut Agric. Expt. Sta. Rpt. of Tobacco Station at Wuilsor, Tobacco Stat. Bull. **10**: 75T–82T, 1928.

A popular general account discussed in the following topics: effect on host, cause?, known facts of mosaic and preventive measures.

Conceicao, C.
O mosaico da canna (Cane Mosaic), Fazenda Fluminense, Brasil, **1**(16): 18, 1930.

Conners, I. L.
Thirteenth annual report of the Canadian plant disease survey 1933. p. I–IX, 1–75, 103–128, 1934.

This report contains notes on false blossoms of cranberry.

Cooley, L. M.
Control of raspberry viruses. Amer. Nurseryman **61**(7): 7, 1935.

Source of raspberry mosaic infections and how to get rid of them. Proc. New York State Hort. Soc. **80**: 273–277, 1935.

Coombs, J.
Black currant reversion. Gloucestershire Farmer **3**: 83–87, 1935.

Costa Lima, A[ngelo] M[oreira]
Sobre o mosaico da canna. (About sugar-cane mosaic). Chacaras e Quintaes (Brasil) **34**(1): 30, 1926.

Costantin, Julien [Noel], Lebard, P[aul] & Magrou, J[oseph]
Influence du séjour en montagne sur la productivité de la Pomme de terre. (Influence of mountainous regions on the productivity of potato). Compt. Rend. de Séances de l'Acad. Sci. **193**: 902, 1931.

Précocité productive et resistance a la dégénérescence. (Premature production and resistance to the degeneration). Compt. Rend. Acad. Agric. France **18**: 661–665, 1932.

A review of work of others on sugar cane.

Selection practique da la Pomme de terre en plains et en montagne en vue de combattre la dégénérescence. Conférence faite le 20 octobre (1932) devant la Ligue nationale du lutte contre les enmemis des cultures, 5 avenue de l'Opéra,

á Paris. (Practical selection of potatoes in low and high lands with the view of fighting degeneration. Conference given on October 20, 1932, before the National League against the enemies of crops. Held. at 5 Opera Ave. Paris). 1932.

Évolution de nos conceptions sur la dégénérescence et la symbiose. (Evolution on our conceptions of the degenerations and symbiosis). Ann. Sci. Nat. Bot. **10**(15): 1–53, 1933.

Sereh disease of sugar cane receives some attention in this paper, but there is very little on the other virus diseases.

Cowland, J. W.

Gezira Entomological Section, G. A. R. S. Final Report on experimental work, 1932–33. Gezira Agric. Res. Serv. Ann. Rpt. for the year ended 31st December 1933, p. 107–125, 1934.

Studies of transmission of cotton leaf curl disease by means of the white fly *Bemisia gossypiperda* on cotton plants and other host plants.

Crew, F. A. E., & Lamy, R.

Autosomal colour mosaics in the Budgerigar. Journ. Genetics **30**: 235–241, 1935.

Cristinzio, M.

La "necrosis" del cuore" dei tubori di Patata. (Heart necrosis of potato tubers). Ricerche, osservazioni ed divulgazioni fitopatologich per la Campania ed il Mezzogiorno (Portici) R. Lab. Pat. Veg. Portici **3**: 3–17, 1934.

Report of tests made on Bohms potato variety affected with heart necrosis. Histological examinations were made and the observations reported. The author attributes the disease to the action of a virus in a localized form. Classified according to Quanjer under category V or pseudo-net necrosis.

Cunningham, H. S.

The lily situation. Bermuda Dept. Agric. Bul. **8**: 12–13, 21–23, 1929.

Visited greenhouses in New York. Found the plants grown from Bermuda bulbs of *Lilium harrisii* superior to those grown from other stock. This is due to eradication of disease (mosaic). The mosaic is more marked in low than in high temperatures.

Currie, J. F.

The production of high-grade seed potatoes in North Wales. Journ. Min. Agric. **40**: 316–326, 1933.

Curzi, M[ario]

Propieta e natura di virus delle piante. (The properties and nature of plant viruses.) Rev. di Biol. **16**(2) : 335–352, 1934.

Discussion on the subject under the three theories that have been advanced trying to explain the nature of plant viruses i. e. (1) autocatalytic, (2) microbial, and (3) ultra-microbial.

Le malattie da virus delle piante. (Virus disease of plants.) Testo della Confrenza tenuta alla riunione dei Tecnici Agricoli Fasciti a Udine il 28 marzo 1933. A–XI ampliato e corredato delle documentazione bibliográfica, p. 3–51, 1934.

Daikuhara, G.

(On dwarf disease of rice plant.) Journ. Tokyo Chem. Soc. **25** : 215–253, 1904. (Imp. Agric. Expt. Sta. Bull. **29** : 163–193, 1904. Journ. Japanese Soc. Agric. **255** : 4–8, 1902.)

Dana, B[liss] F.

Progress in Investigation of curly top of vegetables. Ann. Meeting, Oregon State Hort. Soc. Proc. **49** : 95–99, 1934.

Popular review of work started in 1928.

The curly-top disease of vegetables in the Pacific Northwest. U. S. Dept. Agric. Bur. Plant Ind. (Mimeograph) 4 pp.

Appears to be a progress report but gives suggestions for control in tomatoes.

Davidson, J. & Bald, J[ames] G[rieve]

Description and bionomics of *Frankliniella insularis* Franklin (*Thysanoptera*). Bull. Ent. Res. **21**(3) : 365–385, 1930.

Davis, R[obert] L[esley]

Mayagüez sugar-cane varieties resistant to mosaic. Intrn. Sugar Journ. **34**(407) : 434–435, 1932.

Brief review.

Report of the plant breeder. Puerto Rico Agric. Expt. Sta. Ann. Rpt. **1931–32** : 13–22, 1933.

Report of the results of mosaic-resistant varieties and their distribution in the island.

Sugar-cane variety POJ 2878 in Puerto Rico. Puerto Rico Agric. Expt. Sta. Bull. **35**, 45 p., 1934.

Data on yields of POJ 2878 sugar-cane variety planted in some district to replace B.H. 10(12) as mosaic-resistant variety.

Demandt, E.

Samenvatting van de resultaten der vakkenproeven van oogstjaar 1934 over verschillende onderwerpen. (Summary of the results of plot tests during the harvest year 1934 on various lines of investigations.) Arch. Suikerindus. Neder.—Indië, Deel **2**(26) : 937–946, 1934.

This report contains a great deal of data in regard to yields of mosaic-diseased sugar cane.

Demaree, J. B., Fowler, E. D., & Crane, H. L.

Control of pecan rosette with zinc sulfate. Proc. 28th. Ann. Cont. Southeastern Pecan Growers' Assoc. p. 29–37, 1934.

This is not a virus-disease paper, but as pecan rosette has been considered by some investigators as a disease belonging to the virus group and the symptoms are so similar we decided to include this article which might be of interest to students of the subject.

Desai, S. V.

Studies on the nature of the causative agent of the mosaic diseas of tomatoes. Indian Journ. Agric. Sci. **3**(4) : 626–638, 1933.

Description of experiments to study the nature of the virus of tomato mosaic disease.

Dieselben

Die Blattrollkrankheit und unsere Kartoffelernten. (The leaf roll disease and our potato crop.) Arbt. d/D.L.G. Heft 190, 1911.

La maladie de l'enroulement des pommes de terre. (The leaf roll disease of potatoes.) Ann. Serv. Epih. **6**: 1919.

Dix, W[alter]

Ein Beitrag zur Frage des Abbaues der Kartoffel. (A contribution to the problem of degeneration in the potato.) Landb. Jb. **80**(5) : 769–809, 1934.

A detailed account of the experiments conducted by the author at the Kiel Experimental and Plant Breeding Institute. He attributes this disorder to alcohol formation in the tuber with consequent respiratory disturbance and not, at any rate in the first place to virus infection.

Dobrozrakova, T. L.

Neparzituye zabolevanica Kartofelia. (Non parasitic diseases of potatoes.) Bolezni Rastenu. Morbi Plantarum **16**(2): 121–135, 1927.

Report of observations and studies of mosaic, aucuba, leafroll, crinkle, curly dwarf and necrosis of the parenchyma and the vascular tissues. The effect of temperature on these diseases is given under Leningrad conditions. These studies were based on tuber inoculation which were preferred to top inoculations.

Dodds, H. H., & Fowlie, P.

The effect of streak disease on the yield of Uba Cane. Part II. South African Sugar Journ. **18**(4): 241, 243, 1934.

Report on yields of streak disease of sugar cane. Resistant varieties are recommended as means of control.

Doolittle, S[ear] P[olydore], & Sumner, C. B.

Probable occurrence of Australian spotted wilt of tomatoes in Wisconsin. Phytopathology **24**(8): 943–946, 1934.

Brief account of a disease of tomatoes occurring in Wisconsin, which in behaviour, nature and symptoms is very similar to the Australian spotted wilt.

Dover, C. & Appanna, M.

Studies on insect transmission. Entomological investigations on the spike disease of sandal. (20). Indian For. Rec. **20** (1): 1–25, 1934.

Data of experimental evidence in insect transmission of spike disease of sandal.

Insect transmission of spike disease. Indian For. **40**(7): 505–506, 1934.

The author reports the failure to transmit spike disease of sandal by means of *Moonia albimaculata*. He is unable to accept the statement regarding the complete recovery of the "infected" plants.

Drake, C. J., Martin. J. N., & Tate, H. D.

A suggested relationship between the protoplastic bridges and virus diseases in plants. Science **80**(2067): 146, 1934.

The authors base their conclusion in their studies of yellow dwarf of onions. They suggest that the plasmodesma serve as protoplasmic bridges in the intercellular movements, not only of solutions and other substances of various kinds which have to do with the normal functions of the plant, but also of viruses and other disease-producing agents as suggested by Samuel.

Dubois, P.

Maladies de dégénérescense de la pomme de terre. (Degeneration diseases of the potato.) La Vie Agric. & Rurale **20**: 187, 1922.

A discussion of mosaic, crinkle and leaf roll.

Ducomet, Vital

Les maladies de dégénérescense de la pomme de terre. (The degeneration diseases of the potato.) Rev. Hist. Nat. Appl. **3**(1) : 274–283, 1922.

A discussion of leafroll, curly dwarf and mosaic as used in different countries.

----------, **& Diehl, R.**

La culture de la pomme de terre en montagne et les maladies de dégénérescence. (Potato cultivation in the mountains and degeneration diseases.) Compt. Rend. Acad. d' Agric. France **20**(7) : 228–238, 1934.

From the writers' comparative observations they concluded that the influence of the "degeneration" diseases on yield was at least as marked at the higher altitude as at the lower. The symptoms were more sharply defined in the mountains. In some instances the activity of insect vectors of virus diseases was much greater in the low-lying localities.

---------- **&** ----------

La culture en montagne et les maladies de dégénérescence de la Pomme de terre. (Mountain cultivation and degeneration diseases of the potato.) Ann. Agron. **4**(3) : 355–372, 1934.

A more detailed account than the paper above by the same authors.

Dufrénoy, Jean

(Biochemical factors of local immunity in plants.) Rpt. & Proc. 10th Intern. Hort. Congr. 1932. Soc. Nat. Hort. France, Paris, 1933.

Les maladies á virus. Rpt. & Proc. 10th Intern. Hort. Congr. 1932. Soc. Nat. Hort. France, Paris, 1933.

----------, **& Dufrénoy, M. L.**

Cytology of plant tissues affected by viruses. Phytopathology **24**(6) : 599–619, 1934.

The authors discuss the subject and describe their observations in comparison with healthy material. A bibliography of 48 titles is appended.

----------, **& Shapovalov, Michael**

Cytological changes in the callus of the graft union in connection with curly top in tomatoes. Phytopathology **24**(10): 1116–1118, 1934.

Brief description of observations made by the authors.

Un virus des Renonculacées transmissible au *Nicotina tabacum.* (A virus of Ranunculaceae transmissible to *Nicotiana tabacum.*) Comp. Rend. Soc. Biol. (France) **117**(30): 346–348, 1934.

Brief note reporting a virus of Peonia transmissible to tobacco.

Le zinc et la croissance de la Vigne. (Zinc and vine growth.) La Potasse **75**: 137–139, 1934.

Gives the successful results obtained in the application of zinc sulphate and potash in checking "court-noué" of the vine. This disease of obscure nature is regarded by some authors as of a virus origin and very frequently is cited in the virus-diseases literature.

L'immunité des plantes vis-a-vis des maladies á virus. (Plant immunity against virus diseases.) Ann. Inst. Pateur **54**: 461–512, 1935.

Duggar, B[enjamin] M[inge,] & Livingston, L. G.

The location and concentration of the virus of tobacco mosaic within the cells. Amer. Journ. Bot. (Abstract) **20**(10): **679,** 1933.

----------, **& Hollaender, A.**

Irradiation of plant viruses and of micro-organisms with monochromatic light. I. The virus of typical tobacco mosaic and *Serratia marcescens* as influenced by ultraviolet and visible light. II. Resistance to ultraviolet radiation of a plant virus as contrasted with vegetable and spore stages of certain bacteria. Journ. Bact. **27**(3): 219–256, 1934.

Detailed account of the experiments and description of the procedure, giving results obtained.

Thermal inactivation of some tobacco viruses: Standardization, technique and illustrative data. Phytopathology (Abstract) **25**(1): 15, 1935.

----------, **& Mc Alister, D. F.**

Some factors affecting "Longevity" *in Vitro* of viruses of to-

bacco mosaic and of tobacco ring spot. Phytopathology (Abstract) **25**(1) : 15, 1935.

Earle, F[ranklin] S[umner]

Cane shortage coming. Facts About Sugar **22**(44) : 1057, 1927.

A brief note in which the author predicts a shortage of cane production in Cuba as a result of the spread of mosaic.

Eckart, C. F.

The Fiji disease. Hawaiian Planters' Rec. **3**(4) : 175–176, 1910.

Brief note warning the planters against the danger of importing plants and soil from Fiji. At that time the true cause of the disease was not known.

Emmerez de Charmoy, D[onald]d'

Nouvelle contribution a l'étude du streak. (A new contribution to the study of streak.) Rev. Agric. de L'Ille de la Réunion n.s. **39** : 193–202, 1934.

Results of experimental work. The author describes two forms of streak. One of them being destructive.

Erikkson, Jakob.

The mycoplasm theory. Is it dispensible or not? Phytopathology **11**(10) : 385–388, 1921.

This paper is devoted to the mycoplasm theory as applied to parasitic fungi, but at the bottom of page 388 the author states that he suspects "the occurrence of a mycoplasm symbiosis in the life cycle of several other plant pathogens". He gives a list of parasitic fungi and includes tobacco mosaic.

Esau, Katherine

Localization of symptoms during the early stages of curly-top infection in the sugar-beet. Phytopathology (Abstract) **24** (10) : 1144, 1934.

Autogeny of the phloem in sugar beets affected by the curly-top disease. Amer. Journ. Botany. **22**(2) : 149–163, 1935.

The virus causes phloem degeneration, but one or more of the primary sieve tubes differentiate before degenerations becomes perceptible. Pericycle or phloem parenchyma becomes hypertrophied and dies, that is primary hypertrophy and primary necrosis. Cells some distance from sieve tubes are stimulated to growth and division; that is hyperplasia. A large number of the hyperplastic cells undergo

changes characteristic of differentiating sieve tubes, develop slime bodies and plastids; and both slime bodies and nuclei degenerate. The cytoplasm is reduced in amount and the cell walls thicken.

Fawcett, G[eorge] L[orenzo]

El enrolamiento de las hojas de la tomatera. (Leaf rolling of the tomato plant.) Rev. Ind. Agric. Tucumán **20**(3–4): 49–54, 1924.

Agallia stricticollis, which transmits beet curly top in Argentine is capable of transmitting a disease of tomatoes resembling leaf roll if allowed to feed on tomatoes after feeding on diseased beet. Conclusions drawn from cage tests.

Fawcett, H[oward] S[amuel]

Is psorosis of *Citrus* a virus disease? Phytopathology **24**(6): 659–668, 1934.

Description of symptoms of psorosis appearing as a mosaic-like disease affecting young leaves of citrus which was discovered in May 1933. The author believes, based on experimental and observational evidence, that the disease belongs to the virus group.

Folsom, Donald

Important papers published during the past year on degeneration or virus diseases of the potato. Proc. Potato Assoc. Amer. p. 29–33, 1922.

The effect of difference in region upon the natural spread of potato degeneration diseases. Agric. Bull. Bermuda **4**(7): 5–6, 1925.

A résumé of paper by Schultz and Folsom in Journ. Agric. Res. 30(6): 1925.

Tuber-unit seed plots in Maine. 1925. Potato News Bull. **2** (8): 304–305, 1925.

Popular.

Is uniformity of potato seed certification rules possible and desirable. Amer. Potato Journ. **3**: 377–378, 380, 382, 1926.

Die Kartoffelanerkennung in den Vereingten Staaten von Nordamerica. (The certifying of seed potatoes in the United States). Illus. Landw. Zeitung **47**: 43–46, 1927.

A review of Folsom's work by Dr. H. W. Wollenweber.

Comparison of "healthy" Green Mountain Strains and tuber lines in Maine. Proc. of the 17 Annual Meeting of the Potato Asso. of America. December 1930.

Popular.

Potato virus diseases in 1932. Amer. Potato Journ. **10**(11): 224–233, 1933.

Review of recent American and European literature on potato virus diseases.

Potato virus diseases in 1933. Amer. Potato Journ. **11**(9):235–242, 1934.

A summary of 133 papers. (1) An unusually large number of publications; (2) Naming and classifications of virus diseases far from solution; (3) Experimental transmission and field observations have given new light on insects; (4) Natural dissemination varies with location; (5) Knowledge of geographical distribution increasing; (6) Some German workers still claim that virus diseases are due to growing conditions; (7) Virus diseases are important factors in seed improvement; (8) Tuber-indexing impractical without growing the plants; (9) Scientific knowledge of physiology increasing; (10) Data on yields increasing; (11) Progress in seed plot studies; (12) Seed free from masked mosaic and streak increasing; (13) New data on many diseases; (14) Psyllid yellow not proved to be a virus; (15) Cause of Giant Hill unsolved; (16) Some but not all of internal discolorations due to a virus.

Fukushi, Teikichi

Plants susceptible to dwarf disease of rice plant. Trans. Sapporo Nat. Hist. Soc. **13**(3): 162–166, 1934.

Report based on a cage and glass-tube experiments of attempted transmission of dwarf disease of rice plant to cultivated and wild grasses; hosts and the insect vectors are given.

(The relation between *Nepholettix apicalis* Motsch. var. *cineticeps* Uhler and dwarf disease of rice plant). Agric. and Hort. **9**: 669–676, 879–890, 1091–1094. 1934.

Studies on the dwarf disease of rice plant. Journ. Fac. Agric. Hapkaido Imp. Univ. **37**(2): 41–164, 1934.

A most complete discussion of the subject in which the author gives the history and geographical distribution of the disease, a review of the literature, the symptoms and the results of his own

work. Intracellular bodies are always present in diseased rice, in wheat, rye, *Panicum miliaceum, Echinochloa crus-galli,* sub sp. *colona* var. *edulis* and *Alopecurus fulvus,* but were not found in the insects. *Nephotettix apicalis* Motsch. var. *cinticeps* Uhl. is the only vector. Some individuals are viruliferous while others are not. The progeny of infected insects may or may not be viruliferous while those from a cross between uninfective females and infective males are non-viruliferous.

(The earliest record of the insect-transmission of virus diseases.) Journ. Plant Protection. **22**(1) : 38–45. 1935.

Gandrup, Joh.

Verslag over het jaar 1923 door Dr. W. H. Arisz. Med. Beosoekisch Proefstation **36**: 20, 1924.

Refers to "kroekoek and krekoh" which are probably the same as "Kroeppoek".

Garbowski, L[udwik]

Choroby virusowe Ziemniakow w okresie 1928–1932. (Virus diseases of potato during the period 1928–1932.) Prace Wydz. Chor. Tosl. Panstw. Inst. Nank. Gosp. Wiejsk. Bydgoszczy. **13**: 1–136, 1933.

Gardner, Max W[illiam] & Whipple, O. C.

Spotted wilt of tomatoes and its transmission by thrips. Phytopathology (Abstract) **24**(10) : 1136, 1934.

---, **Tompkins, C[hristian] M[ilton] & Whipple, O. C.**

Spotted wilt of truck crops and ornamental plants. Phytopathology (Abstract) **25**(1) : 17, 1935.

Ghimpu, V.

Sur les maladies á virus de *Nicotiana* spp. en Rumaine. (The virus diseases of *Nicotiana* in Rumania). Compt. Rend. 2 Congr. Intern. Pathol. Comp. Paris, **2**: 453–456, 1931.

Giddings, N[ahum] J[ames]

Testing sugar beets for resistance to curly-top. Phytopathology (title) **24**(10) : 1135, 1934.

Gigante, Roberto

Risultati di un espirienza sull ereditarieta della maculature interna dei tuberi di patata. (Results of an experiment on the hereditary nature of internal spotting of potato tubers). Boll. R. Staz. Pat. Veg. ns. **12**(3) : 275–277, 1932.

Report of the results of an experiment with potato affected with a disease which resembles hereditary spotting. The plants did not show the symptoms of the disease but the tubers were affected. The condition is considered to be a virus disease due to the pseudo-net necrosis virus of Quanjer, and transmitted by *Myzus persicae*.

Nota preliminaire sulla "Necrosi del cuore" del tuberi di patata. (Preliminary note on the "heart necrosis of the potato tuber.) Boll. R. Staz. Patol. Veg. n. s. **13**: 155–159, 1933.

Un caso di elevata recettivitá per le malattie da virus presentato da piante di patata provenienti da riproduzione sessuale. (A case of high receptivity to the virus disease shown by potato plants produced by asexual reproduction.) Boll. R. Staz. Pat. Veg. **14**(3): 334–338, 1934.

"The behaviour of potato cultures, sexually and agamically reproduced, against virus diseases is described. The plants obtained from seed have shown a great susceptibility to virus diseases in comparison with the plants obtained from tubers of the variety 'Bianca di Como'. It appears from this that in some cases it is easier and quicker to obtain an improvement from the selection made from tubers than with sexual reproduction."

Gilbert, A[lfred] H[olley]

Spindling-tuber. A new potato disease. Vermont Agric. Ext. Ser. Circ. **28**, 1923.

Popular.

Gladwin, Fred E.

A non-parasitic malady of the vine. New York (Geneva) Agric. Expt. Sta. Bull. **449**: 99–110, 1918.

Account of a disorder of the vine occurring in the experimental grounds. The author does not regard it as a virus disease, but its symptoms are characteristic of diseases belonging to this group.

Goes, O[scar] C[ampos]

O mosaico como factor de perturbacao economica. (Mosaic as a factor in economic perturbation). Est. Expt. Barreiros. Brasil, 1930.

Goidánich, G.

Ricerche sul "deperimento" dei Susini. (Researches on the plum wilt.) Boll. Staz. Pat. Veg. Rome, n.s. **14**(3): 339–381, 1934.

This disease is not attributed to a virus by the author, but due to the similarity of the symptoms described for other phloem necrosis of virus origin we decided to include it.

Grainger, J[ohn]

Some economic aspects of virus diseases in potatoes. The Naturalist pp. 151–153, 1933.

This is a brief paper showing the heavy losses in the potato crop of England as a result of virus diseases.

Graber, L. F. & Sprague, V. G.

Alfalfa yellows. Science n.s. **78**(2026) : 385–386, 1933.

This disease is due to insect injury. It is mentioned here because the name "yellows" may mislead. Some persons who have not seen the paper may suppose that the disease is due to a virus.

Gram, Ernst

II Filtrabelsygdomme hos. Planter. (Filterable virus of plants.) Nordiske Jordbrugsforskeres Kongres i Oslo 1926, Nordisk Jordbrugsforakning Heft **4–7** : 681–685, 1926.

Gratia, André

Pluralité, heterogenetié, autonomie antigénique des virus des plantes et des bacteriophages. (Multiplicity, heterogeneity, antigenic autonomy of the virus of plants and bacteriophages.) Compt. Rend. Soc. Biol. **114** : 1382–1383, 1933.

Des analogies entre les virus des plantes et les bacteriophages; rapprochement avec les tumeurs. (Analogies among plant viruses and bacteriophages; comparison with the tumors.) Compt. Rend. Soc. Biol. **115** : 189–192, 1934.

A brief discussion from the medical viewpoint.

---, **& Manil, P.**

Différenciation sérologique des virus X et, Y de la Pomme de terre chez les plantes-infectées ou portenses de ces virus. (The serological differentiation of the X and Y potato viruses among plants infected by these viruses or carriers of them.) Compt. Rend. Soc. Biol. **117**(31) : 490–492, 1934.

In continuation of previous work of the senior author they prepared sera from potatoes from different sources. The experimental results obtained are discussed.

Les complexes de virus des plantes et la méthode sérologique.

(The complexes of plant viruses and the serological method.) Compt. Rend. Soc. Biol. **117**(31) : 493–494, 1934.

Brief account of experimental results obtained in trying to separate the complex of virus by serological methods following the technique in vogue with certain bacteriophages.

----------, **& Manil, P.**

De quelques échecs de la méthode sérologique appliquée aux virus des plantes. (On some failures of the serological method applied to plant viruses.) Comp. Rend. Soc. Biol., Paris, **118**(4) : 379–381, 1935.

Report of the negative results in serological experiments with a differential view point. The authors ascribe such failures to the mixture of viruses with divergent antigenic properties.

Graves, C. E.

The spindle tuber disease in Irish potatoes. Bienn. Rept. Kansas State Hort. Soc. **39** : 146–148, 1928.

Popular.

Green, D. E.

The virus of spotted wilt in Gloxinias. Gard. Chron. **96**(2488) : 159, 1934.

Note reporting *Gloxinia speciosa, Vicia faba,* and *Convolvulus arvensis* as hosts for the tomato spotted wilt virus.

Gulley, A. G.

The results of legal efforts to remove peach yellows. Trans. Peninsula Hort. Soc. p. 73–75, 1896.

A discussion primarily of costs.

Gulyás, Antal

A dohánylevél elzalagosodása és a mosaikbetegség. Magyar Nemzeti Könyv-Debrecen p. 21–28, n. d.

Guyot, A. L.

Essais de lutte practique contre la chlorose de Pécher. (Test on the practical control of peach chlorosis). Rev. Path. Veg. & D'Ent. Agric. **13** : 66–69, 1926.

Hall, Frank H.

Some disappointing seed potatoes. New York (Geneva) Agric. Expt. Sta. Bull. **422**, (Popular Edition) 8 p., 1916.

This is a brief review of Bulletin No. 422 of the same institution, "Observations on some degenerate strains of potatoes", by F. C. Stewart.

Hamilton, M. A.

Further experiments on the artificial feeding of *Myzus persicae* (Sulz.) Ann. Appl. Biol. **22**(2): 243–258, 1935.

This paper is not a discussion of virus diseases, but it is of interest because the insect is a vector of virus diseases.

Hansen, Henning P.

Inheritance of resistance to plant diseases caused by fungi, bacteria and vira. A collective review with a bibliography. Yearbook Roy. Veter. & Agric. Coll. Copenhagen, **1934**: 1–74, 1934.

Hansford, C[lifford] G[erald]

Annual Report of Mycologist 1933. Uganda Dept. Agric. Ann. Rpt. **1933**(2): 48–51, 1934.

This report includes notes on a serious mosaic disease of *Phaseolus* sp. and soy-beans.

Harrenveld, Ph[ilippus] van

De bibitvoorziening bij de Java-suikerindustrie in verband met de Sereh of zeefvatenziekte. Meded. Proefst. Java Suikerindus. **5**, 33 p., 1917.

Harris, R. V.

Mosaic disease of the raspberry in Great Britain. I Symptoms and varietal susceptibility. Journ. Pomol. & Hort. Sci. **11**(3): 237–255, 1934.

This is a progress report of several years' work and investigations on the symptomatology of raspberry mosaic in England.

The "degeneration" of the strawberry. Imp. Br. Fruit Prod. Tech. Communication **5**: 11–15, 1934.

This work includes four different articles. In the second paper yellow edge, crinkle and gold disease are briefly discussed.

Harrison, A. L.

The effect of mosaic on transpiration of the bean. Phytopathology (Abstract) **25**(1): 18, 1935.

Hartish, J.

Stoffwechselphysiologische Untersuchungen über die Blattrollkrankheit der Kartoffelpflanze. (Metabolic and physiological investigations on potato leaf roll.) Planta **22**(5): 692–719, 1934.

Continuation of previous work. The results obtained so far are surveyed and discussed. Results are given as to his observations on healthy and diseased plants in regard to the metabolic process of dextrine formation. A preliminar study of the effect of increased dextrin production on certain physiological processes, such as assimilation and respiration, has yielded promising results.

Heald, F[rederick] D[e Forest] & Burnett, Grover.

A virus disease of perennial *Delphiniums*. Bulletin Amer. Delphinium Soc. **2**(2) : 14–21. 1934.

This paper reviews the history of this disease, describes the symptoms and gives the results of inoculations of 16 species of wild plants.

Hill, Helen Deuss

A comparative study of certain tissues of Giant-hill and healthy potato plants. Phytopathology **24**(6) : 577–598. 1934.

The author gives her observations comparing healthy plants with giant-hill potato plants. In conclusion she states that the disorganizations observed in photosynthetic and conducting tissues of the giant-hill plants are of related order though of less degree than the disorganizations reported for plants affected with some other virus diseases.

Hino, I.

(Early important records on phytopathological science in the Orient). Agric. & Hort. **2** : 1223–1232, 1927.

List of plants susceptible to mosaic and mosaic-like diseases. Miyakaki Coll. Agric. & Forst. Bull. **5** : 99–111. 1933.

(Teratological ferns caused by a virus disease.) Journ. Japanese Bot. **10** : 377–380, 1934.

Hoggan, Ismé A[ldyth]

Two viruses of the cucumber mosaic group on tobacco. Ann. Appl. Biol. **22**(1) : 27–36. 1935.

This paper was prepared for the purpose of describing two viruses of cucumber mosaic group which may or may not have been described. (1) A yellow cucumber mosaic virus which appeared during experiments with tobacco mosaic. (2) A second virus on tobacco appears to belong to the same group but shows differences from cucumber mosaic virus.

Holmes, Francis O[liver]

A masked strain of tobacco-mosaic virus. Phytopathology **24** (8) : 845–873, 1934.

Account of a masked strain of tobacco-mosaic virus. The author gives description of symptoms and behavior of the disease.

Inheritance of ability to localize tobacco-mosaic virus. Phytopathology **24**(9) : 984–1002, 1934.

The author states that localization of tobacco mosaic virus in *Capsicum frutescens* was to be determined by a dominant Mendelian factor. He describes the effect of tobacco-mosaic virus in plants possessing this factor.

Increase of tobacco mosaic virus in the absence of chlorophyll and light. Phytopathology **24**(10) : 1125–1126, 1934.

Brief account of results obtained in laboratory experiment.

Hopkins, J. C.

Suspected "streak" disease of maize. Rhodesia Agric. Journ. **32** : 234–236, 1935.

Horne, W[illiam] T[itus]

Avocado diseases in California. California Agri. Exp. Sta. Bull. **585**, 72 p., 1934.

This bulletin on diseases in general contains a description of sun blotch (p. 4–6).

Hungerford, Cha[rle]s W[illiam]

Calico and russet dwarf disease of potatoes. Idaho Agric. Expt. Sta. Ann. Rpt. **1920** : 42, 1920. (Idaho Agric. Expt. Sta. Bull. **122**, 1920.)

Brief note on calico and russet dwarf disease of potatoes. This paper does not attribute the cause of the disease to a virus but suggests a bacterial origin.

Hutchins, Lee M[ilo]

Phony peach. A new and dangerous peach disease. Maryland State Hort. Soc. Proc. 34th Ann. Meeting 1932, p. 42–51, 1932.

Popular.

Hyslop, G. R.

Seed production in relation to mosaic diseases. Seed World **33**(13) : 22–24, 1933.

Popular.

Imai, Y.

Studies on the transmission of broad bean mosaic. Trans. Sapporo Nat. Hist. Soc. **13**(3) : 241–245, 1934.

Aphis rumicis, Macrosiphum pisi and *Myzus persicae* were found to be transmiters of mosaic of *Vicia faba*. Details are given of the incubation period. Needle inoculations were successful in transmitting the disease.

Ishikawa, T.

(The merit of Hatsuzo Hashimoto, the earliest investigator of dwarf disease of rice plant.) Journ. Plant Protect. **15**: 218–222. 1928.

Iyengar, A. V. Varadaraja

Conrtibutions to the study of the spike disease of Sandal. (*Santalum album* Linn.) Part XV. The role of plant acids in health and disease. Journ. Indian Inst. Sci. **16A**(13): 139–152. 1933.

The author describes methods by which he compared the composition of spiked and healthy leaves of Sandal. He gives also the composition of both according to his findings.

Contributions to the study of spike-disease of sandal (*Santalum album* Linn.) XVI. Distribution of arsenic in sandal wood treated with sodium arsenite. Journ. Indian Inst. Sci. **17A**: 131–139, 1934.

Deamination in virus-infected plants. Nature **135**(3409): 345, 1935.

This paper is a study of spiked sandal.

Jaggar, Ivan C. & Chandler, Norman.

Big vein a disease of lettuce, Phytopathology **24**(11): 1253–1256, 1934.

This disease was observed by Jagger several years ago. The cause is not definitely known but the disease has some of the characters of the soil borne-mosaic of wheat.

Jivanna Rao, P. S.

Spike disease in sandal, I. The virus theory repudiated. 8p. Reprint from Hindu. Oct. 17, 1932.

The author repudiates the virus theory and supports the physiological theory. To support his point of view he gives a review of the literature and the studies of other workers so far.

Spike disease in sandal. II. Killing of trees condemned. 2p. Reprint from Hindu. Oct. 21, 1932.

Brief popular article. Discusses the nature of the disease and condemns the practice of killing trees.

Spike disease in sandal. III. Physiological cause explained. 2p. Reprint from Hindu. Dec. 16, 1932.

The author discusses briefly the physiology of spiked sandal.

Correspondence. Spike disease of sandal 2 p. Reprint from Madras Agric. Journ. **21**(4), 1932.

Brief notes explaining the physiological theory of spike disease.

Jochems, S[arah] C[amelia] J[ohannes]

Overzicht van de ziekten en plagen der Deli-tabak in het jaar 1931. Med. Deli-Proefst. 2 ser. **73**: 16–17, 1932.

Johnson, Howard W.

Nature of injury to forage legumes by the potato leafhopper. Journ. Agric. Res. **49**(5): 379–406, 1934.

This paper is not strictly on virus diseases, but is of interest to students on the subject. The author makes some statements of particular interest as "all available evidence indicates that the pathological symptoms caused by the potato leafhopper on forage legumes are not due to the transmission of a virus by this insect". He inserts a detailed discussion of the pathological changes underlying the various pathological symptoms.

Johnson, James & Hoggan, Isme A[ldyth]

A descriptive key for plant viruses. Phytopathology **25**(3): 328–343. 1935.

The authors explain the desirability of some system for the classification based on characters of the viruses rather than on symptoms on host plants. They discuss the most important diagnostic characters and present a series of tables, the last one being a descriptive key.

Jöhnsen, Alfred

Zur Blattrollkrankheit der Kartoffel. (On the leaf-roll disease of potato.) Kartoffel Zeitsch. Kartoffelhanges **13**: 150, 1933.

Über die Reisigkrankheit der Rebe. (On the twig disease of the vine.) Der Deutsche Weinbau, **1913**(17–20) 10 p., 1933.

The author describes fully his experiments and observations which lead him to conclude that the disease under study is a true virus disease. He considers the disease to be due to the same cause as "roncet", "court-noué", and leaf curl in other countries.

Jones, Leon K[ilby]

Virus diseases of raspberries in Washington. Proc. Washington State Hort. Assoc. **28**: 279–284, 1932.

A survey giving brief statements concerning virus diseases on red and black raspberries.

Red raspberry mosaic. Washington State Coll. Ext. Serv. Circ. **22**, 4 p., 1934.

A popular discussion of symptoms, cause, importance, distribution and control.

---, **Anderson, E. J., & Burnett, G[rover]**

The latent virus of potatoes. Phytopath. Zeitsch. **7**(1): 93–115, 1934.

The authors give the results of their experiments and observations fully discussed and tabulated. They reach the conclusion that the latent or "healthy potato" virus of American workers is the same as X virus of English workers, while the veinbanding virus often associated with it corresponds to the Y virus of the English, and that Z virus is probably an attenuated form of the Y virus.

Tobacco mosaic on spinach. Phytopathology (Abstract) **24**(10): 1142, 1934.

The rate of spread of the veinbanding on potatoes. Phytopathology (Abstract) **24**(10): 1144, 1934.

Jordi, E.

Di Blattrollkrankheit der Kartoffel. (The leaf roll disease of potato.) Arbtn. d. Auskunftstelle a. d. Landw. Hochschule Riitti-Zollikofen, 1919.

Jorstad, I[van]

Melding om plantesykdommer i land-og hagebruket. VIII. Sykdommer pa Tomater og Agurkvekster. (Report on plant diseases in agriculture and horticulture. VIII. Tomato and cucumber diseases.) Landbruksdirektorens Arsmelding, Tilleg C, 55 p., 1934.

Notes on the symptoms, etiology and control of virus diseases of tomato and Cucurbitaceae of occurrence in Norway.

Karatchevsky, I. K.

(Virus diseases of tomatoes in the Crimea. A year of field observations and experiments. In virus diseases of plants in

the Crimea and the Ukraine) State Publ. Office for the Crimea, Simferopol p. 39–58, 1934.

Detailed descriptions are reported in this paper of the author's observations in regard to "fruit woodiness" of tomato and tobacco mosaic (fern leaf). It concludes with an account of wild host influence and control measures.

(Biochemical studies of the "stolbur" disease of the tomato. In virus diseases of plants in the Crimea and the Ukraine.) State Publ. Office for the Crimea, Simferopol. p. 74–78, 1934.

"Stolbur" is the local popular name for the "fruit woodiness" disease. After discussing the biochemical studies on diseased plants concludes that, based on Dunlap's attempt to classify virus diseases according to the C/N ratio in the diseased plants the result appeared to support the Russian researches in that tomato "stolbur" belongs to the "yellows" group of virus diseases rather than to the true mosaic group.

Die Viruskrankeiten der Tomaten in der Krim. (The virus diseases of tomatoes in the Krim.) In Rischkov, V. L. Viruskrankheiten der pflanzn in der Krim und Ukraina. Forshungs Isnt. der Krim & Inst. f. Pflanzensch. der Ukraine, Krimisdat, p. 39–58, 1934.

Report of observations of the behavior and effect of tomato virus diseases.

Biochemische Untersuchung der Tomatenfruch verholzung. In Rischkov, V.L. Viruskrankheiten der pflanzn in der Krim und Ukraina. Forshungs Inst. der Krim & Inst. f. Pflanzensch. der Ukraine, Krimisdat, p. 74–78, 1934.

Comparison of healthy and virus-diseased tomatoes in these biochemical studies.

Kawamura, T.

Historical review of X-body, with one example, 1934.

Kendrick, James B[lair]

Cucurbit mosaic transmitted by muskmelon seed. Phytopathology **34**(7) : 820–823, 1934.

Based on controlled experiments made by the author it was found that cucurbit mosaic can be transmitted by muskmelon seed.

Keuchenius, P. E.

Waameningen over Ziekten en plagen bij tabak, Med. Bosoekisch-Proefstation **14**: 12, 1915.

Refers to "Kroepoek" and "Knepoh" which are probably same as "Kroepoek".

Keur, John Y.

Studies of the occurrence and transmission of virus diseases in the genus Abutilon. Bull. Torrey Bot. Club. **61**(2): 53–70, 1934.

Detail report of the experiments conducted by the author in seed transmission and grafting of different species of *Abutilon*. The disease is transmitted by grafting and very rarely by seeds.

Klapp, E. L., & Spennemann, F.

Ökologie und Abbau der Kartoffel. (Ecology and potato degeneration.) Pflanzenbau, Pflanzenschutz u. Pflanzenzucht **9**(8): 303–313, 1933.

Continuation of previous work done by the senior author. Discussion of theories in regard to degeneration and environmental effect.

----------, & ----------

Strichelkrankheit und Schernabbau der Kartoffel. Versuch der Analyse eines Falls schwerer, fortschreitender Wuchsstörungen. (Streak disease pseudo-degeneration of the potato. An attempted analysis of a case of severe, progressive growth disturbances.) Pflanzenbau **11**(2): 67–68, 1934.

Report of four years' investigation on the study of the failure of certain potato varieties. The authors believe that it is due to the combined effects of adverse environmental conditions acting on an impaired constitution. They regard the influence of virus diseases, if present, of minor importance.

Klinkowski, M.

Der Kartoffelabbau und seine Diagnose. (Potato degeneration and its diagnosis.) Die Umschau **37**: 198–202, 1933.

Knowlton, George F[ranklin]

Beet leafhopper notes. Utah Acad. Sci., Arts & Letters **11**: 238–239, 1934.

This is not an article on virus disease, but is of interest to students on the subject.

Koch, James, & Johnson, James

A comparison of certain foreign and American potato viruses. Ann. Appl. Biol. **22**(1): 37–54, 1935.

Viruses were obtained from nine foreign countries and compared with American forms, with emphasis on mottle, ring spot and vein-banding viruses. Mottle and ring spot viruses were present in potatoes from all nine countries. Veinbanding was found in potatoes from six countries.

Köch, Karl

Aphid transmission of potato yellow dwarf. Phytopathology **24**(10) : 1126–1127, 1934.

Brief account of test to determine the vector of potato yellow dwarf disease, so far *Myzus persicae* Sulz. appears to be the active vector under field conditions.

Köhler, E[rich]

Die Viruskrankheiten der landwirtschaftlichen Kulturpflanzen. (The virus diseases of agricultural plants.) Mitt. Deut. Landw. Ges. **48** : 572–573, 1933.

Untersuchungen über die Viruskrankheiten der Kartoffel. III. Weitere Versuche mit Viron aus der Mosaik gruppe. (Investigations on the virus diseases of the potato. III. Further experiments with viruses of the mosaic group.) Phytopath. Zeitschr. **7**(1) : 1–30, 1934.

A rather lengthy, comprehensive, and fully tabulated account is given of the writer's further studies on the potato mosaic viruses M 23, H 19, R 77 and G. A.

Virus Krankheiten. In Sorauer's Handbuch der Pflanzenkrankheiten. 1. Band. 2. Teil. 6. Aufl. Berlin, pp. 329–511, 1934.

An extensive review of the subject and description of the diseases, arranged after the host plants.

Beiträge zum Studium des Kartoffelabbaus. Beobachtungen auf dem Dahlemer Versuchsfelde der Biologischen Reichsanstalt. (Contribution to the study of potato degeneration. Observations on the Dahlem experimental field of the National Biological Institute.) Landw. Jahrb. **79**(2) : 205–217, 1934.

The author observed that the Aphis, *Myzus persicae*, from peach trees transmitted leaf roll from diseased to healthy potato plants to the extent of 100 per cent. He described the symptoms produced by different viruses experimentally transmitted.

Ueber die blattrollkrankheit und andere abbouursachen. Kartoffel. Zeitschr. Kartoffelbanges. **14**: 12–13, 1934.

Der nachweis von virus-infektionen an Kartoffelpflanzgut mit der stecklingsprobe, Der Züchter **7**: 62–65, 1935.

Mischimfektionen mit verschiedenen stämmen der ringmosaikvirus (X–virus gruppe) der kartoffel. (Untersuchungen uber die viruskrankheiten der Kartoffel, IV.) Augew, Bot. **17**: 60–74, 1935.

Kramer, S[imon] P[endleton]

Bacterial filters. Science. **68**(1754): 88, 1928.

A brief discussion of bacterial filters which do not permit the virus of mosaic disease of tobacco to pass.

Kranzlin, G[ottfried]

Beitrag zur Kenntnis der Kräuselkrankheit der Baumwolle. (Contribution to the knowledge of the curl disease of cotton.) Der Pflanzer, Daressalam **6**(6–12): 129–145. 161–170, 1910.

Relation of cicadas to leaf curl of cotton.

Beiträge zur Kenntnis der Krauselkrankheit der Baumwolle. (Contribution to the knowledge of the curl disease of cotton.) Der Pflanzer, Daressalam **7**(4): 327–329, 1911.

Relation of cicadas to leaf curl of cotton.

Kunkel, L[ouis] O[tto]

Amoeboid bodies associated with *Hippeastrum* mosaic. Science **57**: 693, 1923.

Reports bodies in *Hippeastrum equestre.*

Plant pathology and man. W. B. Saunders Co. 207 p. 1926–27.

A lecture.

Heat treatment for the cure of yellow and rosette of peach. Phytopathology (Abstract) **25**(1): 24, 1935.

Possibilities in plant virus classification. Bot. Rev. **1**(1): 1–17, 1935.

Discusses the different methods proposed by other workers, the importance of biological carriers, evidence of relationships among the plant viruses and the new methods for differentiating plant viruses. Includes a bibliography of 88 titles.

Kuprewicz, V. F.

(Contribution to the physiology of diseased plants. Physiological data on the injury caused to cultivated plants by some fungus and virus diseases.) Thesis Acad. Sci. U. S. S. R. Bot. Inst. Leningrad 71 p., 1934.

The virus diseases included in this report are potato mosaic, leaf-roll and aucuba mosaic. Conclusions are based on experimental data.

Küster, Ernst

Beiträge zur Kenntnis der panaschierten Gehölze XXIII–XXVII. Mitt. Deutsch. Dendrol. Ges. **41**: 347–356, 1929.

Beiträge zur Kennthis der panaschierten Gehölze XXXI—XXXII Mitt. Deutsch. Dendrol. Ges. **43**: 343–348, 1931.

Beiträge zur Kenntnis der panaschierten Gehölze XXXVII—XXXI. Mitt. Deutsch. Dendrol. Ges. **45**: 286–293, 1933.

Beiträge zur Kenntnis der panaschierten Gehölze. XXXII—XXXIV. Mitt. Deutsch. Dendrol. Ges. **46**: 116–121, 1933.

Beiträge zur Morphologie der panaschierten Gewächse. Bioch. Zentralblatt. **54**(1½): 89–95. 1934.

Kuwana, S. L.

Important diseases of rice crop in Japan. Proc. 4th Pacific Sci. Cong. Java **4**: 203–207, 1929.

Among other diseases reported by the author he mentioned a rice-virus disease called "ine no ishhiku byo" which occurs in the nursery beds and fields while the rice is young.

Lackey, C[harles] F[rankin]

Restoration properties of *Erodium cicutarum* on the attenuated curly-top virus. Phytopathology (title) **24**(10): 1135, 1934.

Larter, L. N. H. & Russell, T. A.

Leaf stripe in maize. Dissertation for the A. I. C. T. A. 1930–31.

Unpublished.

Lehman, S[amuel] G[eorge]

Contaminated soil and cultural practices as related to occurrence and spread of tobacco mosaic. North Carolina Agric. Expt. Sta. Tech. Bull. **46,** 43 p., 1934.

Based on carefully planned tests the author gives yield and loss data. Also reports his observations in regard to soil transmission, its causes, and suggestions to prevent it. The spread of mosaic by laborers chewing or smoking while handling plants is also discussed.

Letoff, A. S.

(Some notes on diseases of newly cultivated bast yielding plants in Daghestan (1930). Diseases and pests of new cultivated textile plants.) Int. New Bast Raw Material, Moscow, p. 37–43, 1933.

In these notes report is made of a disease of *Hibiscus cannabinus* which has characteristics of a virus disease. The disease is described.

Likhité, V. N.

(The nature and relations of the intracellular inclusions present in the mosaic of tobacco.) Meded. Landb Wageningen **34** (1) : 3–28, 1929.

Virus diseases of the tomato. Journ. Indian Bot. Soc. **9**(273) : 114–125, 1930.

The author gives a brief history of the subjects and gives evidence that there is more than one virus.

---, **& Dessi, G. H.**

Starch accumulation in stenosised plants. Current Sci. **3**(8) : 356, 1934.

A brief note calling attention to the accumulation of starch in diseased plants.

Linford, M[aurice] B[lood]

Yellows (*Delphinium* sp.) Plant Dis. Reporter Supp. **65**: 42, 1927.

A record.

Linz, C. V.

Phony peach disease bearing. Flor. Exchange **78**(3) : 33–34, 1934.

Report on quarantine bearing. A new test perfected. Origin of name explained.

List, G. M., & Daniels, L. B.

A promising control for psyllid yellows of potatoes. Science n.s. **79**(2039) : 79, 1934.

This disease has not been proved to be due to a virus but is of interest to workers in virus diseases. The loss in Colorado in 1932 was 8 million bushels of potatoes. No evidence supporting bacterial or virus theories. Believed to be a toxin injected into the plant by *Paratrioza cockerelli.*

Livingston, L. G., & Duggar, B[enjamin] M[inge]

Experimental procedures in a study of the location and concentration within the host cell of the virus of tobacco mosaic. Biol. Bull. **67**(3) : 504–512, 1934.

The results of the studies recorded in this paper are given in the summary as follows: "In this paper experimental evidence is offered which strengthens the view that the virus of typical tobacco mosaic occurs primarily, if not solely, in the protoplasmic components of the cell, rather than in the vacuole. From observations made it would seem clear that the occurrence of intracellular bodies, in hair at least, is coincident with or an accompaniment of relatively high virus concentration. The inclusion bodies, both vacuolate and striated types, are fragile structures disintegrating on contact with the micro-needle or pipette."

Loewenthal, H.

The cultivation of animal and plant viruses. Arch. für Exper. Zellforsch. **15**(2–4) : 403–404, 1934.

A brief summary of the progress in virus cultivation since 1925 is given by the author. He expresses the opinion that progress in the cultivation of plant viruses, as opposed to animal viruses has been delayed by the difficulty of finding a medium analogous to that made from blood plasma and extracts, and suggests that the use of some lately discovered growth-promoting substances might give valuable results along this line.

Loree, R[obert] F[arls]

Virus diseases. In raspberry growing in Michigan. Michigan Expt. Sta. Circ. Bull. **152** : 34–36, 1934.

Brief popular discussion on the following virus diseases of raspberries; mosaic, curl and streak.

Ludwig, O.

Über Viruskrankheiten bei Pflanzen. (On virus diseases of plants.) Med. Klin. **1** : 52–55. 1933.

Ueber Viruskrankheiten bei Pflanzen. (On virus diseases of plants.) Med. Klin. **2** : 1–10, 1933.

Lushington, P. M.

Note on spike disease of sandal. Ind. For. **42**: 65, 1916.

In this note the author reports *Zizyphus aenoplia, Dodonaea viscosa, Pterolobium indicum, Argyreia cuneata, Solanum indicum, Cassia auriculata* and *Ficus tsiela* as presenting similar symptoms as those of spike disease of sandal.

Lutman, B[enjamin] F[ranklin]

Relation of structure of potato leaves to tip burn. Potato Mag. **5**: 1, 6; 22–23, 1922.

Tip burn is severe when degeneration diseases are present.

Mac Clement, D.

Purification of plant viruses. Nature **133**(3368): 760, 1934.

Description of a method adapted from one used by Warburg and Christian for the purification of a water-soluble ferment. It proved effective in the preparation of a purified suspension of any of the "X" group of plant viruses.

Malherbe, I. de V.

Little-leaf or rosette of fruit-trees. Farming South Africa. **9**: 312, 315, 1934.

Malhotra, R. C.

The effect of mosaic on the reserve materials in *Solanum tuberosum*. Biol. Gen. (Vienna) **9**(1): 257–262, 1933.

A brief review of the work of others.

Mandelson, L. F.

Citrus psorosis control. Department of Agric. and Stock. Queensland, Div. of Plant Path. Advisory Leaflet No. **8**. 4 p., 1933.

This paper gives a brief discussion of symptoms, varietal susceptibility, cause, conditions favouring the disease and control. The author states that it is probably due to a very slow growing organism. It is inserted here because there is some recent evidence in America that it is due to a virus.

The importance of tobacco mosaic. Queensland Agric. Journ. **42**(5): 538–545, 1934.

Popular paper warning the growers of the importance of the dangers of tobacco mosaic. Gives symptoms, effects, nature, manner of spread and control of the disease.

Manil, P.

Note sur les nécroses foliaires du tabac dans les cultures de

vallés de la Semois en 1934. (A note on the leaf necroses of tobacco grown in the Semois valley in 1934.) Bull. Inst. Agron. & Stat. Rech. Gembloux **3**(4) : 367–377, 1934.

Report of a disease on tobacco of new occurrence in the Semois Valley, Belgium. The symptoms of the disease are described, no fungus or bacterium was found, although ordinary virus of ordinary tobacco mosaic was isolated in three occasions from the spots, it has not been determined if there is any association of the disease and any virus disease; nevertheless, from the evidence obtained the author concludes that this disorder is due to one or more specific viruses.

De la différenciación de certains virus phytopathogénes par l'action des complexes. (On the differentiation of certain phytopathogenic viruses by the action of complexes.) Compt. Rend. Soc. Biol. Paris **118**(4) : 376–379, 1935.

Report of experimental work on inoculations under controlled conditions.

Manns, T[homas] F[ranklin]

Our present knowledge on the dissemination of yellow and little peach. Trans. of Penn. Hort. Soc. Bull. of the State (Delaware Board of Agric. **23**(5), 1933.

The author reports that *Macropsis trimaculata* (the vector for peach yellows and little peach) lives primarily on the plum. He finds it in abundance *P. salicina* and other Japanese varieties and very sparing on the peach. Budding experiments show that plums are carriers of these diseases and that they show few or no symptoms. The death rate in peach orchards is greater when near plums than when not near them. "It is quite probable that the plums are the hosts which have brought the viruses of little peach and yellows into America.

---, **& Manns, M. M.**

Plums as factors in the dissemination of yellows and little peach. Trans. Penn. Hort. Soc. **24**(6) : 72, 1934.

This is a continuation of the preceding paper. The Japanese plums (*Prunus salicina* and *P. simonii* and varieties) carry peach yellows and little peach, with more or less masking. The European (*P. domestica*) and American (*P. Americana*) are not as good carriers as the Japanese species. The Oriental specie (*P. myrobalon* and *P. munsoniana*) are capable of masking both yellows and little peach.

NOTE: Dr. Manns has been studying peach yellows and little peach for many years. Reports of progress are in the Annual Reports of the Delaware Agricultural Expt. Station.

--- **& Adams, J[ames] F[owler]**

Department of Plant Pathology. Delaware Agric. Expt. Sta. Ann. Rpt. **1932–33** (Bull. **188**) : 36–46, 1934.

Studies on the masking of yellows and little peach in other species of *Prunus* indicated that some varieties of plums may act as carriers of these diseases, which can be disseminated from them by the leafhopper *Macropsis trimaculata* which lives principally in plums and very seldom is found on peach trees. Budding experiments are described.

Manzoni, L.

Attenti al roncet. (Attention to roncet). Battaglie Rurali **2**(3) : 2, 1933.

The author regards "roncet" or leaf roll of the vine to be caused by a filterable virus. In his opinion the disease was introduced into Italy with susceptible varieties from America. He gives some methods of control.

Marchal Emile [Jules Joseph]

Observations et recherches effectués á la Station de Phytopathologie de l' Etat pendant l'année 1933. (Observations and researches carried out at the State Phytopathological Station during the year 1933). Bull. Inst. Agron. and des Stat. de Res. de Gembloux. **III**(2) : 97–106, 1934.

This report contains interesting notes on sugar beet, mangolds, tobacco and dahlia mosaic.

Martin, W[illia]m H[ope]

Influence of degenerative diseases on yield. New Jersey Hints to Potato Growers **7**(6) Oct. 1926.

Popular.

Martinoff, S. I.

(Mosaic or Reisigkrankheit of the vine.) Agriculture, Sofia **38**(2) : 6, 1934.

As results of an official survey, the author found a vine disease, which was of some importance in 1930, spreading rapidly and causing considerable losses. He states that the disease shows much resemblance to vine mosaic in Czecho-slovakia. Reviewing the literature on the subject he notes what has been named as "mal nero", "reisigkrankheit", "roncet", "court-noué", "rougean", "brunissure", etc., are but different manifestations of the virus troubles, either due to varietal responses of the host or to ecological conditions. Description is given of the symptoms. No conclusion has been reached as to insect vectors, although the author suggests the possibility of two forms of *Phylloxera* (*P. vastatrix*, *P. gallicola* and *P. radicicola*). In the opinion of the author this disease is widely distributed all over the world and control measures are needed.

Martyn, E[ldred] B[ridgeman]

Mosaic disease of cane. Agric. Journ. British Guiana **2**(2): 112–113. 1929.

The author states that there is some doubt among a number of planters and others concerned with the sugar industry of the colony as to the appearance and nature of mosaic disease of sugar cane. Although the disease has been wide-spread over the world it is absent from British Guiana. The author gives a brief description of symptoms.

Botanical and Mycological Division. Annual Report 1929. Agric. Journ. British Guiana **3**(4): 226–233, 1930.

(This annotation is to correct the one on **18**(1–2): 239 which is wrong.) "The mosaic disease, though not yet known in British Guiana, occurs in Surinam, and I was enabled to see instances of it on an estate in the neighborhood of Nickerie. The cane principally affected was D–625, the variety so universally grown in this colony. At one time, I was informed, several fields had been badly infected and no steps having at first been taken to control the disease, up to 100 per cent infection had occurred upon 3rd and 4th ratoons. But by rouguing and replacing with the more resistant D–109, or in the worst areas by Uba, the disease has subsequently been kept under control, and has never spread from the section of the estate where it originally appeared." (Change made at the request of the author.)

Report of the botanical and mycological division for the year 1932. British Guiana Dept. Agric. Divisional Rpts. **1932**: 117–121, 1934.

Brief notes on sugar-cane mosaic and Liberian coffee-phloem necrosis.

Massee, A. M.

On the transmission of the strawberry virus "yellow edge" disease by the strawberry aphis together with notes on the strawberry tarsonemid mite. Journ. Pom. & Hort. Sci. **13** (1): 39–53, 1935.

This paper is a record of the study of the transmission of this disease by insects. The disease is transmitted by the strawberry aphis (*Capitophonis fragariae* Theo.) during June but not during the latter part of July and August. It is not transmitted by the strawberry tarsoneimid mite (*Tarcossemus fragariae* Zimm.).

Massey, R. E.

Section of Botany and Plant Pathology, G. A. R. G. Final Reports on experimental work in 1932–33. Gezira. Agric. Res.

Serv. Ann. Rpt. for the year ended 31st December 1933, p. 126–146, 1934.

In this report cotton-leaf curl disease is included and description of experimental work described as well as its insect vector *Bemisia gossypiperda.*

Matsumoto, Takashi, & Somazawa, Koetsu.

Immunological studies of mosaic diseases 1. Effect of formolization trypsinization and heat--inactivation on the antigenic properties of tobacco mosaic juice. (The Phytopathological Laboratory, Taihoku Imperial University, Contribution No. 8.) Journ. of Society of Tropical Agriculture **3**: 24–33, 1931.

Immunological studies of mosaic diseases. IV. Effects of acetone, lead subacetate, barium hydroxide, aluminium hydroxide, trypsin and soils on the antigenic property of tobacco mosaic juice. Journ. of the Soc. of Trop. Agric. **6**: 671–682, 1934.

The author partially purified tobacco mosaic by the use of acetone, lead subacetate, barium hydroxide, aluminium hydroxide, etc., and found that mosaic juice remained antigenic as long as it was infectious. Confirmed Lojkins and Vinson's results that trypsin was able to inactivate the infectivity of virus when treated with acetone. The antigenic property and infectivity of the virus were impaired by passing through soils, especially sterilized and dry soils. The serological reaction may be due not to modified plant protein, etc., but to antigenic properties of the virus. It appears probable that the virus may be absorbed by the roots of plants in nutrient solutions.

Mc Carthy, C.

Progress Report of Forest Adminitration in Coorg for 1898–1899.

This report contains the first published record on spike disease of sandal that has come to the attention of the compilers.

Unpublished memorandum on the future supply and culture of sandal in Coorg, 1899.

In this memorandum the external symptoms of spike disease of sandal are well described.

Mc Clean, A. P. D.

Streak disease of sugar cane. Proc. South African Sugar Tech. Asso. **7**: 73–79, 1933.

Mc Cubbin, W[alter] A[lex]

Peach yellows report 1929. Proc. State Hort. Assoc. Pennsylvania **7**: 113–118, 1930.

A survey.

Mc Kenny-Hughes, A. W.

Les pucerons comme vecteurs du *breaking* des tulipes. (Plant lice as vectors of "breaking" of tulips.) Deux. Cong. Int. Path. Comp. Paris. II Compt. Rend. et Comm. p. 447–449, 1931.

A brief review with special reference to insects.

Mc Kinney, H[arold] H[all]

Etude sur les mélanges de virus. (Study on virus mixtures). Deux. Cong. Int. Path. Comp. Paris 1931. II Comp. Rend. et Comm. p. 449–453, 1931.

A brief review.

Mc Rae, W[illiam]

Effect of mosaic on the tonnage and the juice of sugar cane in Pusa. Indian Journ. Agric. Sci. **1**(5): 527–533, 1931.

Based on analysis and experiments the author illustrates with tables the results obtained in general. The loss in purity is more appreciable in Coimbatore seedlings than in tonnage.

----------, **& Subramaniam, L[ekshminarayapuram] S[ubrania]**

Effect of mosaic on the tonnage and the juice of sugar cane in Pusa, Part III. Indian Journ. Agric. Sci. **3**(5): 870–880, 1933.

In continuation of previous work it was found during the season 1932–33 from the experimental plots, that less juice was extracted from mosaic-infected than from healthy canes. There was slightly less glucose from diseased canes than from healthy ones, but the other differences were so small that they were not statistically significant.

Effect of mosaic on the tonnage and the juice of sugar cane in Pusa, IV. Indian Journ. of Agri. Science. **4**(5): 787–796. 1934.

This paper gives the results of field studies.

Mc Whorter, F[rank] P[aden] & Bouquet, A. G. B.

Suggestions for the control of tomato and tomato mosaic and streak. Oregon Agric. Expt. Sta. Circ. Inf. **84**, 4 p. 1933.

English form of tomato spotted wilt found in Oregon greenhouse. U.S.D.A. Br. Plant Indus. Plant Disease Reporter **18**(5) : 25–26, 48.

Based on observations made on the ornamental plants or native weeds in association with tomatoes affected by the English form of spotted wilt, it is concluded that the disorder must have been introduced on seed imported direct from England.

Megaw, W. J.

The improvement of stocks of potatoes by selection. Journ. Min. Agric. Northern Ireland **3** : 131–136, 1931.

A discussion of rate of degeneration, prevention of deterioration, propagation of selected stock and data on yields.

Mejía, R.

El mosaico de la caña en Antioquía. Bol. Agric. Soc. Antioquía Agric. Colombia. **8**(194) : 3–9, 1934.

Merkenschlager, F[ritz]

Zur Diagnose und Pronose des Pflanzwerters der Kartoffeln. (On the diagnose and prognose of the value of the potato plants.) Der Kartoffelbau **16** : 109–111, 1932.

--- **& Klinkowski, M.**

Ueber die Degeneration der Kartoffelabbau). (On the degeneration of the potatoes.) Wiener Landw. Ztg. **82** : 67–68, 1932.

Meyer, H[ans]

Das Chlorose—und Panachureproblem bei Chlorellen. II. Beih. Bot. Centralbl. **51** : 170–203, 1933.

Mikhailova, P. V.

(Anatomy of tomato plants affected with fruit woodiness. In virus diseases of plant in the Crimea and the Ukraine.) State Publ. Office for the Crimea, Simferospol p. 79–92, 1934.

The author states that tomato fruit woodiness present in the structure of the mesophyll and in the diseased leaves is characteristic of virus diseases.

--- **& Pivovarova, R. M.**

(Consideration on the anatomical method of diagnosing virus diseases of the potato. In virus diseaes of plants in the Crimea and the Ukraine.) State Publ. Office for the Crimea. Simforopol, p. 93–108, 1934.

Based on the anatomical changes that virus diseases cause in the tuber of the potato plant, the authors discuss and give the results of their studies arranged in tabular form. The results given show that they are inclined to accept Quanjer's classification rather than the views given by von Brehmer and Rochlin.

M'Intosh, T. P.

Potato notes. Gard. Chron. **91**: 66–67, 1932.

Brief popular notes.

Moore, E[nid] S[tella]

The kromnek disease of tobacco and tomato in the eastern Cape Province. Farming So. Africa **8**: 378–380, 1933.

General popular account.

---------- & ----------

Über die anatomische Diagnotikmethode der Kartoffelnviruskrankheit. (On the Anatomical diagnostic method of potatoes virus disease.) In Rischkov, V., Viruskrankheiten der pflanz in der Krim und Ukraina. Forshung Inst. der Krim & Inst. f. Pflanzench. der Ukraine, Krimisdat p. 93–108, 1934.

Same as above.

----------, **& Wager, V. A.**

Kromnek: A serious tomato disease. Farming in South Africa, June, 1934.

Popular account warning the growers.

The leaf curl or crinkly dwarf disease of tobacco. Farming in South Africa, June, 1934.

Brief popular description of the disease giving control measures.

Morgenthaler, O.

Die Blattrollkrankheit der Kartoffel, eine Infektion oder eine Ernährungs-storung? (Leaf roll disease of the potato, an infection or a nutritional disturbance?). Mitt. Naturforsch. Gesellsch. Bern. 1933, p. 44–45, 1934.

Brief notes on the history of leaf-roll disease during the last thirty years.

Morris, O. M.

Apple rosette. Wahington Agric. Expt. Sta. Bull. **177,** 30 p., 1923.

The author states that apple rosette is a functional or nutritional disorder of apple and pear trees. After discussing its relation to

soil, fertilizer, pruning and other cultural practices concludes that growing a leguminous cover crop or shade crop in the affected orchard gave the best results in recovery from the disease. The author does not consider apple rosette as a virus disease.

Morstatt, H[ermann] A[lbert]

Die Degeneration bei unseren Kulturpflanzen, (Degeneration in our crop plants.) Blatt Pflanzenbau u. Pflanzenzucht **1**: 49–51, 1923.

Brief popular account on deterioration.

Viruskrankheiten der Pflanzen. (Virus diseases of plants.) Pflanzenbau **1**: 57–58, 1924.

Brief popular account.

Entartung, Altersschwache und Abbau bei Kulturpflanzen, insbesondere der Kartoffel. (Degeneration, senile decay and running out of cultivated plants, especially the potato.) Naturw. und Landw. Heft. **7**, 74 p., 1925.

The author discusses the concepts of degeneration, senile decay and running out, and develops his theory of an ecological basis for deterioration, especially of potatoes.

Der gegenwartige Stand unserer Kenntnis der Degeneration. (The present position of our knowledge on degeneration.) Angeu. Botanik **13**: 81–83, 1931.

Brief account of ecological theory of deterioration at this time.

Degeneration bei Kulturpflanzen und die Frage ihres Vorkommes bei Sisal. (Degeneration on cultivated plants and the question of its occurrence on Sisal). Tropenpflanzen **34**(3): 95–99, 1931.

Essentially the same as the above citation. There seems to be no deterioration in Sisal.

Morwood, R. B.

Potato diseases. Queensland Agric. Journ. **40**: 382, 395, 1933.

Potato diseases. Queensland Dept. Agric. & Stock. Phytopath. Leaflet **23**, 12 p., 1933.

Popular.

Mottet, S[eraphin Joseph]

La dégénérescence de la pomme de terre. (The potato degeneration.) Journ. Soc. Nat. Hort. France **4**(23) : 263–268, 1922.

Murata, T. J.

(Insect pests of the rice and barley and their control.) 364 p., 1915.

(Dwarf disease of rice plant.) Journ. Japanese Agric. Soc. **604** : 47–50, 1931.

Murphy, Paul A[loysius]

Potato inspection service. Agric. Gaz. **6**(3) : 1–7, 1919.

Popular account about potato seed inspection, specially in regard to the so-called "running out" which in the opinion of the author is mainly caused by mosaic or leaf-roll diseases.

Discussion on "Ultra-Microscopic Viruses". Proc. Royal Soc. **B 104** : 540–542, 1929.

The author expresses the opinion that plant virus diseases are more homogeneous than animal virus diseases. He divides the plant virus disease into two groups—those that are transmitted mechanically and those only by grafting or budding. He also states that he has found individual plants of the "up-to-date" variety of potato that were virus-free. He states that "perhaps the most important difference between plant and animal virus diseases consists in the very great regularity with which insects act as vectors of the virus diseases of plants".

Nattrass, R. M.

Annual Report of the Mycologist for the year 1933. Cyprus Dept. Agric. Ann. Rpt. **1933** : 48–57, 1934.

This report contains brief notes on potato and tomato virus diseases.

Neill, J. C., Brion, R. M., & Chamberlain, E. E.

"Sore-shin": a virus disease of blue lupines. New Zealand Journ. **49**(3) : 139–146, 1934.

The authors report the occurrence of a disease on blue lupine in New Zealand and call it "sore-shin". Description is given of the symptoms of the disease. From diseased material a *Fusarium* and bacteria were isolated but both failed to transmit the disease. Virus inoculation resulted in successful reproduction of the disease. No transmission occurred through the seed.

Nelson, Ray

Mosaic disease problems. Light thrown on this serious disease by a new discovery by Ray Nelson of Michigan Station. Michigan Quart. Bull. **5**(3) : 128–130, 1923.

Popular notes in relation to the author's paper which appeared in Tech. Bull. 58 of Michigan Agricultural Experiment Station.

Neuwiler, E.

Kartoffelanbannesluucke der Vereiningung Schwerzerijcher Verjuchs = Vermittlungegtellen fur Saatkartoffeln. (Potato cultivation experiments of the Association of Swiss Experiment Stations and Agencies for seed potatoes.) Landw. Jahrb. der Schweiz **45** : 513–538, 1931.

The results of field studies over a series of years.

Newton, William

Virus diseases of potatoes in British Colombia. Amer. Potato Journ. **8**(1) : 13–15, 1931.

Popular. Degeneration strains of potatoes were recognized by farmers before we knew much about virus diseases.

Nielson, Olaf

Kartoffelsorter og Kartoffelsygdomme. Fortsatte orienterende underspgelsen. (Potato varieties and potato diseases .) Saertryk of Tidsskrift for Planteavl, Kobenhavn. Bind. **40** (1) : 105–118, 1934.

This paper includes a very brief discussion of mosaic and leaf-roll diseases of potatoes.

Noble, R[obert] J[ackson]

Australia: Success in control of bunchy top disease of banana in New South Wales. Intern. Bull. Plant Prot. **7** : 195, 1933.

Australia: summary of plant diseases recorded in New South Wales for the season 1932–1933. Internt. Bull. Plant Protect. **8**(1) : 3–5, 1934.

The virus diseases reported were: Fig mosaic is widespread but generally unimportant. Pecan rosette was recorded for the first time. Mosaic occurred on *Zinnia elegans* and Iceland poppy (*Papaver nudicaule*).

Ocfemia, G[eraldo] O[ffimaria]

The transmission of the Fiji disease of sugar cane by an insect vector. University of the Philippines Nat. and Appl. Science Bull. **3**(3) : 277–280, 1933.

The author gives evidence that this disease is transmitted by *Perkinsiella vastatrix*. This is followed by a comparison of this disease with the bunchy-top of Abacá which is transmitted by *Pentalonia nigronervosa*.

Bunchy-top of Abacá: its nature and control. Philippine Agric. **23**(3): 174–186, 1934.

Continuation of previous work cited above.

---------- **& Buhay, G. G.**

Bunchy-top of Abacá or Manila hemp: II. Further studies on the transmission of the disease and a trial planting of Abacá seedlings in a bunchy-top devastated field. Philippine Agric. **22**(8): 567–581, 1934.

Due to the fact that bunchy-top of Abacá in the Philippine Islands has not been found spontaneously on bananas and that the authors were unable to transmit it by means of *Pentalonia nigronervosa* they concluded that the virus of banana bunchy-top in Australia is different from that on Abacá in the Philippines. Data are given on the transmission experiment and procedure for rehabilitation of devastated fields due to the disease.

Bunchy-top of Abacá. Univ. of the Phil. Coll. of Agric. Exp. Station. Circ. **27**, 13 p., 1934.

A popular paper giving symptoms of the disease, methods of spread and control.

Our work on plant diseases. Philippine Agric. **23**(5): 467–475, 1934.

A popular account for the Silver Jubilee number in which the author gives brief notes on the work done on plant pathology in the Philippine College of Agriculture during the last 25 years. He gives special attention to the following diseases: Abacá bunchy-top, Fiji and mosaic diseases of sugar cane.

Ogilvie, L[awrence], Swarbrick, T., & Thompson, C. R.

A note on a strawberry disease resembling the American "crinkle". Agric. & Hort. Res. Sta. Univ. Bristol, Ann. Rpt. **1933**: 96–97, 1934.

Brief note reporting a disease very similar to that described by Zeller and Vaughan in United States known as crinkle and classed under the virus group diseases.

Oortwijn Botjes, J. G.

Die Gesunderhaltung der Saatkartoffeln. (The maintenance of healthy-potato seed.) Pflanzenbau (Berlin). Feb. 15, 1925.

The results of studies on field experiments in which the fields were planted on different dates.

De invloed van abnormale minerale bemestingen op de aardappelplant. De vatbaarheid voor viruszickten in verbani met abnormale Kalibemestingen. (The effect of abnormal mineral fertilizing on the potato plant. The susceptibility to virus diseases in connection with abnormal potassium fertilizing.) Landbouwk. Tijdschr. Maandbl. Nederl. Genootsch. Landbouwwetensch. **44**: 749–754, 1932.

Opitz, K., Tamm, E., Goepp, K., Rathsack, K., & Soltan, F.

Beiträge zur Kartoffelbau, insbesondere zum Abbauproblem. (Contribution to potato cultivation, specially in connection with the degeneration problem.) Landw. Jahrb. **79**(5): 737–781, 1934.

This is a detailed report of five years, experiments conducted by the authors. It is very comprehensive and the results obtained well tabulated. Reference is made to the works of others in regard to degeneration diseases of the potato.

Orton, C[layton] R[oberts]

The virus diseases of plants. Bull. Amer. Delphinium Soc. **4**: 26–33, 1933.

Popular.

Orton, W[illiam] A[llen]

Phytopathology. Environmental influences in the pathology of *Solanum tuberosum*. Journ. Washington Acad. Sci. **3**(7): 180–190, 1913.

In this paper the only reference in relation to virus diseases is on page 189, which states: "Both leafroll and curly dwarf develop suddenly from hitherto healthy stocks and both are transmitted by planting tubers from diseased plants. That whole districts should be affected as in Westphalia in 1907 and in Colorado in 1911 indicates a physiological deterioration due to environmental relations unless a parasite should be demonstrated, which has not yet been done."

Osborn, H. T.

Incubation of the virus of pea mosaic in the aphid, *Macrosiphum gei*. Phytopathology (Abstract) **25**(1): 31, 1935.

Incubation period of pea mosaic in the aphid, *Macrosiphum pisi*. Phytopathology **25**(2) : 160–177, 1935.

Detailed discussion based on observations made in experimental work. It was difficult to transmit this mosaic by mechanical methods.

Palm, B[jorn] T[orwald]

The gametophytes in a composite affected with "Aster yellows". Svensk Bot. Tidskr. **27**(4) : 420–437, 1933.

In plants of *Troximon glaucum* spontaneously affected by aster yellows in Colorado, the author did not observe any abnormalities in the development of the anthers, tapetal, and sporogenous tissues, and male gametophytes. Other abnormalities or occurrence are described. He assumes the theoretical possibility that the pollen from an infected plant may convey the virus to the embryo sac of a healthy one in the process of fertilization.

Pape, H[einrich]

Die Mosaikkrankheit der Lilien. (The mosaic disease of lilies.) Gartenwelt **37** : 324–325, 364, 1933. (Zentrabl. Bakt. (Abstract) II **89**(17–20) : 431–432, 1934.)

Description of the disease which is transmitted by *Aphis gossypii*. Behavior of the disease is given and varietal susceptibility.

Patch, Edith M[arion]

Aroostook potato insects. Journ. Econ. Ent. **15** : 372–373, 1922.

List of 21 species in addition to aphids. Some may transmit degeneration diseases.

Marooned in a potato field. Sci. Mo. **15** : 166–180, 1922.

Discussion of a species of aphid that transmits diseases and overwinters on rose bushes.

Potato aphids. Maine Agric. Expt. Sta. Bull. **323**, 1925.

A discussion of aphids infecting potatoes and other solanaceous plants. This is of interest because some of these insects are vectors of virus diseases.

Park, Malcolm

Bunchy top disease of plantains. Ceylon Dept. Agric. Leaflet **18**, 2 p., 1934.

Popular.

Peacock, Walter M[iller]

Rouguing seed potato fields. Potato News Bull. **1**(10):214-216, 1924.

Popular.

Peltier, George L[eo]

A mosaic disease of wheat. Boston meeting. Plant Path. Section (Abstract) Dec. 27–30, 1922.

Paper read. A record of the disease.

Perret, Claude

Maladies de la pomme de terre. Maladies de dégénérescence. Rapport sur le fonctionnement de l'Institut des Recherches Agronomiques pendant l'année 1931. (Diseases of the potato. Degeneration diseases. Report on the work of the Agricultural Research Institute for the year 1931.) Rep. Francaise, Min. Agric. Inst. Res. Agron. 42 bis, rue Bourgone. Paris VIIe p. 356, 1932.

Pethybridge, G[eorge] H[erbert]

Potato diseases. Journ. Min. Agric. **41**(2):125–136, 1934.

The author makes reference to Bechhold's and Erbe's copper method for diagnosing leaf roll, which appears to be useless.

Petre, A. W.

Factors influencing the activity of tobacco mosaic virus preparations. Cont. Boyce Thompson Inst. **7**(1):19–28, 1935.

The author reports improvements on the Vinson & Petre lead precipitation of mosaic virus from tobacco leaves. He believes that succulent plants are more susceptible than non-succulent plants of the same species.

Petri, L[ionello]

Court-noué de la vigne. Deux. Cong. Int. Path. Comp. Paris 1931. II. Compt. Rend. et Comm. p. 441–443, 1931.

A brief résumé.

Rassegna dei casi fitopathologici osservati nel 1933. (Review of phytopathological records noted in 1933). Boll. R. Staz. Pat. Veg. n. s. **14**(1):1–78, 1934.

This report contains information on "court-noué" of the vine, regarded by some authors as caused by a virus and on bitter pit of apple.

Sull'arricciamento (court-noué) della Vite. (On leaf roll (court-noué) of the vine). Boll. R. Staz. Pat. Veg. n. s. **14** (2) : 273–278, 1934.

In this paper the author expresses his opinion that this disease belongs to the virus group, which has been included in the third edition of Sourauer's Handbook of Plant Diseases. He discusses the assertion by Viala and Marsais who stated that the disease is due to the fungus *Pumilus medullae.*

Degenerazione e necrosi del cambio dei Peri e dei Meli nel Trentino e in Alto Adige. (Degeneration and necrosis of the cambium of pear and apple trees in the Trentino and the Upper Adige.) Boll. Stat. at. Veg. Rome, n. s. **14**(3) : 281–326. 1934.

The author does not regard in this paper the disease as caused by a virus, but we include it due to its similarity of symptoms.

Piemeisel, R. L.

Weedy abandoned lands and the weed hosts of the beet leaf hopper. U.S.D.A. Circ. **229,** 23 p., 1932.

This is not a work on virus diseases of plants but is of interest to students on the subject, *Eutettix tenellus* Baker being the insect vector of beet curly top.

Pierce, W[alter] H[oward]

Resistance to common bean mosaic in the Great Northern field bean. Journ. Agric. Res. **49**(2) : 183–188, 1934.

Report of results obtained in field and greenhouse tests with certain strains of beans. The results showed that nine strains are immune to common bean mosaic and some tolerant to yellow bean mosaic.

Pittman, H. A.

Virus diseases of plants. With particular reference to the spotted or bronzy wilt disease of tomatoes. Journ. Dept. Agric. Western Australia ser. 2., **11**(1) : 123–140, 1934.

This paper deals almost entirely with spotted wilt of tomatoes. Gives the behavior of a considerable number of hosts to this disease. Gives detailed account of symptoms.

Plantegen, María H. J.

Pathologische verandenrigen in het phoem. (Pathological disturbance in the phloem.) Thesis Univ. Utrecht (Hollandbrukkery, Baarn). 108 p., 1932.

In this rather extensive paper the author reports his experimental studies on the transmission and inoculations in regard to the phloem necrosis disease of coffee in Surinam.

Porter, D. R.

Infectious nature of potato calico. Hilgardia **6**(9) : 277–294, 1934.

This appears to be a virus disease. The author gives a history of the disease, symptoms, results of field and experimental studies, methods of transmission and effects on yield.

Relation of virus diseases to potato production in California. California Agric. Expt. Sta. Bull. **587**, 32 p., 1935.

This bulletin gives a discussion of some of these diseases, including variations of symptoms, tuber indexing, rate of spread in the field, insects in relation to spread, varietal resistance and yield. The yield of table stock may be reduced 25 per cent.

Price, W[illiam] C[onway]

Isolation and study of some yellow strains of cucumber mosaic. Phytopathology **24**(7) : 743–761, 1934.

Descriptions of the behavior on different hosts of some yellow strains of cucumber mosaic and method of isolation.

Purdy Beale, Helen

Specificity of the precipitin reaction in tobacco mosaic disease. Journ. Expt. Med. **54** : 463–473, 1931.

The serum reactions as an aid in the study of filterable virus of plants. Contrb. Boyce Thom. Inst. **6**(3) : 407–435, 1934.

The author reviews the literature on the subject and a precipitin reaction is recommended as a qualitative test in the detection of masked carriers in the identification of new hosts and in the classification of plant viruses. The reaction of several hosts is given to precipitin reaction with antiserum of tobacco virus I. A method for determining the antigenic content of saline extracts of tobacco virus I to which phenol has been added is described in detail. Description is given of Holmes' method of estimating local lesions. Results are given in using different numbers of plants (*Nicotiana glutinosa* L.) inoculated with known concentrations. A quantitative relation is shown to exist between the antigenic content and the active virus concentration of 11 separate extracts compared in eight different pairs. In seven out of eight of these pairs the concentration of virus probably does not vary more than 80 per cent and in the case of the remaining pairs, probably less than 50 per cent. The value of serologic technique is discussed.

Quanjer, H[endrick] M[arius]

De stand der onderzoekingen over de bladrolziekte, de mosaickziekte, de Krinke en de stippelstreepziekte. (The point reached by the investigations into the leafroll disease, the mosaic disease, the crinkle disease and the speckle streak disease.) Voordracten Eersten Aardappeldag Cent. Com. Inzake Kenring v. Gewassen, Wageningen p. 9–19, 1922.

(Influence of fertilization on the health of the potato.) Die Ernährung der Pflanze, **25**: 194–198, 1929.

A general paper with some attention to virus diseases.

De selectie van den aardappel en de invloed van nitwedige omstandigheden, speciaal van de bemesting, op het resultaat daarvan. Voordr. 4 Aardappeldag Centr. Com. Kenr. Gewass., Gehoud. Wageningen. (Selection of potatoes and the influence of external circumstances, especially of fertilizing on the results of this selection.) Lecture **4**, Potato day Central Committee for testing—Wageningen.) July 1930, p. 542–552, 1930.

Results of studies on virus diseases in which fertilizers were tested for control. The insect *Myzus persicae* does not thrive in certain regions.

---, **Thung, T. H., Elze, D[avid] L[eon], & Likhité, V.**

De virusziekten der planten. (Virus diseases of plants.) Landbouw. Tijdschr. der Ver. van Landbuw. **5**(10): 793–836, 1930.

A rather extensive and very comprehensive account of virus diseases of plants with the following chapters: 1. Introduction. Agricultural and scientific importance of the virus diseases. Definition of virus diseases. Bacteriophagy. 2. Genetical research. Deterioration of potatoes and sugar cane. Infection and variability of host plants and parasites. Asexual propagation of cultivated and wild plants, mosses and fungi. Breeding resistant varieties of sugar cane and potatoes. 3. Ecological methods. "Running out" or "Abbau". Importation of sugar-cane cutting from mountain regions. Importation of seed potatoes. Mycorrhiza-hypothesis. Climate and manure. 4. Grafting and budding as methods of investigations. Infectious chlorosis of ornamental plants. "Peach yellows". Comparison of viruses of potatoes, sandal tree and hop. Quercinatype of *Datura stramonium*. 5. Bacteriological, mycological and helminthological methods. Sereh disease of sugar cane. "Curly top" of sugar beet. Leafroll of potato. Tobacco mosaic. 6. Enzymatic methods. Investigations of the juice of mosaic tobacco and other plants. Comparison of different

mosaic diseases. Physiological investigations of leafroll and healthy potato plants. Spread of virus thru different manured plants. 7. Morphological and histological methods. Four types: 1. Chlorosis, (Phloemnecrosis, gummosis of vascular bundles, Pseudo-virus diseases); 2. Mosaic diseases; 3. Necrotical diseases. 4. Alloiophylly. 8. Methods of pure culture of virus diseases and of virus free plants. Sources of infection. Masking of symptoms. Varietal response to infection. "Carriers". Increase of virulence. Source of infection among related and non-related plants. Vira of tobacco and cucumber mosaic. Decrease of virulence. Specialization of mosaic vira. Polyphagy of mosaic vira. Separation of vira. 9. Advantages of tobacco mosaic in virus studies. Filtration and centrifugation. Fractional filtration. Quantitative determination. Chemical purification. Laboratory culture experiments. Corpuscular nature of vira. 10. Cytological methods. Iwanoski's results. Vacuolate bodies. Crystalloid material. Flagellate bodies. Elytrosomes and scolecosomes. Other cytological investigations. Vacuolate bodies as probable cause. 11. The importance of insects in the spread of viruses. Classification of the virus diseases according to their transmission by insects. Diseases which are spread by a specific insect. Diseases which are spread by different insects. Relation between insects, virus and plants. Spread by other animals than insects. Control. 12. Control of virus diseases. Empirical methods. Exclusion, protection, eradication, curing, immunization. Control of potato viruses. Other plants. 13. Bacteriophagy.

La sélection des pommes de terre et l'influence des conditions extérieuses, des engrais en particulier. (Selection of potatoes and the influence of external conditions, in particular the fertilizer.) Bull. Soc. Intern. des Sélectionneurs de Plantes de Grande Culture **3**(2): 1–9, 1930.

This paper has been cited on page 302 of the Bibliography. It appears also in Ernährung der Pflanze **27**(1): 1–8, 1931.

Die Auslese der Kartoffeln und der Einfluss der ausseren Umstande, insbesonderes der Dungung. Landw. Tijdschr., **42,** 542 p., 1930. (Zeitschr. Pflanzenkr. (Abstract) **42**: 491, 1932.)

Pflanzenpathologie auf anatomisch-physiologischer Grundlage. (Plant pathology on the anatomical-physiological foundation.) Angew. Bot. Zeitschr. Erfs. Nutzpflanzen **6**(2): 225–237, 1934.

A general discussion on the subject but especially virus diseases of plants, reviewing the work so far.

Enpele kenmerken der "vegelings"—Ziekte van suiker— en volderbielen ter onderscheiding van de "Zwarte horlvaten" —Ziekte (Some symptoms of the "yellowing" disease of sugar and fodder-beet, and its differentiation from the "black wood-vessel" disease of these hosts.) Tijdschr. Plantens. **40**(10) : 201–214, 1934.

This disease resembles a virus disease but there is no absolute proof that it belongs in that group.

Rademacher, B.

Erfahrungen über die wichtigsten Krankheiten der Ackerbohne und ihre Bekäfflpfung. (Experimental observations on the most important broad bean diseases and their control). Deutsch. Landw. Presse **41**(21) : 253–254, (22) : 275–276, (23) : 290, 1934.

The author summarized briefly Boning's investigations on mosaic of *Vicia faba*.

Rangel, E[ugenio] S[antos]

O mosaic (Mosaic). A Lavoura, (Brazil) **31**(8) : 589, 1927.

Rankin, W[illiam] H[oward]

Leaf curl mosaic or yellows of the cultivated red raspberry. Canada Dept. Agric. Interim Rpt. Dom. Botanist **1921–22**: 30–60, 1922.

Ravaz, L.

Sur la chlorose, Progr. Agr. & Vite. **103** : 366–369, 1935.

Rawlins, T[homas] E[lsworth]

Cytology and other studies of curly-top disease of the sugar beet. Unpublished thesis in the Library of the University of California, 1926.

---------- **& Parker, K. G.**

Influence of rootstocks on the susceptibility of sweet cherry to the buckskin disease. Phytopathology **24**(9) : 1029–1031, 1934.

Reports the author's observations of the effect of the so-called "buckskin" disease of the sweet cherry in grafting experiments.

----------, **& Tompkins, C[hristian] M[ilton]**

The use of carborundum as an abrasive in plant-virus inoculations. Phytopathology (Abstract) **24**(10) : 1147, 1934.

Read, W. H.

Physiological investigations of mosaic disease of the tomato. Cheshunt Expt. and Res. Stat. Herkfordshire. 19th Ann. Rpt., **1933**: 64–67, 1934.

Account of analysis of samples of healthy and artificially inoculated with aucuba mosaic tomato plants, taken at intervals of two hours throughout a period of twenty-four hours.

Reddick, Donald

Some diseases of wild potatoes in Mexico. Phytopathology **22** (6): 609–612. 1932.

This paper includes a brief statement concerning virus diseases. No virus diseases were observed in the wild plants in Mexico. Wild mosaic developed on two wild and three cultivated plants when grown in Ithaca, New York. Leafroll was not seen in Mexico. However, the author reported a spot disease which was found later to be due to a virus.

Reed, H[oward] S[prague] & Dufrénoy, Jean

Éffects de l'affection dite "mottle leaf" sur la structure cellulaire des citrus. (Effect of the so called "mottle leaf" disease on the cytological structure of citrus trees.) Rev. Gen. Bot. **46**(541): 33–44, 1934.

As the title implies this is a study of the cytological structure of diseased citrus leaves, but we include it on account of its similarity to a virus disease.

---------- ----------

Modification in cell structure accompanying mottle leaf of the orange. Amer. Journ. of Botany **22**(3): 311–328, 1935.

This is not a virus disease but is of interest to students of virus diseases because of its resemblance to them.

Reinmuth, E[rnest] Friedrich & Finkembrink, W.

Experimentelles zur Frage der Eisenfleckigkeit der Kartoffel. (Experimental tests concerning the iron spot disease of the potato.) Ztschr. Pflanz. **43**: 21–28, 1933.

Riha, J., & Blattny, C[tibor Eugen Marie Karel]

Ergenbnisse des Versuches über das Verbreiten der Viruskrankheiten und der durch sie verursachten Degeneration der Kartoffeln in verschiedenen Gegenden der Tchechreslovakischen Republik in den Jahren 1926–29. Ochrana Rostlin **9**: 97–108, 1929.

Rischkov, V[itolij L.]

Einige neue wildwachsende buntblättrige Pflanzen. (Some new variegated wild plants.) Biol. Zentralbl. **47**(1) : 18–25, 1927.

A discussion of chlorophyll in chlorotic leaves.

----------, **& Karatschevsky, I. K.**

Chlorophyllmangel und Eunzymwskung. I. Katalasewirkung bei Panaschierung und mosaikkrankheit. Biologie der Pflanzen. **20** : 199–220, 1933.

---------- & ----------

Ueber die Entstehung von "Fern-Leaf" bei Tomaten. (On the origin of "fern-leaf" in tomatoes.) Phytopath. Zeitschr. **7** (3) : 231–244, 1934.

Report of results obtained in inoculation experiments.

----------, & ----------

(Experiments on the artificial transmission of virus diseases of the tomato. In Virus diseases of plants in the Crimea and Ukraine.) State Publ. Office for Crimea, Simferopol p. 7–30, 1934.

This paper contains a brief enumeration of virus diseases of tomato which have been hitherto described as *Stolbur* disease of tomatoes attributed to a virus and wide-spread in Crimea and also observed in Ukraine. Discussion on transmission and inoculation experiments.

(Filterable virus as a cause of virescence of flowers. In Virus diseases of plants in the Crimea and the Ukraine.) State Publ. Office for the Crimea, Simferopol, p. 59–73, 1934.

This paper is a continuation of the preceeding one. It contains a detailed account of the teratological changes observed by the author in tomato flowers naturally or experimentally infected with "woodiness virus". The observations of the author agrees with those of Bald and Eardley. He believes that the Crimean disease is identical to that of Australia known as "big bud". They also agree with those described by Kostoff.

----------, **Karatschevsky, I. K.**

Die Viruskrankheiten der Tomaten und ihre experimentelle Übertragung. (Tomato virus diseases and its experimental transmission.) In his Viruskrankheiten der pflanzn in der Krim und Ukrania. Forshungs Inst. der Krim & Inst. f. Pflanzensch. der Ukraine, Krimisdat, p. 7–30, 1934.

Report of experimental transmission of tomato virus.

Ultravirus als die Ursache der Vergrünungsercheinungen. In his, Virus Krankheiten der pflanzn in der Krim nud Ukraina. Forshungs Inst. der Krim & Inst. f. Pflanzensch. der Ukraine, Krimisdat, p. 59–73, 1934.

---, & **Mikhailova, P. V.**

(On the nature of *Pseudocommunis* sp. In Virus diseases of plants in the Crimea and Ukraine.) State Publ. Office for the Crimea, Simferopol p. 114–121, 1934.

The authors report their cytological studies of potato tubers from plants affected with virus disease. As a result of their observations they found constantly intracellular bodies which appeared to be identical to Debray's *Pseudocommunis vitis*. They state that microchemical tests showed that these bodies are a product of cell metabolism under the influence of certain pathological processes.

---, & ---

Über die Natur d. *Pseudocommunis* sp. (On the nature of *Pseudocommunis* sp.) In his Viruskrankheiten der pflanz in der Krim und Ukraine. Forshungs Inst. der Krim & Inst. f. Pflanzensch. der Ukraina, Krimisdat, p. 114–121, 1934.

See preceding citation by the same authors.

Robbins, William J.

Isolation of the infective principle of virus diseases. Science **80**(2073) : 275–276, 1934.

The author reviews briefly the outstanding works of other investigators on the subject and based on assumptions establishes a table given the dilution and quantity of juice taken 100,000 the molecular weight of the infective material.

Roberts, J. I.

The tobacco capsid (*Egytatus* volucer, Kirk.) Rhodesia. Bull. Ent. Res. **21** : 169, 1930.

Refers to crinkle which is probably the same as "Kroepoek".

Rosa, J[oseph] T[ooker]

Relation of potato viroses to yield. Amer. Pot. Journ. **5**(7) : 190–191, 1928.

Popular. Contains interesting data on yields.

Rosenfeld, A[rthur] H[inton]

Variedades de caña de azúcar inmunes o muy resistentes al

mosaico. (Sugar-cane varieties immune or very resistant to mosaic.) La Hacienda **25**: 489–490, 1930.

Popular.

Ross, A. F.

The effect of proteoclastic enzymes on purified preparations of tobacco mosaic virus. Phytopathology (Abstract) **25**(1): 33, 1935.

Ruhland, W., & Wetzel, K.

Zur Physiologie der sogenannte Blattrollkrankheit der Kartoffelpflanze. (On the physiology of the so-called leafroll disease of the potato plant.) Ber. Verhandl. Sachs. Akad. Wiss. Leipzig. Math.–Phys. Kl. **85**(3): 141–149, 1933.

It was found in leaves and tubers, dormant and sprouted potato, a higher content of dextrin and a lower content of sugar in leaf-rolled as compared to healthy ones. It was also found that in diseased material the diastatic activity was much lower, accompanied by a reduction of assimilatory capacity, transpiration and respiration.

Salaman, R[edcliffe] N[athan]

Discussion on "Ultra-Microcospic" Viruses. Proceedings of the Royal Soc. **B 104**: 550–552, 1929.

The author states that he has repeated the work of Johnson and Schultz in which the former inoculated tobacco with juice from apparently healthy potatoes and the latter inoculated potatoes with juice from apparently healthy potatoes. His results were negative. He called attention to the tolerance of host varieties, to attenuation of virus and varietal reaction in host plants.

---------- & Bawden, F. C.

An analysis of some necrotic virus diseases of the potato. Proc. Roy, Soc. ser. **B. 111**: 53–73, 1932.

Summary:

"A summary of the literature on streak is given, from which it appears that two distinct chemical states can be isolated.

"One of these is that described by Orton and commonly known as stipple-streak or leaf-drop streak and later designated, on the grounds of its histopathology, as acropetal necrosis. The other, known as top-necrosis, has been described by Quanjer on basis of its histopathology as acronecrosis.

"It has been shown that the former is the distinctive reaction in certain varieties of the Y virus of Kenneth Smith.

"Acronecrotic or top-necrosis has been shown to be divisible into at least four distinct groups based on its varietal reaction, and here designated as top-necrosis X, top-necrosis A, top-necrosis B, and top-necrosis C.

"The first three are alike in that when they do produce a top-necrosis in any given variety, it is unaccompanied by any mosaic symptom. Top-necrosis C, on the other hand, differs clinically by the fact that necrotic and mosaic symptoms occur together.

"It is shown that top-necrosis X is due to the action of the virus acting alone.

"Top-necrosis A is shown to be due to a complex containing X and Y, possibly associated with the virus X.

"Top-necrosis B is shown to be due to a complex containing both Z and Y.

"Top-necrosis C is likewise shown to be due to the presence of both viruses X and Y.

"Top-necrosis X and C complexes are capable of transmission by needle inoculation to other potato varieties, though it by no means follows that the resultant lesion is a top-necrosis. Top-necrosis B is uninoculable, and so is top-necrosis A, except that it can be conveyed to the varieties Arran Crest and Epicure by the needle.

"Carriers of top-necrosis A are found among many of our widest grown varieties ——."

---------- **& Hurst, C. C.**

Discussion on the microscopy of the filterable viruses. Journ. Roy. Micros. Soc. **52**: 237–238, 1932. (Univ. Cambridge School Agric. Mem. (Abstract) **5**: 30–31, 1933).

A brief discussion of the inclusion bodies in varieties of potatoes infected with different viruses.

Virus disease research in relation to the cultivation of the potato. Hort. Educ. Assoc. Year Book. D. (Wye. Kent) **2**: 45–50, 1933.

Popular.

The raising of blight-resistant varieties and virus-free stocks. 4 pp. (No place of publication or date on the copy in the hands of the compiler.) (1934).

Explains the efforts of the Potato Virus Research Station made to secure varieties free from virus diseases.

Sandford, G. B.

A malady of the potato in Alberta similar to psyllid yellows. Sci. Agric. **15**(1): 46–48, 1934.

Not a virus disease paper, but may be of interest due to similarity of symptoms.

Sarker, B. N., & Dutt, K. M.

Effect of mosaic disease on the tonnage and the juice of sugar cane in Patna. Indian Journ. Agric. Sci. **4**(5) : 797–802, 1934.

Summarized results of yields for 1933–34, following the same scope of Mc Rae and Sulvia-Maniam for previous years.

Sarrant, A.

(Sugar-cane Fiji disease.) Journ. Sta. Agron. Guadalupe **1** (4) : 116–119, 1921.

Brief discussion on the spread of Fiji disease of sugar cane and the legislative action taken in Guadalupe to prevent its introduction.

Savastano, Giulio

El mosaico del fagiolo in Italia. (Bean mosaic in Italy.) Boll. R. Staz. Patol. Veg. **12**(4) : 377–394, 1932.

Popular notes on the distribution, symptoms, description of the disease, varietal resistance to mosaic and control.

Schapiro, S. M.

Die Federblätter der *Lappa*. (The feather-leaf of *Lappa* sp.) In Rischkov, V. Viruskrankheit der pflanzn in der Krim und Ukraina. Forshungs Inst. der Krim & Inst. f. Pflanzensch der Ukraine, Krimisdat, p. 100–113, 1934.

Brief account of this disease attributed to a virus.

(Note: This article is cited under Schapiro, S. M. page 204.)

Schreven, D. A. van

Kalkgebrek als oorzaak van mergnecrose bij Aardappelpnollen (Lime deficiency as the cause of medullary necrosis of potato tubers.) Tijdschr. over Plantenziekten **40**(11) : 225–255, 1934.

Although this paper is not a discussion on virus diseases specially, it contains comparison on necrosis due to virus and other valuable data on the subject.

Schultz, E[ugene] S[chultz]

Potato diseases of virus group due to cause not yet known. U.S.D.A. Yearbook **1927** : 522–525, 1928.

Brief popular notes on potato virus diseases. Gives a list of the types of virus diseases, means of dissemination and experiments in control.

---------- **& Folsom, Donald**

Recent potato virus-disease information contributing to the production of better seed potatoes. 15 Annual meeting of the Potato Association of America pp. 203–227, 1928.

A popular review.

----------, **Bonde, Reiner, & Raleigh, W. P.**

Isolated tuber unit seed plots for the control of potato virus diseases and blackleg in northern Maine. Maine Agric. Expt. Sta. Bull. **370**, 32 p. 1934.

A thorough account of experiments on the control of virus diseases in potatoes.

Schweizer, J.

Verslag over het jaar 1925 door Dr. W. H. Arisz. Med. Bevekisch Proefstation **41**: 14. 1926.

Refers to "Kroepolk" and "Krepoli" which are probable same as "Kroepoek".

Serrano, Luis A.

Mosaico. (Mosaic). Imp. Bolívar, Caracas (Venezuela) 16 p., 1927.

Report of a trip of inspection made by the author to the northern sugar-cane region of Venezuela. It includes a brief review of the work of others and a short historical sketch. Gives symptoms and methods of control and prevention.

Servazzi, O.

Nota sulla classificazione isto-pathologica del Quanjer, deller virosi nolla potato. (Notes on the histo-pathological classification of Quanjer of the virus of the potato.) Defesa Pianta Torino **27**: 20–24, 1932.

Severin, Henry H[erman] P[aul] & Thomas, William W.

Notes on the beet leafhopper, *Eutettix tenella* Baker. Journ. Econ. Ent. **11**(3): 308–312, 1912.

This paper has no mention of virus diseases, but we considered it of interest to students on the subject due to the long list of host plants of the beet leafhopper which is a recognized insect vector of beet-curly top.

Natural enemies of the sugar beet leafhopper in California. California State Comm. Hort. Mo. Bull. **4**(5–6): 277–280, 1915.

Notes on the beet leafhopper *Eutettix tenella* Baker. Journ. Econ. Ent. **11**(3): 308–312, 1918.

Notes on the behavior of the beet leafhopper (*Eutettix tenella* Baker) Journ. Econ. Ent. **12**(4): 303–308, 1919.

Detailed account of his studies. Although not a strictly virus disease paper it is of interest to students of the subject.

The beet leafhopper (*Eutettix tenella* Baker). California Agric. Expt. Sta. Ann. Rpt. **1918–19**: 70, 1919.

Brief notes on migration of this leafhopper and relation of species to beet-curly top disease.

Beet blight. California Agric. Expt. Sta. Ann. Rpt. **1919–20**: 62–63, 1920.

Brief note in relation to the progress of studies on beet-curly top.

The beet leafhopper, *Eutettix tenella* Baker. California Agric. Expt. Sta. Ann. Rpt. **1920–21**: 41–42, 1921.

Brief note reporting field experiments.

Summary of life history of beet leafhopper (*Eutettix tennella* Baker). Journ. Econ. Ent. **15**(5): 433–436, 1921.

Mosaic and curly leaf disease of sugar beets. Journ. Econ. Ent. **15**(3): 247, 1922.

Brief reviews of Robbins' paper (Phytopathology **11**: 349–365) and Bonequet's (Phytopathology **7**(4): 269–289) on mosaic of sugar beets.

Infective beet leafhoppers (*Eutettix tenella* Baker), do not transmit curly leaf daily. Journ. Econ. Ent. **15**(4): 318–1922.

Brief note giving results of experiments.

---, **Schwing, E[dward] A., & Hartung, W. J.**

Relation of leafhopper migrations to time of sugar beet plantings. Use of nicotine dust against the beet leafhopper, *Eutettix tenella* Baker. Curly leaf transmission experiments with leafhopper. California Agric. Expt. Sta. Ann. Rpt. **1921–22**: 83–85, 1922.

Brief notes reporting field experiments and observations.

Natural enemies of beet leafhopper (*Eutettix tenella* Baker). Journ. Econ. Ent. **17**(3): 369–377, 1924.

Although not a virus disease paper it is of interest to workers on the subject.

Natural enemies of beet leafhopper (*Eutettix tenella* Baker); curly leaf transmission experiments; chemical substance or toxin; incubation period. California Agric. Expt. Sta. Ann. Rpt. **1923**: 125–128, 1924.

Life-history of beet leafhopper (*Eutettix tenella* Baker) in California. Univ. California Pub. in Ent. **5**(4): 37–88, 1930.

Not a virus disease paper, but of interest to students of the subject.

----------, **& Freitag, Julius H.**

List of ornamental flowering plants experimentally infected with curly top. U. S. Dept. Agric. Plant Disease Rep. **17** (1): 2–5, 1933.

----------, & ----------

Ornamental flowering plants naturally infected with curly-top and aster-yellows viruses. Hilgardia **8**(8): 233–260, 1934.

"Fourteen species of ornamental flowering plants in 13 genera belonging to 10 families have been found to be naturally infected with curly-top in California. Previously non-infective beet leafhoppers transferred the curly-top virus from the following 14 species of naturally infected plants to sugar beet: *Kochia scoparia* var. *trichophila*, *Celosia argentea* var. *cristata*, *Mirabilis jalapa*, *Dianthus plumarius*, *Pelargonium hortorum*, *Tropaelum majus*, *Viola tricolor* var. *hortensis*, *Viola cornuta*, *Petunia hybrida*, *Scabiosa atropurpurea*, *Zinnia elegans*, *Cosmos bipinnatus*, *Coreopsis tinctorea*, *Helichrysum bracteatum*. The curly-top virus was not recovered from *Dianthus caryophyllus*, *Mathiola incana*, *M. incana* var. *annua*, although showed reliable symptoms of the disease."

"Eight species and 3 varieties of ornamental flowering plants in 7 genera belonging to 4 families have been found to be naturally infected with aster yellows in California. Previously non-infective *Cicadula divisa* transferred the yellows virus from the following species and varieties of naturally infected plants to asters or celery; *Ranunculus asiaticus*, *Eschscholtzia californica*, *Godetia grandiflora*, *Chrysanthemum segetum*, *Zinnia elegans*, Double Giant Pink, Dahlia Flowered mixed, lilliput Scarlet Gem, *Tagetes patula*, I. *erecta* and *Helichrysum bracteatum*."

Weed host range and overwintering of curly-top virus. Hilgardia **8**(8): 261–280, 1934.

"The weeds growing on the uncultivated plains and foot hills and in the cultivated areas infected with curly top include 57 species in 28 genera belonging to 16 families. The wild plants growing on the uncultivated area demonstrated to be naturally infected with curly

top include 14 species in 13 genera belonging to 8 families. In the cultivated areas 26 species of weeds in 15 genera belonging to 9 families were found to be naturally infected with the disease. The curly top overwinters in 11 species of annuals and 3 species of perennial wild plants growing on the uncultivated area. Previously non-infective beet leafhoppers repeatedly recovered the virus from a perennial-ballscale (*Atriplex fruticulosa*) during a period of six months, when the tests were discontinued. Four species of perennials and 3 species of weeds sometimes annuals and sometimes perennial, growing in the cultivated area were demonstrated to be naturally infected with curly top.'' Several economic plants overwinter and were naturally infected with curly top. A list is given of ornamental flowering plants which were infected with curly top under natural conditions. The curly-top virus rarely overwinters in the male beet leafhopper since most of the males die during the winter.

Transmission of California aster and celery-yellows virus by three species of leafhoppers. Hligardia **8**(10) : 339–361, 1934.

Detailed account of experimental studies in aster and celery yellows virus transmission by means of *Cicadula divisa, Thamnotettix montanus* and T. *germinatus.*

---, **& Haasis, Frank A.**

Transmission of California aster yellows to potato by *Cicadulla divisa.* Hilgardia **8**(10) : 329–335, 1934.

Report of experiments on transmission of aster yellows to potato by means of *Cicadula divisa* Uhl. (*Cicadula sexnotata* Fall.) Description of symptoms, incubation period and behavior of the virus is given.

Expriments with aster-yellows virus from several states. Hilgardia **8**(10) : 305–325, 1934.

After a brief review of the work of other investigators of the subject the author gives the results of his studies based on experimental data.

Shapiro, S. M.

(A curious case of mosaic in *Lappa* sp. In virus diseases of plants in the Crimea and the Ukraine.) State Publ. Office for the Crimea, Simporopol p. 109–113, 1934.

A brief morphological account reporting a ''fern leaf'' disorder of a species of *Lappa.* It is the first record in weeds of such malformation of leaves. It is attributed to a virus.

Shapovalov, Michael

Some host response in graft transmissions of dieback streak of tomatoes. Phytopathology (Abstract) **24**(10) : 1149, 1934.

Chemical splitting of the tomato "Combination-Streak" virus complex. Phytopathology (Abstract) **25**(1) : 33, 1935.

Sheffield, Frances M[arion] L[ena]

Intracellular inclusions in plant virus diseases. Deux Cong. Int. Path. Comp. Paris, 1931. II. Compt. Rend. et Comm. p. 481–482, 1931.

A brief note.

Experiments bearing on the nature of intracellular inclusions in plant virus diseases. Ann. App. Biol. **21**(3) : 430–453, 1934.

The author gives the results of a series of experiments and concludes that it is not possible to identify virus diseases by cytological characters alone but states that they are of value in supplementing other diagnostic features. She compares the behavior of these bodies produced by aucuba mosaic, Hy III and tobacco mosaic viruses. She also gives the results of treatments with chemicals and states that lactic acid induced the formation of amoeboid bodies like the X-bodies of tobacco mosaic but that they persisted for only a few hours.

Shull, J. M.

Comments on the "breaking" of tulips. Florida Growers **21**: 164, 1934.

Silberschmidt, Karl

Neuere Arbeiten über die Strömungsrichtung der Assimilate im Pflanzenstengel. (New works on the assimilation stream direction within the plant stem.) Die Naturwissenschaften Heft 43, **26**(10) : 725–728, 1934.

A review of recent literature.

Simmonds, J[ohn] H[oward]

The spotted wilt of tomatoes. Queensland, Australia, Dept. Agric. & Stock, Phytopathological Leaflet **1,** 3 p., 1927.

A brief, popular discussion of symptoms, cause and control.

Bunchy top of the banana and its control. Queensland, Australia, Dept. Agric. & Stock. Advisory Leaflet 12, 4 p., 1934.

A popular discussion of symptoms and control.

Slate Jr., Wm. L.

Comparative analysis of healthy and of diseased (calicoed) leaves of tobacco, tomato and petunias. Connecticut Agric. Expt. Sta. Bull. **258**: 372–373, 1924.

Brief note giving the analysis.

Smith, E. H.

Spotted wilt disease of tomatoes. Expt. & Res. Stat. Cheshunt, Circ. **7**, 1933. (Gard. Chron. **94**: 350, 1933.)

Smith, J[ohn] Henderson

Remarks on the size of plant viruses. Arch. für Exper. Zellforsch. **15**(2–4): 454–456, 1934.

The author discusses the "living entity" theory in regard to plant viruses. He is opposed to the term "living".

Smith, Kenneth M[anley]

Discussion of "Ultra-Microscopic Viruses". Proc. Royal Soc. **B 104**: 545–546, 1929.

The author raises the question as to whether the insect is an obligate alternate host for the virus or merely a mechanical carrier. He summarizes his discussion by saying: "By certain manipulation of the virus of potato mosaic, it is therefore possible to change its character and, by starting with the normal mild disease upon potato, to produce from it four apparently distinct diseases, or perhaps it would be more accurate to say, four distinct modifications of the virus, i. e., (1) ringspot of tobacco; (2) its highly infections counterpart in potato; (3) the virulent form of disease in tobacco; (4) the aphis produced green line disease, also in tobacco."

Nature complexe de certain virus causant des mosaïques de la pomme de terre. (The complex nature of certain virus causing mosaic of potato.) Compt. Rend. 2 Congr. Intern. Pathol. Comp. Paris, **2**: 443–445, 1931.

---, **& Dufrénoy, Jean**

Botanique—Sur le virus Y des Solanées. (Botany—On virus Y of Solanaceae.) Proc. Roy. Soc. **B. 109**: 241–267, 1931. (Compt. Rend. Acad. Sci. (Abstract) **199**(21): 1147–1150, 1934.)

This annotation is taken from the abstract. The authors report their observations.

Insects in relation to virus diseases of plants. Agricultural Progress **11**: 86–88, 1934.

This is a very brief and clear summary of our knowledge of this subject. It is a summary of a paper read at Cambridge, July, 1933.

Some virus diseases of the potato and other farm crops. The Scottish Jour. Agric. **16**(4) 11, 1933.

After a brief discussion of the subject, the author gives a discussion of the mechanics of aphid attacks on plants. This is followed by a popular discussion of the virus diseases of the potato and of the potato viruses.

Some aspects of the plant virus problem. Agricultural Progress **11**: 88–92, 1934.

A very brief but very clear review of the most important aspect of these problems. It is a summary of a paper read in London, December, 1933.

The plant virus in the insect vector. Arch. für Exper. Zellforsch. **15**(2–4): 459, 1934.

The author states there are three kinds of relationship between plant viruses and the insect vectors: (1) purely mechanical, (2) semi or group specific and (3) specific. Other valuable data on the relations of the viruses and insect vectors are given.

The mosaic disease of sugar-beet and related plants. Journ. Minis. Agri. **41**(3): 269–274, 1934.

This is a popular paper giving a brief history of the disease, symptoms, weed hosts, method of spread and effects on the yield.

Report of experimental observations of the virus Y on Solanaceae, especially the tobacco plant.

A virus disease of *Primula obconica* and related plants. Ann. Appl. Biol. **22**(2): 236–238, 1935.

A description of the disease and the results of inoculating the virus into other species of plants.

A virus disease of cultivated crucifers. Ann. Appl. Biol. **22** (2): 239–242, 1935.

A description of the disease and the results of inoculations into other species of plants. The virus is usually fatal on *Nicotiana glutinosa*.

Smith, Loren B[arlett]

Notes on spinach breeding. Proc. for 1920. Amer. Soc. Hort. Sci. **17**: 146–155. 1921.

Report of experiments and methods in the production of a mosaic-resistant spinach at the Virginia Truck Experiment Station.

Smith, M. Smith

(The plant virus in the insect vector.) Archiv fur Exper. Zellforsch. Band **15**: 459, 1934.

A very brief review of the subject in which the author divided the insect vectors into three groups: (1) those in which the insect is a mechanical vector, (2) in which an insect species has a particular affinity for a plant virus which may be transmitted by other insect vectors, (3) in which a specific insect alone is able to transmit a definite virus.

Sommer, H.

Nochmals: Mosaikkrankheit an Kakteen. Gartenwelt **37**: 37-38, 1933.

Sorauer, Paul [Carl Moritz]

Handbuch der Pflanzenkrankheiten (Die nicht parasitaren Krankheiten). vol. I: 893–964, 1924.

Much of this is devoted to a review of virus diseases.

Soukhoff, K. S.

(Contribution to the physico-chemical characterization of the filterable viruses of mosaic. In Virus diseases of plants in the Crimea and Ukraina.) State Publ. Office for the Crimea, Simferopol, p. 31–38, 1934.

Report of experiments and observations made by the author in relation to the virus of tomato "fern-leaf" disease. The properties of the virus observed by the author do not agree with those reported by Mogendorff in which it was suggested that the virus under study belongs to the cucumber-mosaic group. The author believes that it belongs to the tobacco-mosaic group. Other valuable data is given.

Spencer, Ernest L.

Influence of nutrition on host susceptibility to yellow tobacco mosaic. Phytopathology (Abstract) **25**(1): 33, 1935.

Effect of nitrogen supply on host susceptibility to virus infection. Phytopathology **25**(2): 178–189, 1935.

A study was made of the effect of nitrogen supply on host susceptibility to virus infection. Turkish tobacco, *Nicotiana glutinosa* L. and Early Cluster beans were the hosts under study with different types of mosaic virus. The results of the study showed that

there is a definite correlation between host nutrition and host susceptibility to virus infection and that host susceptibility is not governed mainly by host vigor as judged by the rate of growth, but by some limiting factor the nature of which is at present unknown. Data are given as to rate of growth and host susceptibility in relation to nitrogen level.

Influence of phosphorus and potassium supply on host susceptibility to yellow tobacco mosaic infection. Phytopath. **25** (5) : 493–502, 1935.

The author says: "Data obtained in this investigation indicate that phosphorus supply has apparently only an indirect influence on susceptibility of tobacco to infection with yellow tobacco mosaic, inasmuch as susceptibility and growth seem to be correlated directly with each other. Potassium supply, on the other hand, has a direct influence in changing host susceptibility to infection. When more than 20 milligrams of potassium were added each day, susceptibility decreased appreciably. Growth, however, was not retarded noticeably until the addition of potassium exceeded 200 milligrams per day."

Sreenivasay, M.

Spike disease of sandal. Quinquennial survey investigations. Perfum. & Essent. Oil. Rec. **24**: 265–266, 1933.

---, **& Rangaswami, S.**

Field studies on the spike-disease of sandal (*Santalum album* Linne) I. Observations on the natural dissemination of spike. Proc. Indian Acad. Sci. **1**(B) : 143–154, 1934. (Mem. Indian Inst. Sci. No. 26, 1934.)

The fact that spike disease of sandal discontinued its natural spread and that it occurred in isolated outbreaks in areas far from all source of infection lead the authors to believe in virus-infected seeds and long-distance dispersal of the insect vectors effected by wind or other agencies. Data of observation on this subject is given in this paper.

Contribution to the spike-disease of sandal. (*Santalum album* L.) LXVIII, Hydrogen-ion concentration and buffering capacity as factor of disease resistance. Journ. Indian Inst. Sci. **17A** : 153–164, 1934.

Insect transmission of spike disease. Nature **133**(3358) : 382, 1934.

The author makes the distinction of the stunting non-infectious condition of sandal due to environmental conditions and the highly infectious spike disease. Gives details of transmission experiment and positive results obtained in tests using *Moonia albimaculata*.

Ssuchov, K.

Material zur phsico-chemischen charakteristik des filtrierbaren Viruses. In Rsichkov, V. L. Viruskrankheiten der pflanzn in der Krim und Ukraine Forshungs Inst. der Krim & Inst. f. Pflanzensch. der Ukraine, Krimisdat, p. 31–38, 1934.

Investigations on the filtration of viruses and the different filters in use.

----------, & **Lanschina, M. N.**

Pathologische Veräderungen in den pflanlichen Zellen der Kj. Wirkung und das Problem der X–Körper. In Rischkov, V. L. Viruskrankheiten der pflanz in der Krim und Ukraina. Forshungs Inst. der Krim & Inst. f. Pflanzensch der Ukraine, Krimisdat, p. 122–124, 1934.

Stahel, Gerold

De tegenwoordige stand van het onderzoek naar den overdrager der zeefvatenziekte van den Koffie. (The present status of the investigation on the vector of phloem necrosis of coffee). Landbouwproefstat. Suriname Meded. **7**, 9 p., 1934.

The author reports his observations in connection with his experiments on the insect vector of phloem necrosis of coffee. He observed that *Lincus* bugs may be concerned in the process of transmission.

Stanley, W. M.

The action of high frequency sound waves on tobacco mosaic virus. Science **80**(2076) : 339–341, 1934.

The author reviews Takahashi and Christensen's work (Science 79: 415, 1934). Describes his experiment and concludes: "The results indicate that inactivation of virus by supertonic radiation is associated with cavitation of dissolved gas and with the presence of extraneous matter found in untreated juice, since high frequency sound waves of great intensity have practically no effect on purified virus under a high vacuum."

Chemical studies on the virus of tobacco mosaic. I. Some effects of trypsin. Phytopathology **24**(10) : 1055–1085, 1934.

Discussion, based on experiments, on the decrease of infectivity of different viruses and the reaction on different hosts in relation to the interference of trypsin.

Chemical studies on the virus of tobacco mosaic. II. The pro-

teolytic action of pepsin. Phytopathology **24**(11):1269–1289, 1934.

The author reports his studies and observations and concludes that since pepsin inactivates virus only under conditions favorable for proteolytic activity and since the rate of inactivation of virus varies directly with the concentration of active pepsin, it is concluded that the inactivation of virus is due to the proteolytic action of pepsin. This suggests the virus of tobacco mosaic is a protein, or very closely associated with a protein, which may be hydrolyzed with pepsin.

Chemical studies on the virus of tobacco mosaic III. Rates of inactivation at different hydrogen-ion concentrations. Phytopathology **25**(5):475–492, 1935.

This paper is a record of experimental studies of tobacco-mosaic with reference to temperature hydrogen-ion concentration and virulence on *Nicotiana tobacum, Nicotiana glutinosa* and *Phaseolus vulgaris*. Experiments with tobacco ring spot and cucumber-mosaic viruses showed that they were less stable than tobacco-mosaic virus.

Stevens, Neil E.

An attempted analysis of the economic effects of cranberry diseases. U. S. Dept. of Agric. Plant Diseases Reporter **19**(8):112–136, 1935.

Two pages are devoted to the false blossom. It is considered the most serious disease of this crop. It is the cause of heavy losses but may have prevented over production.

Stevenson, F. J., & Clark, C. F.

New potato varieties. Amer. Potato Journ. **11**(4):85–92, 1934.

Report of new potato varieties resistant to disease, among them is the Katahdin potato variety which is resistant to mild mosaic.

Stewart, Fred Carlton

Observations on some degenerate strains of potatoes. New York (Geneva) Agr. Expt. Sta. Bull. **422**:319–357, 1916.

"This is a detailed account of the behavior of a large number of potato plants of known parentage and belonging to degenerate strains of several different varieties. The object of the study was to increase our knowledge of the diseases or forms known as leaf-roll, curly-dwarf, mosaic and spindling-sprout or other forms of degeneration. The conclusion is reached that leaf-roll, curly-dwarf and mosaic are closely related disorders due to the same general, undetermined cause. In some respects they behave like bud varieties; but they present also important points of difference. All are transmitted

through the seed tuber. The progeny of affected plants almost invariable become affected. The heredity of spindling-sprout is still undetermined and its cause is largely a matter of conjecture. However, it may be stated that spindling-sprout is not correlated to leaf-roll, mosaic or curly-dwarf.''

Stone, R[oland] E[lisha]

Winter blight or streak of tomatoes. Paper read before the Canadian Phytopathological Society, December 1924 (unpublished).

Not seen by the compilers.

Storey, H[arold] H[aydon]

Streak disease of maize. Dept. Agric. Union of South Africa. Reprint from Farming in South Africa, September, 1926.

A short popular discussion of symptoms, spread by an insect and control.

Streak disease of maize. Union of South Africa. Dept. Agric. 8 p., 1926.

Popular.

Streak disease of Uba cane. Jamaica Dept. Agric. Microb. Circ. **6**: 38–39. 1926.

Brief popular notes on streak disease. According to the author the disease is not confined to Uba variety only, but has been found to affect 10 other varieties. The insect vector is the leafhopper *Balclutha mbila* Naud. *Aphis maidis* Fitch has always failed to transmit streak. Methods of control are given.

(Studies on the mechanism of the transmission of plant viruses by insects.) Arch. für Exper. Zellforsch. **15**(2–4): 457–458, 1934.

This is a condensed paper on the same subject by the author noted above.

The photodynamic action of methylene blue on the virus of a plant disease. Ann. Appl. Biol. **21**(4): 588–589, 1934.

Description of experiment by which the author concludes that the virus of a plant disease may be inactivated by exposure to light in the presence of a methylene blue and oxygen as has been done with animal viruses and bacteriophage.

Stout, Gilbert L[eonidas]

Peach yellows. Trans. Illinois State Hort. Soc. **63**: 479–493, 1929.

Popular.

Sukhoff, K. S., & Lanshina, M. N.

(Pathological changes in plant cells caused by the action of potassium iodide. (In connection with the problem of the nature of X bodies.) Preliminary communication. In virus disease of plants in the Crimea and the Ukraine.) State Publ. Office for the Crimea Simforopol p. 122–124, 1934.

The authors report in this paper the finding of X bodies in the cells of healthy beet plants treated with a 0.01 percent solution of potassium iodide. These inclusions resemble very closely those found by other workers in cells of beets affected with virus diseases.

Summers, Eaton M.

Types of mosaic on sugar cane in Louisiana. Phytopathology **24**(9): 1040–1042, 1934. (Sugar News **16**(2): 83–84, 1935.)

The author reports the finding of four distinct types of mosaic which were very different in virulence. One of them was much more virulent than the others and very destructive.

----------., & Rands, R. D.

Losses due to planting of mosaic seed cane. The Sugar Bulletin **13**(15): 2–6, 1935.

This is a very important paper for Louisiana growers of sugar cane. It gives the results of studies on germination and yields of Co 281, Co 290, POJ 234 and C P 29/291. It urges the growers to select mosaic-free cane for planting and gives instructions for doing so.

Sundararaman, S.

The mosaic disease of sugar cane. Madras, Dept. Agric. Leaflet **42**, 2 p., 1926.

Brief popular notes on sugar-cane mosaic.

Takahashi, William N., & Rawlins, T[homas] E[lsworth]

Application of stream double refraction in the identification of streak disease of tomato. Phytopathology **24**(10): 1111–1115, 1934.

The author summarizes: "Juice from streaked tomato plants infected with a combination of tobacco-mosaic and potato latent viruses exhibits a stream double refraction indistinguishable from that exhibited by tomato plants infected with tobacco-mosaic virus alone." "Juice from tomato plants infected with dieback streak exhibits a stream double-refraction behavior which is indistinguishable from

that shown by normal plants. This technic may therefore be used to distinguish diseased plants infected with combination streak from those infected with dieback streak."

----------, ----------

The relation of stream double refraction to tobacco mosaic virus. Science **81**(2099): 299–300, 1935.

This is a continuation of previous studies by the authors. They advance three theories for this phenomenon and summarize the results as follows: "Although much of the evidence cited above favors the supposition that the virus particles are the causal agent of most of the stream double refraction exhibited by juice from mosaic plants the evidence remains inconclusive."

Takata, K[azno]

(Results of experiments with dwarf diseases of rice plant.) Journ. Jap. Agric. Soc. (171): 1–4, 1895, (172): 13–32, 1896.

Thiele, R.

Ein Fall typischer Kräuselkrankheit bei Baumwolle im Gewächshaus. (A case of typical leaf-curl on cotton in the glasshouse.) Zeitschr. Pflanzenkrankh. **23**: 198–201, 1913.

Thompson, A.

Diseases of tobacco in Malaya. Malayan Agric. Journ. **22**(6): 263–269, 1934.

The author describes briefly the symptoms, manner of spread and control of tobacco mosaic.

Thornberry, H. H.

Particle size of three strains of tobacco mosaic virus. Phytopathology (Abstract) **25**(1): 36, 1935.

Thung, T. H.

Epidemiologie de quelques maladies de tabac. (Epidemiology of some diseases of tobacco.) Deux. Cong. Intern. Path. Comp. Paris, 1931. II Compt. Rend. et Comm. p. 482–484, 1931.

A very brief discussion of mosaic and Kroepoek.

Jaarverslag 1 Mei 1930—30 April 1931. Proefst. Vorst. Tabak Meded. **71**: 21, 1931.

De epidemiologie van tabaksziekten. Proefs. Vorst. Tabak. Aug. 1931.

The author gives results of experimental work with mosaic and kroepoek.

Phytopathologische waaremingen. Proefes. Vorst. Tabak. Meded. **76**: 20–25, 1932.

This paper gives the results of experimental studies on mosaic and "kroepoek".

Over enkele tabaksvirusziekten, de dovr insecten worden verbreid. Overgedrukt uit Nerslag 13 e. Bijeenkomst van de Vereeniging van Proefstation-Personeel, 1933.

Phytopathologische waarnemingen. (Phytopathological observations.) Proefstat. Vorst Tabak, Meded. **77**: 34–48, 1934.

Notes on a new form of tobacco mosaic characterized by wispy leaves. Proof also has been obtained, that ordinary mosaic of tobacco is disseminated in the fields by laborers.

Bestrijding der krul—on kroepoekziekten van tabak. (The control of curl and crinkle-diseases of tobacco.) Proefs. Vorst. Tabak. Meded. **78**: 3–18, 1934.

This paper is a continuation of the studies reported in No. 72. The disease is carried by *Bemisia* sp. and attacks the following weed hosts: *Ageratum conyzoides, Synedrella nodiflora* and *Vernonia cinerea.*

Tice, C[ecil]

Seed-potato certification in British Columbia. Potato Magazine **4**(10): 6, 1922.

Rules.

Tims, E[ugene] C[hapel]

Severe type of mosaic on sugar-cane variety. Phytopathology (Abstract) **25**(1): 36, 1935.

Tolas, A. G.

Minnesota certification rules. Potato Mag. **4**(9): 10, 18, 1922.

Rules—Legislation.

Tompkins, C[hritian] M[ilton] & Gardner, Max W[illiam]

Spotted wilt of head lettuce. Phytopathology (Abstract) **24** (10): 1135–1136, 1934.

A destructive virus disease of cauliflower and other crucifers. Phytopathology (Abstract) **24**(10): 1136–1137, 1934.

Breaking in stock (*Manthiola incana*), a virosis. Phytopathology (Abstract) **24**(10) : 1137, 1934.

Torres Filho, A[rthur]

O combate do mosaico da canna de assucar. (The control of sugar cane mosaic.) Brasil Agricol. **12**(2) : 65, 1927.

Recommendation of the planting of resistant varieties.

Tropova, A. T.

A contribution to the diseases of American jute. (Diseases and pests of new cultivated textile plants.) Inst. New Bast Raw Material. Moscow p. 58–60, 1933.

Brief note reporting that *Abutilon avicennae* growing near tobacco plantations showed a disease very similar to reticulate mosaic of tobacco. Investigations are still in progress in the study of this malady.

Trotter, Alessandro

La degenerazione della patata e le malattie da virus. (Potato degeneration and virus diseases.) Richerche, osservazioni ed divulgazioni Fitopatologiche per la Campania ed il Mezzagiorno (Portici). R. Lab. Pat. Veg. Portici **3** : 18–48, 1934.

Review of our knowledge so far on virus diseases of the potato. Discusses the different types of the diseases (mosaic, leafroll, witches' broom, pseudo-net necrosis and concentric necrosis). Quanjer's classification, Schander and Bielert necrotic tissue changes and Elze transmission by insects. Control measures are also considered.

Trümpener, Egon

Wie erkennt man den Abbau der Kartoffel? Der Kartoffelbau **17** : 61–62, 1933.

Die Blattrollkrankheit. (The leafroll disease.) Kartoffel **13** : 210–213, 1933.

Tu, C.

Notes on diseases of economic plants in South China. Lignan Sci. Journ. **11**(4) : 489–504. 1932.

This report contains several brief notes on virus diseases of economic plants.

Unite, Juan O., & Capinpin, J[osé] M[ananjaya]

Selection of mosaic free cuttings of sugar. Philippine Agric. **15**(2) : 67–73, 1926. (Planter & Sugar Manuf. **77**(8) : 147–148, 1926.)

A brief paper in which the authors report (1) that first generation plants from mosaic resistant plants in severely infected fields do not show symptoms of disease; (2) that mosaic-free points produce diseased plants; and (3) that buds from infected stalks invariably give diseased plants, although the leaves of these buds do not show symptoms.

Uppal, B[adri] N[ath]

The absorption and elution of cucumber mosaic virus. Indian Journ. Agric. Sci. **4**(4) : 656–662, 1934.

Detailed account, illustrated with tables, of an experiment on the filterability of the cucumber-mosaic virus.

The movement of tobacco mosaic virus in leaves of *Nicotiana sylvestris*. Indian Journ. Agric. Sci. **4**(5) : 865–873, 1934.

The author gives the results of his experimental observations.

The effect of dilution on the thermal death rate of tobacco-mosaic virus. Indian Journal Agric. Sci. **4**(5) : 874–879, 1934.

The effect of dilution on the thermal death rate of the ordinary tobacco-mosaic virus was determined by the local lesion method on *Nicotiana glutinosa*, and a hybrid between it and *N. tobaccum*. He gives the range of temperature and the correlation of dilution. The thermal death point was established close to 70° at a dilution of 1 in 10,000.

Valleau, W[illiam] D[ornay]

Ring spot (Delphinium sp.) Plant Dis. Rept. **65**: 419, 1927.

A record.

A virus disease of Delphinium reported from Kentucky, Plant Disease Reporter **14**: 118. 1930.

A record.

Do tobacco plants recover and develop an immunity from ring spot? Phytopathology (Abstract) **25**(1) : 37, 1935.

The resistance of Ambalema tobacco to different viruses. Phytopathology **25**(1) : 37, 1935.

Varadaraja Iyengar, A. V.

Contribution to the study of spike-disease of sandal (*Santalum album* Linn.) Part XVI—Distribution of arsenic in sandal-wood treated with sodium arsenite. Journ. Indian Inst. Science. **17A**(12) : p. 131–139, 1934.

Large quantities found in bark and sap wood; the movement was noticed in the roots; the greatest concentration in the girdled regions; root suckers often killed; the oil from sprayed trees did not contain the poison.

Venkata Rao, M. G.

A preliminary note on the leaf curl mosaic disease of Sandal. Mysore Sandal Spike Invest. Comm. Bull. **3**, 5 p., 1934.

Preliminary note appearing in Indian Forester 49(12) : 772–777, 1933.

---------- **& Gopalaiyengar, K.**

Studies in spike disease of Sandal. (1) Two types of spike disease. (2) The movement of the virus in Sandal plants. Indian For. **40**(7) : 481–491, 1934. (Mysore Sandal Spike Invest. Comm. Bull. **4**, 1934.)

The authors describe the two types of sandal spike observed in India. By means of grafting experiments the movement of the virus within the plants was observed.

---------- & ----------

Studies in spike disease of sandal. Methods of inoculation and variation of results under different methods. Mysore Sandal Spike Invest. Comm. Bull. **5**, 13 p., 1934.

Different methods of inoculation by which spike disease of sandal can be transmitted are described. The fundamental principle in all methods is the grafting of infected tissue on healthy stocks. Results are given.

----------, **& Iyengar, K. G.**

Studies in spike disease of Sandal. Methods of inoculation and variation of results under different methods. Indian Forester **40**(10) : 689–701, 1934.

Detailed account giving results obtained by the author with the methods successfully used in the transmission of sandal-spike disease.

The role of under-growth in the spread of the spike disease of sandal. Indian For. **41** : 169–188, 1935.

Verhoeven W[ilhelm] B[oudewijn] L[eenweburg]

Invloed van Zeenwsche Blauwen op Eerstelingen. (The influence of zeenwsche Blauwen on Eeerstelingen.) Tijdschr. over Plantenziekten **40**(8) : 173–174, 1934.

Report of the behavior and influence of potato varieties which harbours the streak virus in a marked form.

Verplancke, G[ermain]

La Culture de la Pomme de terre dans l'Etat de Maine (Etats-Unis d' Amérique.) (Potato culture in the State of Maine, United States of America.) 16 p., 1919.

On pages 9 to 14 the author discusses potato virus diseases.

De la valeur de la "Tuber testing method" dans la sélection sanitaire de la Pomme de terre. (The value of the tuber testing method in the sanitary selection of the potato.) Stat. Phytopath. de l'Etat. Gembloux (Belgium) 7 p., 1927.

Report of experiments on tuber testing for mosaic and leaf-roll disease.

Etude sur 1'organisation de 1'inspection sanitaire de la culture de la pomme de terre en Belgique.—Studie over de inrichting van den gezondheidskeuring van de aardappelkweek in België. (Study of the organization of the sanitary inspection of the potato culture in Belgium.) Belgium Min. Agric. 27 p, 1929.

Regulations for seed certification in which virus diseases receive considerable attention.

Experiences sur la transmission des maladies de dégénérescence de la Pomme de terre. II. Résultats des essais faits en 1931: (Experiments on the transmission of degeneration diseases of potato. II. Result of tests made in 1931.) Ann. Gembloux **37**: 345–349, 1931.

A continuation of studies in papers with the same title.

Étude comparative de Pomme de terre d' origines diverses. I. Resultats des expériences faites en 1931. (Comparative studies on potatoes from different origins. I. Results of experimnts made in 1931.) Bull. Inst. Agron. Sta. Res. Gembloux, **1**: 123–145, 1932.

Records and discussion of field results in tabular form.

Les viroses de la Betterave. (The viruses of beet root.) La Sucrerie Belge **53**(1) : 2–10, (2) : 22–32, 1933. (Publ. Inst. Belge Amélioration Betterave, Tirlemont, **6** : 232–248, 1933.)

The author states that mosaic and yellows are the two virus diseases affecting beet in Belgium. He is working on four types of mosaic; speckled, veined, marbled and pocked mosaics. He gives an account on the cytology, transmissibility, wild hosts, varietal resistance and insect vectors.

Étude comparative de pomme de terre d'origines diverses. III. Resultats des expériences faites in 1933. (A comparative study of potatoes of various origins. III. Results of experiments made in 1933.) Bull. Inst. Agron. Stat. Res. Gembloux, **3**(1) : 52–91, 1934.

Continuation of previous work.

Sur une forme nouvelle de la "bigarrure" maladie á virus filtrant de la pomme de terre. (On a new form of the "bigarrure" disease of the filterable virus of the potato.) Soc. Roy. Bot. Belg. **66**(2) : 107–121, 1934.

Description of type of acropetal necrosis of potato plant.

Contibution á l'étude des maladies á virus filtrants de la Betterave. (Contribution to the study of filterable virus diseases of beet root.) Mem. Acad. Roy. Belgique, Cl. Sci. Ser. II, **1451** (III, 1), 104 p., 1934.

This article is an expansion of the author's paper entitled "La virose de la Betterave" cited above.

Etude des propriétés des virus causant les maladies de dégénérescence de la betterave. (Study of the properties of viruses causing degeneration diseases of the beet.) Sucrerie Belge **54** : 142–251, 1934.

General discussion of the properties of the viruses causing degeneration diseases of the beet, specially yellowing and mosaic.

---, **& Asaert, L.**

Immunologische reakties met plantenvirus (Immunological reactions with plant viruses.) Natuur, Tijdschr. **16**(2–5) : 69–74, 1934.

The authors give brief notes of their results obtained with different viruses and different hosts.

Etat actual de la production de la pomme de terre in Belgique. (Actual state of potato production in Belgium.) Reprint from Ann. de Gembloux p. 185–196, May, 1934.

In this paper is given the effect of virus diseases on the production of potatoes in Belgium.

Verwoerd, Len

Storage pit (bitterpit) of apples. Farming So. Africa **8**: 418, 430, 1933.

Vielwerth, V.

Mosaika Americké Revy vinné. (Mosaic of the American vine.) Ochrana Rostlin **13**(3–4): 83–90, 1933.

Account of a disorder on American varieties of vines occurring in Czechoslovakia which is transmitted by grafting and with characteristics of a virus disease. Studies of the disease are in progress.

Vizioli, J[osé]

O mosaic e outras molestias da canna de assucar. (Mosaic and other diseases of sugar cane). Sec. Agric. S. Paulo (Brasil) 1926.

Vos, H[uguette] C[onstance] C[ornelia] A[driana] A[bdia]

De invloed van *Pseudococcus citri* (Risso). Fern. op de plant. Baarn—N. V. Hollandia-Drukkerij pp. 1–81. 1930.

This paper is not a discussion of a virus disease but is of interest to those who are studying the mechanics of plants with viruses spread by means of insect vectors.

Wallace, H. F., & Anders, C. B.

Report Raymond Branch Experiment Station. Mississippi Agric. Expt. Sta. Bull. **231**, 1925.

On pages 14–17 data are given as to the results of seed certification in relation to potato-mosaic disease.

Report of the Mycologist, 1933. Tanganyika Territory Dept. Agric. Ann. Rpt. **1933**: 76–78, 1934.

Notes on virus diseases of cowpeas, other legumes, tobacco and sesame are included in this report.

Weger, F. W.

Die Blattrollkrankheit der Kartoffel. (The leaf roll disease of potato.) Ztschr. f. Pflanzenkrank. 29, 1919.

Bekämpfung der Blattrollkrankheit der Kartoffel. (The fight of leaf roll diseases of potato.) Landw. Brennerei Ztg. Prag. 1920.

Went, F. A. F. C.

Does the Sereh disease exist in the West Indies, more especially in Trinidad? West Indian Bulletin **12**(4) : 554–560, 1912.

He found suspicious cases but no positive evidence of the disease.

Weimer, J[ames] L[e Roy]

Effect of environmental and cultural factors on the dwarf disease of alfalfa. Journ. Agric. Res. **47**(6) : 351–368, 1933.

Continuation of previous work. This is the result of experiments started in 1929. Dwarf disease of alfalfa is not regarded by the author as a virus disease.

Wellman, F. L.

Identification of celery virus 1, the cause of southern celery mosaic. Phytopathology **24**(7) : 695–725, 1934.

Description of a new virus which causes a true mosaic on celery. It is transmitted by *Aphis gossypii* and artificial juice inoculation.

A disease of banana markedly similar to bunchy top, produced by celery virus 1 in U. S. A. Phytopathology **24**(9) : 1032–1034, 1934.

Report of the susceptibility of *Musa sapientum* variety Lady Finger to celery virus I. Gives behavior of the disease in inoculated plants, aphis transmission and description of the disease.

Infection of *Zea Mays* and various other Gramineae by the celery virus in Florida. Phytopathology **24**(9) : 1035–1037, 1934.

The author found that celery virus I, the cause of southern celery-mosaic, has a very wide host range. He gives the host plants affected by this virus. He made inoculations by means of *Aphis gossypii*.

Dissemination of southern celery-mosaic virus on vegetable crops in Florida. Phytopathology **25**(3) : 289–308, 1935.

This virus is carried by *Aphis gossypii* and attacks celery, corn, squash, pepper and several weeds. The author traces its dissemination by *A. gossypii* from weeds throughout the celery fields and illustrates his studies with maps.

The host range of the southern celery-mosaic virus. Phytopathology **25**(4) : 377–404. 1935.

The paper is the result of experimental studies with celery virus I. The author was able to produce the disease by inoculation in 91 species of plants including 29 vegetable crops.

Werner, H[arvey] O[scar]

Spindle tuber, the cause of "run-out" potatoes. Amer. Potato Journ. **3**(1) : 16–18, 1926.

Popular—Some of the work in this paper will be found in Nebraska Sta. Bull. 207.

The tuber-index method in relation to seed potato certification. Proc. 15th Ann. Meeting Potato Assoc. Amer. p. 140–150, 1928.

Brief review of other workers, tuber-index system of eliminating virus diseases. He reports in this paper the results he obtained.

Effect of cultural methods and maturity upon the seed value of Eastern Nebraska potatoes. Nebraska Agric. Expt. Sta. Res. Bull. **45**, 45 p., 1929.

This paper contains valuable information in regard to spindle tuber of potatoes.

Some facts concerning the productivity of irrigated seed potatoes. Proc. 16th Ann. Meeting Potato Assoc. Amer. p. 120–132, 1929.

The author gives the results of his observations in regard to the relation of irrigated and not-irrigated potatoes to spindle-tubers.

Werwoerd, L.

Storage pit (bitter pit) of apples. Farming South Africa **8**: 418–430, 1933.

Whetzel, H[erbert] H[ince]

Diseases of the peony. Amer. Florist **44**: 609–612, 1915.

White, Philip, R.

Multiplication of the viruses of tobacco and aucuba mosaics in growing excised tomato root tips. Phytopathology **24**(9): 1003–1011, 1934.

Description of a method for the cultivation of the viruses of tobacco-mosaic and aucuba mosaic in isolated root tips grown *in vitro*.

White, Richard P[eregrine]

Diseases of ornamental plants. New Jersey Agri. Exp. Station, Circ. **226,** 98 p., 1931.

This is a popular bulletin containing brief descriptions of yellows on *Callistophus chinensis,* stunt on *Dahlia* spp., mosaic on *Gladiolus* spp. mosaic on *Iris* spp., mosaic on *Paconia officinalis,* mosaic on *Rosa* spp. mosaic on *Lathyrus odoratus,* breaking in *Tulipa* spp., and mention of mosaics on several other plants. Parts of this bulletin were published separately. See our original bibliography.

The effect of mosaic on bloom production of the Talisman rose. Phytopathology **24**(10) : 1124–1125, 1934.

Brief notes reported the effects of mosaic on the Talisman rose variety. According to the author it is of relatively slight influence from the practical viewpoint.

Whitehead, T[atham]

Immature *versus* mature seed potatoes. Journ. Mins. Agric. p. 452–455, 1930.

Contains brief notes on potato leaf roll.

Wilcox, R[aymond] B[oorman]

Identification of false blossom in the fall. Proc. Amer. Cranberry Grow. Assoc. **61** : 15–16, 1930.

Popular.

--- **& Beckwith, C[harles] S[teward]**

A factor in the varietal resistance of cranberries to the false-blossom disease. Journ. Agric. Res. **47**(8) : 583–590, 1933.

A study of relative varietal susceptibility based on the poison formula of cranberry to the false-blossom disease.

---, **&** ---

The false-blossom disease of cranberries. New Jersey Agric. Expt. Sta. Circ. **348,** 1935.

Wiles, D. R. D.

Report of Plant Diseases Inspector. Barbados Agric. Journ. **3**(2) : 39–44, 1934.

This report gives data on the present position of sugar-cane mosaic in Barbados.

Wilson, G. L.

Yellow stripe. Daffodil Year Book Roy. Hort. Soc. **1933** : 74, 75, 1934.

Wingard, S[amuel] A[ndrew] & Godkin, J.

Tobacco diseases in Virginia and their control. Virginia Agric. Expt. Sta. Bull. **90,** 31 p., 1924.

Bitter pit of apples. Virginia Fruit **20**: 8–10. 1932.

Woods, Albert F[red]

"Bacteriosis" of carnation. Bot. Gaz. Sept. 1897 p. 200–205, 1897.

It is admitted that the disease does not show the presence of bacteria but the causal agent is not known. The effects of aphid and thrip injuries are considered.

Stigmonose: A disease of carnation and other pinks. U.S.D.A. Div. Veg. Phy. & Path. Bull. **19,** 30 p., 1900.

The author admits that the disease is not caused by fungi nor bacteria, although no definite cause is recognized. The influence of aphis, thrips and red spiders as carriers is recognized.

Mosaic-disease study points to control by resistant varieties. U.S.D.A. Yearbook, **1927**: 464–465, 1928.

Brief popular notes.

Woods, Mark W.

Cellular changes in ring-spot. Contr. Boyce Thomp. Inst. **6** (1): 51–67, 1934.

The author describes the symptoms and gives the results of studies on the histology and cytology in the lesions.

Yenemaru, C[hutaro] & Asa, K[eijiro]

(Report of the investigations of dwarf disease of rice plant.) Kyoto Agric. Expt. Sta. Spe. Bull. 1, 178 p., 1905.

Youden, W. J., & Beale, Helen Purdy

A statistical study of the local lesion method for estimating tobacco mosaic virus. Contr. Boyce Thomp. Inst. **6**(3): 437–454, 1934.

Discussion of the subject and description of the method designed.

---, ---, **& Guttine, John D.**

Relation of virus concentration to the number of lesions produced. Cont. Boyce Thompson Inst. **7**(1): 37–53, 1935.

"Data from six different sources showing lesion counts obtained by applying virus to extracts to leaves in a series of dilutions have

been assembled from the literature. The authors include work with tobacco mosaic, ring spot, cucumber mosaic, and aucuba mosaic viruses. The plants used as hosts have been *Nicotiana glutinosa*, *Phaseolus vulgaris* var. Early Golden Cluster, and *Vigna sinensis* var. Black Eye. The method of evaluating the constants is given and their possible significances discussed. The utility of the function as a guide to the design and interpretation of experiments is shown by examples.''

Zeijistra, Fzn, H. H.

Versuch einer Erklärung der ''Sereh'' Erscheinungen des Zuckerrohrs. Ber. Deutsch. Bot. Ges. **20**(6): 330–333, 1911. (Bot. Centralb. **119**(6): 151, 1912.)

Review of the work so far on this obscure disease.

Zimmermann, A[brecht Wilhelm Philipp] von

Die Kräuselkrankheit des Manioks (Mhogo). 1 und 2 Mitt. (The leaf curl of cassava.) Pflanzer, Tanga **2**: 145–153, 182–183, 1906.

Popular.

Einge Bemerkungen über, Kassave (Mhogo). (Some notes on cassava.) Pflanzer, Tanga, **2**: 257–271, 1906.

Account on botany, varieties and diseases of cassava including leaf curl.

Ueber eine Krankheit der Erdnusse. (*Arachis hypogea.*) (A disease of peanuts.) Pflanzer, Tanga, **3**: 129–133, 1907.

Notes on peanut rosette disease.

Die Deutsch—Ostafrikanischen Maniok—Varietaten (1). (Varieties of cassava in German East Africa.) Pflanzer, Tanga **3**: 258–269, 1907.

Contains reference to susceptible and resistant varieties to curl disease.

Die Kräuselkrankheit des Maniok (Mhogo) und die Abgabe gesunder Stecklinge. (The leaf curl of cassava and the delivery of healthy cuttings.) Pflanzer, Tango **5**: 184–185, 1909.

Popular.

Die Kräuselkrankheit der Erdnusse. 2 Mitt. (Rosette disease of peanuts.) Pflanzer, Daressalam **9**: 59–63, 1913.

Brief account of peanut rosette with illustrations.

Zundel, G[eorge] L[orenzo Ingram]

Rasberry diseases. Pennsylvania Agric. Expt. Sta. Circ. **133**, 20 p., 1934.

This paper includes brief practical notes on several virus diseases of the raspberry.

INDEX

OF FIRST SUPPLEMENTS TO PARTIAL BIBLIOGRAPHY OF VIRUS DISEASES OF PLANTS

	Page
Abacá (See also *Musa*)	
bunchy top	186
mosaic	145
Abutilon	
mosaic	216
virus	139
Agallia sanguinolenta	140
Agallia stricticollis	156
Ageratum conyzoides	215
Alfalfa (See *Medicago*)	
Almond (See *Prunus*)	147
Alopecurus fulvus	158
Altitude	141, 148, 153
Ambalema tobacco	217
Anuraphis padi	136
Aphid	136
Aphis gossypii	188, 222
Aphis maidis	212
Aphis rumicis	165
Apium graveolens	
mosaic	141
mosaic hosts	223
virus 1	222
yellows	204
Apple (See *Malus*)	
Arachis	142
curl	135
rosette	135, 226, 227
Argyscia cuneata	
stripe	175
Aster yellows (See also *Callistephus*)	188, 204
hosts of	203
Atriplex fructiculosa	
curly top	204
Attenuation	172
Aucuba mosaic	172, 205, 223, 226
of tomato	144, 146
Avocado (See *Persea*)	

Page

Bacteriophage ---- 160
Bacteriosis
carnation (See also *Dianthus*) ---- 225
Balclutha mbila ---- 212
Banana (See *Musa*)
Barley (See *Hordeum*)
Bean (See also *Phaseolus*)
mosaic ---- 135, 162, 164, 190, 194, 208
Beets (See *Beta*)
Bemisia ---- 215
Bemisia gossypiperda ---- 149, 179
Berberis vulgaris ---- 141
Beta vulgaris ---- 166, 169, 177, 194
curly top ---- 137, 139, 142, 150, 155, 156, 158, 190, 194, 201, 202, 203, 204
degeneration ---- 220
leaf hopper ---- 137
mosaic ---- 207, 220
"Bigarrure" of potato ---- 220
Big bud of tomato ---- 196
Big vein of lettuce ---- 165
Bitter pit (See *Malus*)
Bodies (intra-cellular) ---- 156, 158, 168, 171, 173, 174, 205, 213
Brassica
mosaic ---- 140
virus ---- 215
Break down (See also *Malus*)
apple ---- 145
Breaking (See Tulips)
Buckskin
cherry (See also *Prunus*) ---- 194
Bunchy top (See *Musa*)

Cabbage (See *Brassica*)
Cactus ---- 208
Calico
petunia ---- 206
potato ---- 164, 191
tobacco ---- 206
tomato ---- 206
Cane (sugar) See *Saccharum*
Callistephus chinensis
aster yellows ---- 224
Capitophonis fragariae ---- 146, 178
Capsicum frutescens ---- 164
Carnation (See *Dianthus*)
Cassia auriculata ---- 175
Cassava (See *Manihot*)

Page
Cauliflower (See *Brassica*)
Cell ---- 138
Celery (See *Apium*)
Celosia argentea ---- 203
Certification of seeds ---- 156, 215, 219, 222, 223
Chemistry ---- 137, 144, 153, 165, 168, 179, 191, 205, 209, 210, 213, 216
Cherry (See also *Prunus*) ---- 147
  buckskin ---- 194
Chondriome-plastidome ---- 138
Chlorita flavescens ---- 141
Chlorophyll ---- 164
Chlorosis ---- 181, 196
  citrus ---- 136, 140
  peach ---- 161
Cicadula divisa ---- 204
Cicadula sexnotata ---- 204
Citrus ---- 136
  chlorosis ---- 136, 140
  mottle leaf ---- 195
  psorosis ---- 156
Classification ---- 157, 166, 171, 182
Climate ---- 143
Clover
  mosaic ---- 138
Coffee (See *Coffea*)
Coffea phloem necrosis ---- 178, 191, 210
Convolvulus arvensis
  spotted wilt ---- 161
Coreopsis tinctorea
  aster yellows ---- 203
Corn (See *Zea mays*)
Cosmos pipinnatus
  aster yellows ---- 203
Cotton (See *Gossypium*)
"Court-noue" (see also vine) ---- 146, 154, 189, 190
Cowpea ---- 221
Cranberry false blossom (See also *Vaccinum*) ---- 139, 148, 211, 224
Crinkle
  apple ---- 145
  potato ---- 147, 192
  strawberry ---- 186
  tobacco ---- 182, 197, 215
Crucifers ---- 207, 215
Cucumber (See *Cucumis*)
Cucumis sativus ---- 167
  mosaic ---- 130, 142, 147, 163, 168, 191, 217, 226
Cultivation of virus ---- 174

Page

Curl leaf ---- 141, 174, 226
Curly dwarf of potato ---- 211
Curly top
beet ---- 139, 150, 156, 158, 190, 194, 201, 202, 203, 204
Erodium cicutarum ---- 172
tomato ---- 154
Currant (See *Ribes*)
Cytology ---- 153, 154, 220, 225
Cytoplasm ---- 155

Dahlia
mosaic ---- 131, 138, 143, 177, 224
stunt ---- 143
virus ---- 145
Datura stramonium ---- 138, 192
Deamination ---- 165
Degeneration ---- 170, 183
apple ---- 190
beets ---- 155, 220
pear ---- 190
potato ---- 133, 143, 148, 153, 156, 161, 162, 175, 177, 181, 184, 187, 189, 211, 219
sugar cane ---- 148
Delphinium ---- 163, 173, 217
Dianthus
bacteriosis ---- 225
stigmonose ---- 225
Dianthus plumarius ---- 203
Dodonaea viscosa ---- 175
Dwarf
alfalfa ---- 222
curly ---- 182
onion ---- 152
potato ---- 140, 164, 170, 211
rice ---- 131, 132, 141, 150, 157, 165, 184, 214, 225
sugar cane ---- 139
Drupaceous fruit ---- 136

Echinochloa crus-galli ---- 158
Enzymes ---- 138, 198
Erodium cicutarum ---- 172
Eutettex tenellus ---- 146, 190, 201, 202, 203

False blossom (See also *Vaccinum*) ---- 139, 148, 211, 224
Ferns ---- 163
Fern leaf ---- 196, 200, 208
Ficus (fig) ---- 175, 185
Fiji disease ---- 139, 155, 185, 186, 200
Filters, filtration, etc. ---- 171, 191, 210, 220

Page

Fragaria 146
crinkle 186
degeneration 162
yellow edge 178
Frankiella insularis 150

"Gelestrepenziek" (See mosaic)
Giant hill 163
Gladiolus 224
Gloxinia speciosa 161
Gossypium
abnormality 130
leaf curl 137, 149, 171, 179, 214
stenosis 130

Heat treatment 171
Helichrysum bracteatum 203
Hicoria (Pecan rosette) 151
Histology 149, 175, 195, 225
Hippeastrum 171
Hordeum vulgare 184
Hyperplasia 155

Immunity 144, 154, 197, 220
Intracellular inclusion 205
Inactivation 154
Insects 138, 140, 141, 146, 147, 149, 150, 152, 156, 157, 158, 159, 162, 165, 166, 170, 176, 177, 178, 179, 180, 186, 187, 188, 190, 192, 196, 201, 202, 203, 204, 206, 208, 209, 210, 211, 220, 221
Iris 224
Irradiation 154

Jute (See *Corchorus*)

Kassave (Cassava) (See also *Manihot*) 226
Kochia scoparia 203
"Kroepoek", "Kroeppoek", "Krepoli", Kromnek" etc. of tobacco, tomato, etc. 158, 169, 182, 197, 201, 215

Lactuca 165, 215
Lappa 200, 204
Lathyrus odaratus 224
Leaf curl 137, 149, 171, 179, 182
Leaf hopper 137, 140, 145, 166, 169
Leaf roll 166
cassava 226
potato 142, 143, 144, 151, 162, 166, 167, 171, 172, 182, 184, 192, 211, 216, 221, 224
tomato 156
Leaf stripe of corn 172

Page
Legumes ---- 166, 221
Lettuce (See *Lactuca*)
Light ---- 154, 164
Lily mosaic ---- 149
Lincus bug ---- 210
Little leaf ---- 147, 175, 176
Lupins ---- 184
Lycoperscicon esculentus
aucuba mosaic ---- 144
big bud ---- 196
calico ---- 206
chemical splitting of virus ---- 205
crinkle ---- 197
curly top ---- 154
die back ---- 205
fern leaf ---- 196, 208
"kropoek" ---- 197
"kromnek" ---- 186
leaf roll ---- 156
mosaic ---- 138, 139, 144, 146, 167, 168, 180, 195
necrosis ---- 138
spotted wilt ---- 134, 139, 152, 158, 159, 161, 181, 190, 205, 206, 215
streak ---- 130, 139, 204, 211
virus ---- 130, 147, 167, 168, 173
winter blight ---- 211
woodiness ---- 196

Macropis trimaculata ---- 176, 177
Macrosiphum gei ---- 187, 188
Macrosiphum pisi ---- 165
Maize (See *Zea mays*)
Malus
breakdown ---- 145
bitterpit ---- 135, 136, 145, 223, 225
crinkle ---- 145
degeneration ---- 190
necrosis ---- 190
rosette ---- 182
storage pit ---- 223
virus ---- 147
Manihot ---- 226
Maniock (See *Manihot*)
Manthola incana ---- 216
Masking ---- 163
"Matizado" (See sugar cane mosaic)
Mechanics of insect transmission ---- 207, 222
Medicago
dwarf ---- 122
yellows ---- 160
Methylene blue ---- 140

Mhogo (See *Manihot*)
Page
Mild mosaic of potato ---- 211
Mirambilis jalapa ---- 203
Moonia albimaculata ---- 152, 209
Mosaic ---- 140, 142, 159, 163, 185, 225
abacá ---- 145
abutilon ---- 216, 223, 226
Apium gravelens ---- 141
aucuba ---- 144
beans ---- 162, 164, 190, 194
beets ---- 202, 207
cabbage ---- 140
cane (sugar) ---- 131, 133, 134, 135, 142, 145, 148, 150, 151, 155, 178, 180, 181, 186, 197, 200, 201, 213, 216, 221, 224
celery ---- 141, 223
chemistry ---- 144
clover ---- 138
color ---- 149
cucumber, cucurbits ---- 130, 142, 145, 163, 168, 169, 191, 217, 226
dahlia ---- 131, 138, 143, 177
fig ---- 185
Hippeastrum ---- 171
Iris ---- 224
Lappa ---- 204
Lathyrus odoratus ---- 224
lilies ---- 188
movement of virus ---- 217
Narcissus ---- 147
Nature of ---- 151
Nicotiana glutinosa ---- 208, 211
Papaver nudicaule ---- 185
pea ---- 187, 188
peach ---- 141, 146
Phaseolus ---- 162, 211
poppy ---- 185
potato ---- 138, 144, 157, 170, 171, 175, 182, 184, 192, 211, 216, 221
raspberry ---- 162, 174
rose ---- 143, 224
Santalum ---- 218
seed production ---- 164
soy bean ---- 162
spinach ---- 167, 208
sugar cane (See *Saccharum*)
tobacco ---- 134, 137, 138, 148, 154, 155, 163, 164, 167, 168, 173, 174, 175, 177, 179, 189, 191, 206, 208, 210, 211, 213, 214, 215, 217, 223
tomato ---- 138, 139, 144, 146, 168, 180, 195
vines ---- 140, 177, 221
wheat ---- 189
Zinnia elegans ---- 185

Page
Mottle leaf of citrus ---- 195
Movement of virus ---- 144
Musa
bunchy top ---- 135, 185, 186, 188, 205, 222
celery virus I ---- 222
mosaic ---- 145
Mycoplasm ---- 155
Mythelene blue ---- 212
Myzus persicae ---- 147, 159, 162, 165, 170, 192

Narcissues mosaic ---- 147
Necrosis ---- 160, 190
apple ---- 190
beet ---- 155
coffee ---- 178, 190, 191, 210
pear ---- 190
potato ---- 138, 149, 159, 198, 200, 216, 220
tobacco ---- 176
Nephotettix apicalis ---- 157, 158
Nicotiona glutinosa ---- 138, 144, 208, 211, 217, 226
sylvestri ---- 217
tobacum ---- 158, 176, 211, 217, 225
big bud ---- 196
calico ---- 206
crinkle ---- 187, 215
dwarf ---- 182
fern leaf ---- 168
"kromnek" ---- 135, 182
mosaic ---- 134, 137, 138, 148, 154, 155, 163, 164, 167, 168, 173, 174, 175, 177, 179, 189, 191, 198, 210, 213, 214, 221, 223, 226
necrosis ---- 138
ring spot ---- 147, 155, 226
virus ---- 142, 154
Nochmals ---- 208
Nuclei ---- 156

Organisms ---- 154
Ornamentals ---- 203, 224
Oryza
dwarf ---- 131, 132, 141, 150, 157, 165, 172, 184, 214, 225
Overwintering ---- 140, 203

Panicum miliaceum ---- 158
Papaver mosaic ---- 185
Paratrioza cockerelli ---- 174
Particle size ---- 214

Pea mosaic (See *Pisum sativum*)
Peach (See *Prunus*)
Peanut (See *Arachis*)
Pear (See *Pyrus*)
Pecan rosette (See *Hicoria*)
Page
Pelargonium ---- 141, 203
Pentalonia nigronervosa ---- 186
Peony ---- 223
Peregrinus maydis ---- 137
Perkinsiella vastatrix ---- 186
Persea Persea sun blotch ---- 164
Petunia ---- 203, 206
Phaseolus ---- 162, 211, 226
mosaic ---- 135, 162, 164, 190, 194, 208
Phloem necrosis ---- 160, 190
coffee ---- 178, 191, 210
Phony peach (See also *Prunus*) ---- 164, 173
Photodynamic ---- 140, 212
Physiology ---- 163, 195, 198
Pisum sativus
mosaic ---- 187, 188
Plantain mosaic (See also *Musa*) ---- 188
Plastids ---- 156
Pome fruits bitter pit ---- 135
Poppy mosaic ---- 185
Potato (See *Solanum tuberosum*)
Primula virus ---- 207
Properties of curly top virus ---- 139
Properties of virus ---- 220
Protozoa ---- 140
Prunus Americana ---- 176
Amygdalus ---- 147
Avium ---- 147
cerasus ---- 147
domestica ---- 176
musnoniana ---- 176
myrobalon ---- 176
salicina ---- 176
simoni ---- 171
Prunus
chlorosis ---- 161
phony ---- 164, 173
rosette ---- 171
Pseudococcus citri ---- 221
Pseudocommunis ---- 197
Pseudo-net-necrosis (See necrosis)
Psorosis of citrus ---- 156
Psyllid yellows ---- 157
Pterolobium indicum ---- 175

	Page
Pyrus	
bitter pit	136
degeneration	190
necrosis	190
rosette	182
Quince bitter pit (See also *Malus*)	136
Raspberry (See *Rubus*)	
Refraction	213, 214
Resistance	208, 211, 216
Reversion in currants	148
Ribes reversion	148
Rice (See *Oryza*)	
Ring spot	
potato	170, 171
tobacco	155, 228
Delphinium	217
Roguing	189
"Roncet" of vine	141, 177
Rose	143, 147, 224
Rosette	175
apple	182
fruit trees	147
pecan	151, 185
peach	171
peanut	135, 226, 227
pear	182
Rotterdam D disease	139
Rubus	162, 167, 174, 227
virus	148
Running out of potatoes	184, 223
Saccharum officinarum	
Fiji	135, 139, 155, 185, 186, 200
mosaic	**131, 133, 134, 139, 142, 145, 148, 150, 151, 155, 178, 180, 181, 186, 197, 200, 201, 213, 215, 216, 221, 224**
Sereh	162, 222, 226
streak	**139, 152, 155, 179**
Santalum album	
mosaic	218
spike	**136, 143, 152, 165, 166, 175, 209, 218**
transmission of spike	134
Scabiosa atropurpurea	203
Seed production	164
Seed transmission	168
Sereh (see also *Saccharum*)	162, 222, 226
Serology	147, 160, 161, 191
Serratia marcescens	154
Sesame	221
Sisal	183

Page
Solanaceae ---- 206
Solanum indicum spike ---- 175
Solanum tuberosum ---- 138, 141, 152, 187, 188, 189, 201, 219
anatomy ---- 135
"bigarrure" ---- 220
calico ---- 164, 191
certification ---- 156, 215, 223
concentric necrosis ---- 216
crinkle ---- 147, 192
curly dwarf ---- 211
degeneration ---- 133, 143, 148, 153, 156, 161, 162, 169, 175, 181, 184, 187, 189, 211, 219
dwarf ---- 140, 164, 170, 211
giant hill ---- 163
latent virus ---- 167, 213
leaf roll ---- 142, 143, 144, 151, 152, 162, 166, 167, 171, 172, 182, 192, 195, 198, 211, 216, 221, 222, 224
mosaic ---- 138, 114, 157, 170, 171, 175, 192, 206, 211, 216, 221
mottle ---- 170
necrosis ---- 138, 149, 159, 198, 200, 116, 220
pseudo-net necrosis ---- 216
psyllid yellows ---- 157, 174
resistance ---- 211
ring mosaic ---- 171
ring spot ---- 170
running out ---- 223
spindle-sprout ---- 211
spindle tuber ---- 137, 159, 161, 223
spotting ---- 158
stipple streak ---- 192
streak ---- 218
tuber unit ---- 156
veinbanding ---- 167
virus ---- 130, 157, 158, 159, 160, 169, 171, 181, 182, 185, 197, 206, 207, 221
witches' broom ---- 216
Sore shin of *Lupins* ---- 184
Sorghum ---- 137
Sound ---- 210
Soy bean mosaic ---- 162
Spike disease—
Argyreia cuneata ---- 175
Cassia auriculata ---- 175
Dodonaea viscosa ---- 175
Ficus tsiela ---- 175
Pterolobium indicum ---- 175
Sandal (see *Santalum album*)
Solanum indicum ---- 175
Zizyphus aenoplia ---- 175

Page

Spinach blight 167, 208
Spindling sprout of potato 212
Spindling tuber of potato 159, 161, 223
Spotted wilt 134, 139. 152, 158, 159, 161, 181, 190, 205, 206, 215
Spread of diseases 156
"Stolbur" 196
Starch 173
Stenosised 173
Storage pit 223
Stenosis of cotton 130
Stigmonose of carnation 225
Stipple streak of potato 191
Strawberry (see *Fragaria*)
Streak 140
  cane 139, 152, 155, 179
  corn 164, 212
  die back form 214
  potato 157, 218
  raspberry 174
  roses 143
  tomato 139, 144, 205, 212, 213
Stripe of corn 172
Stripe of *Daffodil* 224
Stunt
  Dahlia 143, 224
Sugar beet (See *Beta vulgaris*)
Sugar cane (See *Saccharum*)
Sunblotch of avocado 164
Synedrella nodiflora 215

Temperature 145, 146, 154, 217
Teratology of ferns 163
Thammotettix 204
Thysanoptera 150
Tobacco (see *Nicotiana*)
Tomato (see *Lycopersicon*)
Transmission 135, 138
  by insects 138, 140, 141, 144, 152, 157, 162, 164, 165, 166, 170, 186, 187, 188, 190, 192, 196, 201, 202, 203, 204, 206, 207, 208, 209, 212, 220
  by seeds 168, 169
  of spike disease 134
  of strawberry virus 146
Treatment by heat 171
Tropaelum majus 203
Troximon glaucum 188
Tuber units 156, 157
Tulip breaking 146, 180, 205, 216, 224
Tumors 160

Page
Ultra-microscopic ---- 143, 184, 198, 206

Vaccinium ---- 139, 184, 211, 198, 224
Variegations ---- 172, 196
Veinbanding of potato ---- 167, 170
Vernonia cenesca ---- 215
Vicia faba ---- 161, 165, 194
Vigna sinensis ---- 226
Vine ---- 146, 154, 159, 176, 177, 189, 221
Viola ---- 203
Virescens ---- 196
Viruses ---- 130, 135, 136, 137, 141, 142, 143, 144, 150, 153, 154, 160, 161, 162, 165, 166, 170, 172, 174, 175, 180, 181, 182, 183, 185, 187, 190, 196, 207, 210, 216
- Abutilon ---- 169
- apple ---- 147
- beet ---- 139
- cauliflower ---- 215
- celery ---- 222
- crucifers ---- 215
- citrus ---- 156
- classification ---- 166
- complex ---- 205
- dahlia ---- 145
- *Delphinium* ---- 163
- *Hibiscus cannabinus* ---- 173
- intracellular inclusions ---- 205
- latent potato ---- 167
- *Nicotiana* ---- 158
- mosaic ---- 140
- movement ---- 144
- potato ---- 156, 157, 159, 160, 169, 171, 197, 207, 221
- *Primula obconica* ---- 207
- property ---- 150, 220
- rice ---- 172
- strawberry ---- 146
- tobacco ---- 208
- raspberry ---- 148, 167, 174, 227
- tomato ---- 130, 147, 167, 168, 173, 205, 208

Virus X ---- 160, 175
Virus Y ---- 160, 207

Water core of apples ---- 145
Weeds ---- 190, 200, 222
Wheat mosaic ---- 189
Wilt plum ---- 159
Winter blight of tomato ---- 212
Witches' broom of potato ---- 216
Woodiness of tomato ---- 168, 167, 171

Page
X-virus 160, 167, 171

Y-virus 160, 167
Yellow—
disease of beets 194
edge of strawberry 178
mosaic 144, 190, 191, 208
Yellows
alfalfa 160
aster 188
Delphinium 173
peach 161, 171, 176, 180, 213
psyllid of potatoes 157
Z-virus 167

Zea Mays 137, 164, 172, 212, 222
streak 164, 212
stripe 137, 172
Zinnia elegans 185, 203
Zizyphus aerophyla spike 175

AUTHORS

	Pages
Abzal, H. M.	130
Ainsworth, G. C.	130
Allen, H. F.	131
Altson, R. A.	131
Anders, C. B.	221
Anderson, E. J.	167
Ando, H.	131
Anonymous	131
Apanna, M.	152
Archibald, E. S.	135
Artsschwager, E. F.	135
Asa, K.	225
Asaert, L.	220
Ashby, S. F.	135
Atanasoff, D.	135
Austin, M. D.	136
Badami, V. K.	136
Bailey, M. A.	137
Baker, R. E. D.	137
Bald. J. G.	150
Ball, E. D.	137
Baribeau, B.	137
Barton-Wright, E.	137
Baudys, E.	137
Bawden, F. C.	138, 198
Beauverie, J.	138
Beale, H. P.	138, 225
Bechhold, H.	138
Beckwith, C. S.	139, 224
Bell, A. F.	139
Bennet, C. W.	139
Berkeley, G. H.	130, 139
Bewley, W. F.	139
Birkeland, J. M.	140
Bitancourt, A.	140
Black, L. M.	140
Blattny, C.	140, 195
Bodine, E. W.	141
Bohme, R. W.	141
Bokura, U.	141
Bolley, H. L.	142
Bonde, R.	201
Boning-Seubert, E.	142
Booberg, K. G.	142
Bouffil, F.	142
Bouquet, A. G. S.	180
Bouriquet, G.	142
Boysen-Jensen, P.	142
Brandes, E. W.	142
Brehmer, G. von	142
Brierly, P.	143
Brierly, W. B.	143
Brion, R. M.	184
Brown. B. A.	143
Buhay, G. G.	186
Burnett, G.	163, 168
Bushnell, J.	143
Butler, E. J.	143
Butler, O.	144
Cladwell, J.	130, 144
Calinisan, M. R.	145
Campbell, A. H.	145
Capinpin, J. M.	216
Carne, H. L.	151
Carne, W. M.	145
Carpenter, C. W.	145
Carter, W.	146
Castolla, F. de	146
Cation, D.	146
Cayley, D. M.	146
Chamberlain, E. E.	146, 184
Chandler, N.	165
Chandler, W. H.	147
Chester, K. S.	147
Chittenden, F. J.	147
Christoff, A.	147
Clark, C. F.	211
Clinch, P.	147
Clinton, G. P.	148
Cockerham, G.	137
Conceicao, C.	148
Conners, I. L.	148
Cooley, L. M.	148
Coombs, J.	148
Coons, G. H.	142
Corbett, W.	138

	Pages
Costa Lima, A.	148
Costantin, J.	148
Cowland, J. W.	149
Crane, H. L.	151
Crawford, R. F.	146
Crew, F. A. E.	149
Cristinzio, M.	149
Cunningham, H. S.	149
Currie, J. F.	149
Curzi, M.	150
Daikuhara, G.	150
Dana, B. F.	150
Davidson, J.	150
Davis, R. L.	150
Demandt, E.	151
Demaree, J. B.	151
Daniels, L. B.	174
Desai, S. V.	151
Dessi, G. H.	173
Diehl, R.	153
Dieselben	151
Dix, W.	151
Dobrozrakova, T. L.	152
Dodds, H. H.	152
Doolittle, S. P.	152
Dover, C.	152
Drake, C. J.	152
Dupbois, P.	153
Ducomet, V.	153
Dufrénoy, J.	153, 195, 206
Dufrénoy, M. L.	153
Duggar, B. M.	154, 174
Dutt, K. M.	200
Earle, F. S.	155
Eckart, C. F.	155
Elze, D. L.	192
Emmerez de Charmoy, D. d'	155
Erbe, F.	138
Erikkson, J.	155
Esau, K.	155
Fawcett, G. L.	156
Fawcett, H. S.	156
Finkembrink, W.	195
Folsom, D.	156, 200
Fowler, E. D.	151
Fowlie, P.	152
Freitag, J. H.	203
Fukushi, T.	157
Gandrup, J.	158
Garbowski, L.	158
Gardner, M. W.	158, 215
Ghimpu, V.	158
Giddings, N. J.	158
Gigante, R.	158
Gilbert, A. H.	159
Gladwin, F. E.	159
Godkin, J.	225
Goepp, K.	187
Goes, O. C.	159
Goidánich, G.	159
Gopalaiyengar, K.	218
Grainger, J.	160
Graber, L. F.	160
Gram, E.	160
Gratia, A.	160
Graves, C. E.	161
Green, D. E.	161
Grillo, H. V. S.	140
Gulley, A. G.	161
Gulyas, A.	161
Guttine, J. D.	225
Guyot, A. L.	161
Haasis, F. A.	204
Hall, F. H.	161
Hamilton, M. A.	162
Hansen, H. P.	162
Hansford, C. G.	162
Harrenveld, Ph van	162
Harris, R. V.	162
Harrison, A. L.	162
Hartish, J.	162
Hartung, W. J.	202
Heald, F. D.	163
Hibbard, P. L.	147
Hill, H. D.	163
Hino, I.	163
Hoadland, D. R.	147
Hoggan, I. A.	163, 166
Hollaender, A.	154
Holmes, F. O.	163
Hopkins, J. C.	164
Horne, W. T.	164
Hungerford, C. W.	164

	Pages
Hutchins, L. M.	164
Hyslop, G. R.	164
Imai, Y.	164
Ishikawa, T.	165
Iyengar, A. V. V.	165
Iyengar, K. G.	218
Jaggar, I. C.	165
Jivana Rao, P. S.	165
Jochens, S. C. J.	166
Johnson, H. W.	166
Johnson, J.	166, 169
Jöhnssen, A.	166
Jones, L. K.	167
Jordi, E.	167
Jorstad, I.	167
Karatchevsky, I. K.	167, 196
Kawamura, T.	168
Kendrick, J. B.	168
Keuchenius, P. E.	169
Keur, J. Y.	169
Klapp, E. L.	169
Klinkowski, M.	169, 181
Knowlton, G. F.	169
Koch, J.	169
Köch, K.	170
Köhler, E.	170
Kramer, S. P.	171
Kranzlin, G.	171
Kunkel, L. O.	171
Kuprewicz, V. F.	172
Küster, E.	172
Kuwana, S. L.	172
Lackey, C. F.	172
Lamy, R.	149
Lanschina, M. N.	213
Larter, L. N. H.	172
Lebard, P.	148
Lehman, S. G.	173
Lettoff, A. S.	173
Likhité, V. N.	173, 192
Linford, M. B.	173
Linz, C. V.	173
List, G. M.	174
Livingston, L. G.	154, 174
Loewenthal, H.	174
Loree, R. F.	174
Loughnane, J. B.	147
Ludwig, O.	174
Lushington, P. M.	175
Lutman, B. F.	175
MacClement, D.	175
Magrou, J.	148
Malherbe, I. de	175
Malhotra, R. C.	175
Mandelson, L. F.	175
Manil, P.	160, 161, 175
Manns, M. M.	176
Manns, T. F.	176
Manzoni, L.	177
Marchal, E.	177
Martin, D.	145
Martin, H.	136
Martin, J. N.	152
Martin, W. H.	177
Martinoff, S. I.	177
Martyn, E. B.	178
Massee, A. M.	178
Massey, R. E.	178
Matsumoto, T.	179
McAlister, D. F.	154
McBain, A.	137
McCarthy, C.	179
McClean, A. P. D.	179
McCormick, F. A.	148
McCubbin, W. A.	180
McKenny-Hughes, A. W.	180
McRae, W.	180
McKinney, H. H.	180
McWhorter, F. P.	180
Megaw, W. J.	181
Mejía, R.	181
Merkenschlager, F.	181
Meyer, H.	181
Mikhailova, P. V.	181, 197
M' Intossh, T. P.	182
Moore, E. S.	182
Morgenthaler, O.	182
Morris, O. M.	182
Morstatt, H. A.	183
Morwood, R. B.	183
Mottet, S.	184
Murata, T. J.	184
Murphy, P. A.	184

	Pages
Nattrass, R. M.	184
Neill, J. C.	184
Nelson, R.	185
Neuwiler, E.	185
Newton, W.	185
Nielson, O.	185
Noble, R. J.	185
Oefemia, G. O.	185
Ogilvie, L.	186
Oortwijn, Botjes, J. G.	187
Opitz, K.	187
Orton, C. R.	187
Orton, W. A.	187
Osborn, H. T.	187
Palm, B. T.	188
Pape, H.	188
Patch, E. M.	188
Park, M.	188
Parker, K. G.	194
Peacock, W. M.	189
Peltier, G. L.	189
Perret, C.	189
Pethybridge, G. H.	189
Petre, A. W.	189
Petri, L.	189
Piemeisel, R. L.	190
Pierce, W. H.	190
Pittman, H. S.	190
Pivovarova, R. M.	181
Plantegen, M. H. J.	190
Porter, D. R.	191
Price, W. C.	191
Purdy Beale, H.	138, 191, 225
Quanjer, H. M.	192
Rademacher, B.	194
Raleigh, W. P.	201
Rangel, E. S.	194
Rankin, W. H.	194
Rangaswami, S.	209
Rathsack, K.	187
Ravaz, L.	194
Rawling, T. E.	194, 213
Read, W. H.	195
Reddick, D.	195
Reed, H. S.	195
Reinmuth, E. F.	195
Riha, J.	195
Rischkov, V. L.	196
Robbins, W. J.	197
Roberts, J. I.	197
Rosa, J. T.	187
Rosenfeld, A. H.	197
Ross, A. F.	198
Ruhland, W.	198
Russell, T. A.	172
Salaman, R. M.	198
Sandford, G. B.	199
Sarker, B. N.	200
Sarrant, A.	200
Savastano, G.	200
Schapiro, S. M.	200
Schelesinger, M.	138
Schereven, D. A. van	200
Schultz, E. S.	200
Schweizer, J.	201
Schwing, E. A.	202
Serrano, L. A.	201
Servazzi, O.	201
Severin, H. H. P.	201
Shapiro, S. M.	204
Shapovalov, M.	154, 204
Sheffield, F. M. L.	205
Shull, J. M.	205
Silberschmidt, K.	205
Simmonds, J. H.	205
Slate Jr., W. L.	206
Smith, E. H.	206
Smith, J. H.	206
Smith, K. M.	206
Smith, L. B.	208
Smith, M. S.	208
Soltan, F.	187
Somazawa, K.	179
Sommer, H.	208
Sorauer, P.	208
Soukhoff, K. S.	208
Spencer, E. L.	208
Spennemann, F.	169
Sprague, V. G.	160
Sreenivasaya, M.	209
Ssuchov, P.	210
Stahel, G.	210
Stanley, W. M.	210

	Pages
Stevens, N. E.	211
Stevenson, F. J.	211
Stewart, F. C.	211
Stone, R. E.	212
Storey, H. H.	212
Stout, G. L.	213
Subramaniam, L. S.	180
Sukhoff, K. S.	213
Summers, E. A.	213
Sumner, C. B.	152
Sundararaman, S.	213
Swarbrick, T.	186
Takahashi, W. N.	213
Takata, K.	214
Tamm, E.	187
Tate, H. D.	152
Thiele, R.	214
Thomas, W. W.	201
Thompson, A.	214
Thompson, C. R.	186
Thornberry, H. H.	214
Thung, T. H.	192, 214
Tice, C.	215
Tims, E. C.	215
Tolas, A. G.	215
Tompkins, C. M.	158, 194, 215
Torres Filho, A.	216
Tropova, A. T.	216
Trotter, A.	216
Trümpener, E.	216
Tu, C.	216
Unite, J. O.	216
Uppal, B. N.	217
Valleau, W. D.	217
Varadaraja Iyengar, A. V.	218
Venkata Rao, M. G.	136, 218
Verhoeven, W. B. L.	219
Verplancke, G.	219
Verwoerd, L.	221
Vielwerth, V.	221
Vizioli, J.	221
Vos, H. C. C. A. A.	221
Wager, V. A.	182
Wallace, H. F.	221
Weger, F. W.	221
Went, F. A. F. C.	222
Weimer, J. L.	222
Wellman, F. L.	222
Werner, H. O.	223
Werwoerd, L.	223
Wetzel, K.	198
Whetzel, H. H.	223
Whipple, O. C.	158
White, P. R.	223
White, R. P.	224
Whitehead, T.	224
Wilcox, R. B.	224
Wiles, D. R. D.	224
Wilson, G. L.	224
Wingard, S. A.	225
Woods, M. W.	225
Woods, A. F.	225
Yenemaru, C.	225
Youden, W. J.	225
Zeijistra, F. H. H.	226
Zundel, G. L.	227
Zimmermann, A. von	226

INDEX

PARTIAL BIBLIOGRAPHY ON VIRUS DISEASES OF PLANTS

Journal of Agriculture of the University of Puerto Rico

Vol. XVIII, Nos. 1–2, p. 1–410, 1934.

Page

Abaca. See *Musa textilis*

Abelmoschus esculentus (okra) ---- 286

Abutilon

chlorosis ---- 69, 108, 138, 178, 205, 207

variegated ---- 94, 205, 309

Acer pseudoplatanus ---- 78

Aceratogallia sanguinolenta ---- 142

Acromania of cotton ---- 95

Acronecrosis

of potato ---- 318

(See also necrosis, net-necrosis, pseudo net-necrosis.)

Acropetal necrosis

of potato ---- 318

Adenoropium gossypifolium mosaic ---- 94

Aegeria exitiosa ---- 190

Aesculus (horse chestnut) ---- 175

Agallia stricticollis ---- 143

Agave cantala mosaic ---- 22

Ageing of virus *in vitro* ---- 177

Ageratum mosaic ---- 307

Aleurodidae ---- 8, 157, 160, 206, 217

Alfalfa. See *Medicago*

Alloiphyllie. See *Anemone*

Altheae rosea crinkle ---- 206

Altitude, effect of ---- 61, 97, 98, 99, 122, 222, 224, 235, 390

Amarillis sp.

mosaic ---- 95

Ambrosia trifida (virus disease) ---- 237

Amphoraphora rubi ---- 306

Ananas Ananas (Pine apple)

diseases ---- 223

mealy-bug ---- 81, 190

yellow spot ---- 81, 190, 229, 230

wilt ---- 81, 190

Anatomy

of mosaic tobacco plant ---- 164

Anemone ---- 207, 208

Anoectochilus ---- 262

Anthurium ---- 384

Antigens, antibodies, etc. ---- 165, 166, 240, 241, 297, 342

Antirrhinum Page
virus disease ---- 237, 384
Anuraphis tulipae ---- 170
Aphids ---- 7, 16, 22, 61, 64, 65, 108, 118, 121, 126, 180, 181, 186, 214, 238, 249, 250, 273, 284, 351, 362, 369, 377, 391, 400, 403
transmission of cucumber mosaic by ---- 117
migrations of *A. maidis* ---- 64
Aphis abreviata ---- 287
Aphis fabae ---- 59, 276, 323, 389
Aphis forbesi ---- 293
Aphis fragaefolia ---- 293
Aphis gossypii ---- 101, 117, 118, 169, 193, 392, 395
Aphis labruni ---- 68, 85
Aphis leguminosae ---- 68, 365, 366
Aphis maidis ---- 7, 22, 63, 64, 83, 84, 101, 102, 118, 172, 214, 238, 278, 288, 331, 357, 358, 364, 366, 367, 377, 403
Aphis rubiphila ---- 163, 305, 306, 346
Aphis rumicis ---- 61, 118, 140, 323
Aphis runisit ---- 389
Aphis spiraceae ---- 409
Apple. See *Malus*
Apricot. See *Prunus*
Arachis hypogea. (Peanut or ground nut) ---- 125, 264, 317, 409
clump disease ---- 370
dwarf ---- 174
leprosy ---- 377
mosaic ---- 139, 246
rosette ---- 17, 21, 24, 26, 61, 68, 85, 109, 139, 175, 213, 365, 366
streak ---- 139
virus disease ---- 125, 264
Armour expedition ---- 251
Asclepias spp. ---- 237
Asters. See *Callistephus*
Attenuation ---- 78, 79, 198, 222, 223
Avocado. See *Persea*
Azuki bean mosaic ---- 240

Bacteriophage ---- 166, 392, 404
Bacillus morulans of sugar beet ---- 58
Bacterium prodigiosum ---- 126
Bacterium ruber ---- 51
Balclutha. See *Cicadulina mbila*
Banana. See *Musa*
Barley. See *Hordeum*
Bean of all kinds. See *Phaseolus*
Beets. See *Beta*
Begonia ---- 280
Bemisia ---- 157, 206, 240, 367, 374

Page

Berry plants. See *Rubus*

Beta vulgaris ---- 7, 140, 230, 265, 277, 362, 379, 384, 386, 402

blight (western). See curly top

curly top ---- 7, 24, 25, 38, 58, 71, 78, 79, 80, 81, 82, 96, 102, 106, 108, 137, 142, 152, 170, 175, 201, 208, 209, 222, 228, 230, 234, 248, 260, 262, 266, 274, 276, 277, 307, 330, 332, 333, 334, 335, 336, 337, 351, 352, 355, 357, 358, 359, 370, 376, 384

curling ---- 109, 142, 274, 355, 402

immunity ---- 377

deformation ---- 60, 61

leaf curl ---- 142

leaf hopper (*Eutettix tenella*) ---- 13, 38, 78, 79, 81, 82, 108, 110, 145, 170, 173, 175, 208, 209, 332, 333, 334, 335, 336, 352, 357, 370

mosaic ---- 24, 25, 26, 27, 59, 61, 73, 121, 144, 182 201, 223, 228, 229, 262, 267, 268, 276, 277, 297, 314, 323, 326, 340, 341, 343

phloem necrosis ---- 267

virus disease ---- 61, 386

yellows ---- 377

Bidens pilosa

yellow spot ---- 230

Big bud

of tomatoes ---- 321

of currants ---- 227

Bitter pit. (See *Malus* and *Pyrus*)

Blackberry. See *Rubus*

Black locust. See *Pseudacacia.*

Blight (virus forms only). See *Beta, Spinaceae, Lycopersicon esculentum*, etc.

Bodies of various kinds ---- 37, 49, 65, 102, 116, 153, 160, 161, 180, 183, 204, 207, 208, 211, 215, 217, 218, 222, 228, 229, 235, 242, 250, 272, 273, 285, 323, 339, 346, 347, 348, 384, 405.

Bougainvilea spectabilis virus disease ---- 237

Brambles. See *Rubus*

Branchysm. (See *Gossypium*)

Brassica (cabagge, mustard, turnips) ---- 86, 196, 327, 340, 369, 372

Breaking of tulips. (See Tulips).

Breviocoryne brassicae ---- 86, 102

Browallia speciosa mayor ring spot ---- 280

Brown spot of potato ---- 35

Browning (stem end) ---- 119

Buckskin of cherry. (See *Prunus*)

Buckwheat. (See *Fagopyrum*)

Bullet disease of *Passiflora* ---- 274

Bunchy top. See *Musa, Carica, Helichrysum, Physalis & Lycopersicon*

	Page
Cabbage. See *Brassica*	
Cactaceae	286
Cajanus indicus	261
Cainito rosette	68
Calendula sp. (rosette)	68
Calendula officinalis (virus disease)	51
Calico	
of potato	294
of tobacco	88, 158, 179, 369
Calliopsis rosette	68
Callistephus chinensis (asters)	
chlorosis	88
curly top	237
wilt	202, 234
wire screen control	219
yellows	113, 153, 175, 202, 218, 219, 220, 234, 237, 244, 246, 279, 335, 351
yellows of tomato	219
Calocoris bipunctatus	237, 270
Campanula pyramidalis	280
Campanulales (virus disease)	382
Caña brava. See *Gynerium saggitatum*	
Cane. See *Saccharum officinarum*	
Cantaloupe ring spot	382
Capsicum annuum	156, 330, 379
leaf curl	189, 190, 255, 256
mosaic	56, 95, 125, 184, 256, 379
virus	125
Carica papaya	
bunchy top	94, 95
mosaic	94, 95
Carnation. See *Dianthus*	
Carrot. See *Daucus carota*	
Cassava. See *Manihot*	
Catnip. See *Nepeta*	
Cauliflower. See *Brassica*	
Celastrus scandens	89
Celery	
mosaic	117, 294, 395
yellows	175, 220, 335
Cell contents	
effect of mosaic	92, 93
function	161
inhibition by virus	95
structure affected by mosaic	93, 94, 95, 124, 125
Cell inclusions. (See Bodies)	
Cephalanthus occidentalis	379
Ceratoma trifurcata	344

Page

Chemistry ---- 37, 38, 41, 57, 58, 65, 66, 71, 76, 89, 90, 102, 108, 128, 138, 139, 145, 147, 152, 153, 157, 196, 231, 235, 240, 266, 272, 283, 307, 314, 325, 338, 341, 345, 346, 354, 355, 356, 358, 369, 370, 378, 380, 387, 388, 389, 392, 405

Chenopodium murale curly top ---- 78, 223

Cherry. See *Prunus*

Chicory. See *Cichorium*

Cichorium mosaic ---- 237

Chlamydozoa ---- 231, 235, 285

Chlorophyll ---- 90, 128, 139

Chlorophyllase ---- 290

Chloroplasts ---- 93, 94, 95, 273, 354

Chlorosis ---- 88, 108, 131, 138, 139, 168, 228, 260, 274, 284, 392, 397, 405

- of *Abutilon* ---- 108, 138, 178
- of Asters ---- 88
- of avocado ---- 186
- of cane ---- 131, 143, 264, 274, 364
- of beans ---- 88, 139
- of citrus ---- 377
- of *Celastrus scandens* ---- 89
- of Cucurbit ---- 185
- of *Evonymus* ---- 313
- of *Fraxinus* ---- 43
- of *Laburnum* sp. ---- 42, 43
- of Lupin ---- 309, 326
- of *Ligustrum* ---- 42, 43
- of *Malvaceae* ---- 42, 178
- of peach ---- 88
- of *Ptelea* ---- 43
- of *Ribes vulgaris* ---- 89
- of Rose ---- 260, 261, 273, 397
- of *Rumex* ---- 163
- of *Soja Max* ---- 89
- of *Sorbus* ---- 43
- of Sweet potato ---- 315
- of tobacco ---- 88, 168
- of tomato ---- 311
- of *Zea Mays* ---- 109, 340

Chlorotic

- diseases of hops ---- 319, 320
- spot of rose ---- 105
- streak of cane ---- 238

Chrysanthemum yellows ---- 8, 273

Chrysophyllum rosette ---- 68

Cicadulina mbila ---- 296, 364, 365, 366, 367

Cicadula sexnotata ---- 113, 118, 175, 218, 220, 335, 336

Cicadula divisa ---- 336

Page
Cichorium intybus mosaic ---- 237
Cineraria mosaic ---- 111
Citrus
chlorosis ---- 377
psorosis ---- 144, 236
variegations ---- 292
Classification ---- 73, 136, 166, 198, 199, 302, 347, 366
Clerodendron fragrans mosaic ---- 54
Climate ---- 166, 396
Clover. See *Trifolium*
Club leaf. See *Gossypium*
Coffea phloemnecrosis ---- 19, 170, 356, 357
Coix Lachryma-Jobi ---- 64, 214, 238
Commelina ---- 94, 117, 395
Compositae ---- 62
"Contagium vivum fluidum" ---- 45, 46
Convallaria majalis mosaic ---- 54
Corn. See *Zea Mays*
Cotton. See *Gossypium*
Cowpea. See *Vigna*
Cranberry. See *Vaccinium*
Crinkle dwarf. See *Nicotiana tabacum*
Crinkle ---- 42
of *Althea rosea* ---- 206
of cherry ---- 25
of cotton ---- 8
of potato ---- 270, 317
of strawberry ---- 383, 409
of tobacco ---- 374, 375
Crocus virus disease ---- 170
Crotalaria striata mosaic ---- 94, 95
Crotalaria juncea virus ---- 154
Crucifer
mosaic ---- 86, 165, 372
virus disease ---- 27
Cuban streak of cane ---- 296
Cucumber. See *Cucumis sativa*
Cucumis sativa
mosaic ---- 13, 25, 50, 85, 86, 96, 102, 105, 115, 116, 117, 135, 159, 167, 181, 182, 185, 193, 198, 199, 204, 224, 246, 256, 261, 267, 294, 295, 321, 331, 341, 375, 387, 391
virus disease ---- 212
white pickle ---- 88, 96, 193, 256
yellow pickle ---- 375
Cucurbitacea mosaic ---- 106, 134
virus ---- 382
Cucurbit ---- 115, 116, 185, 193, 267, 391, 340

Page
Cultures
of virus ---- 161, 163, 266, 297
of insects ---- 171
Curl
of beets ---- 109, 274
of cotton ---- 8, 141, 160, 228
of *Pelargonium* ---- 154, 326
of potato. See *Solanum tuberosum*
of Poinsettia ---- 285
of raspberry ---- 47, 48, 112, 256
(See leaf curl of raspberry)
of *Rumex* acetosa ---- 402
of *Sesame* ---- 392
of tobacco ---- 61, 375
of tomato ---- 359
Curly dwarf of potato. (See *Solanum tuberosum*)
Curly top
of *Callistephus chinensis* ---- 237
of *Chenopodium murale* ---- 78, 223
of aster ---- 237
of beans ---- 79, 234
of beets (sugar). See *Beta*
of cucurbits ---- 106
of squash ---- 248
of tomato ---- 338, 392
Currants. See *Ribes*
Cyphomancha betavea ---- 256
Cytology ---- 122, 123, 125, 126, 176, 180, 229, 346, 378
of *Anemone* ---- 207
of bean mosaic ---- 273
of beets ---- 137, 307, 343
of Fiji disease of cane ---- 218, 237
of *Hippeastrum* ---- 183
of insects ---- 113
of mosaic ---- 92, 123, 160
of mosaic of sugar cane ---- 65, 91, 92
of pecan rosette ---- 304
of potato mosaic ---- 347, 348
of potato spindle tuber ---- 384, 385
of potato virus disease ---- 87, 347, 384
of *Santalun Album* ---- 271
of spike ---- 272
of sugar beet ---- 307, 343
of sugar cane ---- 91, 92, 218
of tomato mosaic ---- 354
of tobacco ---- 160, 290, 307, 378, 405
of virus ---- 122, 124, 125, 176, 180, 207, 229
of wheat ---- 251

Page

Daffodil ---- 107, 167
Dahlia
mosaic, dwarf and stunt ---- 62, 67, 161, 237, 280, 307
Dandelion (fall) yellow disease ---- 264
Datura (Jimson weed) ---- 53, 74, 184, 256, 262, 320, 329, 349, 358
Daucus carota yellows ---- 60, 220, 235
Degeneration. See *Solanum tuberosum*
Delphacodes steriatellus ---- 220
Delphinium
stunt A ---- 73
virus disease ---- 176, 189, 237, 382, 397
Desication of potato tuber ---- 38
Dewberry. See *Rubus*
Dianthus (Carnation)
bacteriosis of carnation ---- 32
yellows ---- 223, 287, 288
Digitaria horizontalis (streak of cane) ---- 356, 366
Dilution of tobacco virus ---- 10, 251
Disonycha triangularis ---- 162
Distribution of sugar-cane disease ---- 93
Dock. See *Rumex*
Dodonacea viscosa spike ---- 322
Dolichos biflorus mosaic ---- 379
Dolichos lablab mosaic ---- 379
Dwarf
of barley ---- 14
of blackberry ---- 408
of cane ---- 46
of corn ---- 397
of horse radish ---- 106
of logan berry ---- 408
of mulberry ---- 190, 370, 408
of onion ---- 118, 177, 257
of peanut ---- 174
of potato. See curly dwarf of *Solanum tuberosum*
of rice ---- 153, 185
of tomato ---- 228
of tobacco ---- 268

Egg plant. See *Solanum Melongena*
Electricity ---- 45, 58, 281
Electrophorosis ---- 371
Eleusine indica streak ---- 214, 365, 366
Elm ---- 304
Elytrosomos ---- 228
Emilia ---- 230
Empoasca ---- 8, 87, 165, 206, 274, 352
Enzymes ---- 28, 231, 405
Epiphyllum spp. mosaic ---- 55, 286
Epiterix cucumeris ---- 87, 162

Page

Equisetale mosaic (fossil) ---- 171
Eradication ---- 18, 244, 277, 284, 296, 379, 386, 387
of bunchy top of banana ---- 132
of cane mosaic ---- 40, 129, 130, 131, 141, 152
of celery mosaic ---- 117
of cucumber mosaic ---- 116
Erigeron yellows ---- 246
Eriophyes ribis ---- 12, 227
Erythroneura Zealandica ---- 100
Eucharis amazonica mosaic ---- 95
Euphorbia pulcherrima ---- 285
Evonymous sp. ---- 43, 108, 313
Euscelis striatulus ---- 45, 113, 401
Eutettix tenella ---- 58, 79, 80, 81, 82, 108, 110, 152, 170,
(See also *Beta vulgaris* leafhopper) 173, 208, 209, 330, 332, 333, 334, 335, 336, 337, 338, 352, 357, 370

Fagopyrum esculentum (aster yellows) ---- 218
False blossom. See *Vaccinium*
Fern leaf
of tobacco ---- 261
of tomato ---- 132, 216, 256, 261
Ficus Carica mosaic (fig) ---- 90, 91
Fiji. See *Saccharum officinarum*
Filiform leaf of tomato ---- 330
Fillicosi ---- 105
"Filosité" of potato ---- 15, 121, 286, 289
Filters, filtration, etc. ---- 9, 51, 62, 81, 126, 146, 147, 153, 157, 215, 216, 219, 230, 234, 235, 236, 250, 252, 263, 266, 307, 313, 314, 335, 350, 353, 358, 367, 370, 384, 386, 387, 390
Flagellates ---- 49
"Flenkenkrankheit" ---- 46
Fragaria (strawberry) ---- 239, 293, 294, 383, 408, 409
crinkle ---- 383, 409
degeneration ---- 293
dwarf ---- 293
Gold disease ---- 168
mosaic ---- 48, 49, 168
stunting ---- 168
virus ---- 173, 294
witches' broom ---- 408
yellow edge ---- 173, 293
yellows (June) ---- 293, 294
xanthosis ---- 293
Frankliniella sp. ---- 262, 320, 321
Fraxinus chlorosis ---- 43
Freesia virus disease ---- 170

	Page
Frenching of tobacco	380
Fuchsia virus disease	237
"Gelbflechigheit" of spinach	62
"Gele strepenziekte". (See yellow stripe of sugar cane)	
Geographic distribution of sugar-cane diseases	93
Giant hill of potato. (See also *Solanum tuberosum*)	158, 159
Gislkia mosaic	307
Gladiolus	
virus disease	118, 170, 238
Gossypium	
acromania	95
brachysum	95
club leaf	95, 406
crazy top	95, 205
crinkle	8
curl	8, 141, 160, 189, 200, 228
cyrtosis	95, 406
leaf cut	95, 204, 372
leaf crinkle	206
leaf curl	8, 141, 160, 189, 200, 206, 228, 239, 240
leaf roll	200
malformation	95
tomotosis	95, 204
Grape. See *Vitis*	
Grass	391
mosaic	62, 63, 217, 226, 278, 330, 366
yellow stripes	226, 363
Grasshopper	162
Greenhouse	
mosaic in	116, 117, 165
streak in	200
yellows in	273
virus in	212
Ground nut. See *Arachis hypogea*	
Gynerium saggitatum	7
Haemaria	262
Helichrysum bunchy top	218
Hemiptera	348
Hemp (Manila). See *Musa textilis*	
Hibiscus cannabinus	206
Hibiscus esculentus	206
Hibiscus rosa-sinensis	237
Hibiscus sabdariffa	206
Hicoria. (Pecan rosette)	9, 110, 145, 253, 283, 304, 343
Hippeastrum	
bodies in	183, 217, 250
mosaic	183, 217, 279
virus disease	235

Page

Hippeastrum Johnsonii mosaic ---- 250
Histology ---- 93, 122, 123, 124, 125, 176, 186, 230, 231, 280, 314, 346, 405
of alfalfa ---- 165
of beet ---- 266
of cane ---- 92
of coffee ---- 357
of dahlia ---- 161
of fiji disease ---- 218
of *Hippeastrum* ---- 183
of peach ---- 91
of potato ---- 33, 43, 66, 378, 384, 385
of *Solanaceae* ---- 180
of sugar cane ---- 92, 218
of tobacco ---- 160
of tomato ---- 156
Holodiscus discolor, witches' broom ---- 409
Homoptera ---- 271
Hops. See *Humulus*
Hordeae ---- 252
Hordeum
barley dwarf disease ---- 14
barley stripe disease ---- 371
Horse chestnut. See *Aesculus*
Horse radish. See *Radicula armoracia*
Hot water treatment ---- 56, 63, 221, 401
Humulus **(Hops)**
chlorotic diseases ---- 319, 320
"Kaderavost" ---- 53
leaf roll ---- 206
mosaic ---- 85, 234, 318, 319, 374
Nettle head ---- 122
virus ---- 54, 318
Hunchback of tobacco ---- 354
Hyacinth ---- 279
Hydrastis canadensis virus diseases ---- 237
Hydrogen-ion ---- 55, 174
Hyoscyamus
mosaic ---- 184, 256, 339
virus ---- 171
Hypochaeris radicata mosaic ---- 285

Ilex ---- 78
Illionia solanifolia (Macrosiphum gei) ---- 249
Immunity & immunology ---- 18, 24, 51, 100, 131, 179, 205, 220, 241, 283, 297, 309, 342, 377, 387
Inactivation of virus ---- 32, 38, 281
Incubation ---- 213, 218, 332, 338

Page

Infection by stomata, tobacco 127, 196
Infra-red photography 44
Inhibitory action of mosaic 94, 95, 135
Inoculation 116, 134, 162, 201, 219, 246
  mechanics of 63
  Internal brown spot 35
Insects. (See also latin names) 8, 10, 13, 19, 22, 23, 38, 39, 41, 44, 47, 51, 58, 60, 61, 62, 63, 64, 65, 67, 68, 79, 80, 81, 82, 83, 84, 85, 86, 87, 97, 100, 101, 102, 108, 110, 112, 113, 118, 126, 129, 132, 134, 135, 136, 139, 140, 145, 152, 157, 159, 160, 162, 163, 165, 169, 170, 171, 172, 173, 175, 180, 184, 185, 190, 193, 206, 208, 209, 212, 217, 218, 220, 221, 227, 229, 230, 232, 235, 238, 239, 240, 246, 248, 249, 250, 261, 262, 267, 269, 270, 271, 273, 274, 276, 277, 278, 280, 284, 285, 288, 293, 294, 296, 297, 304, 305, 306, 314, 320, 321, 323, 324, 327, 330, 331, 332, 333, 334, 335, 336, 337, 338, 344, 346, 348, 350, 351, 352, 357, 358, 362, 363, 364, 365, 366, 367, 369, 370, 371, 372, 373, 374, 381, 384, 389, 399, 402, 403, 407, 408, 409
Insects
  cultures of insects 171
  eggs of 154
  transmission. See transmission.
Intracellular bodies. See bodies.
Ipomoea
  chlorosis 315
  mosaic 40, 72, 94, 136, 174, 272, 315, 393, 394
Iris 67, 154, 170, 238, 397

Jimson weed. See *Datura*

"Kaderovost" of hops 55
Kassoer cane 109
Katahdin 28
Kavangerie cane 63, 103, 104, 131, 243, 312, 315, 377
"Kringerigheit" of potato 205, 301
"Kromnek" or "Kat" disease
  of tobacco 262
  of tomato 262
"Kroepoek" (leaf curl)
  of tobacco 205
  of *Zinnia elegans* 205
"Kroepoek-Ziekte" of tobacco 375

Page
Laburnum sp. chlorosis 42, 43
Laetuca sativa
mosaic 62, 106, 193, 279, 307, 335
río grande disease 77, 218
white heart 218
yellows 101, 335
Lagerstroemia rosette 68
Lantana camara, spike 146, 180
Lathyrus odoratus mosaic 116
Latent virus of potato 201, 212
Leaf blotch of potato 185
Leaf crinkle of cotton 206
Leaf curl
of beet 142
of tobacco 185, 205, 367
of cassava 409
of cotton 141, 160, 189, 200, 228, 239, 240
of Poinsettia 285
of potato 76, 100, 185, 239, 261
of raspberry 166, 180, 305, 346
of *Zinnia* 205, 240
Leaf cut of cotton 95, 204, 372
Leaf deformity of *Rumex* 61, 163
Leaf hopper
of alfalfa 165, 200
of asters 218
of beets. See *Beta vulgaris*
of cane 278
of clover 182
of corn 367
of cranberry 44, 113
of potato 224
of sugar cane 278, 364, 367
Leaf roll. (See *Beta vulgaris*) 204
of cotton 200
of hops 206
of potato. See *Solanum tuberosum*
of tomato 168
of the vine 291
of weeds 129
Leaf Spot of tobacco 45, 46, 94, 168
Leconium corni mosaic 369
Legume mosaic 116, 134, 240
Leontodon autumnale yellow disease 264
Leprosy
of manioc 287
of peanut 377
Leptomonas in mosaic 273
Leptinotarse decemlineata 162

Page
Leptonecrosis of potato ---- 147, 298
Lesions
tobacco ---- 183, 194
bean ---- 295
tomato ---- 220
Lettuce. See *Lactuca sativa*
Light ---- 32, 83, 127, 183, 196, 266
Ligustrum chlorosis ---- 42, 43
Lilium ---- 279
mosaic ---- 101, 169, 176, 244, 279, 280
yellows flat ---- 176
Lily ---- 167, 168, 169, 176, 368, 405
virus disease ---- 170, 279, 280, 361, 368
of the valley mosaic ---- 54
Lima bean. See *Phaseolus lunatus*
Lithiasis of pear ---- 77
Little leaf
of apple ---- 175
of avocado ---- 186
of cherry ---- 175
of peach ---- 175
of walnut ---- 169
Little peach. See *Prunus*
Locheria rosea virus disease ---- 110
Logan berry. See *Rubus*
Longevity of virus ---- 292
Locust (black). See *Robina Pseudacacia*
Lupin chlorosis ---- 309, 326
Lycium mosaic ---- 184
Lycopersicon esculentum ---- 50, 189, 197, 272, 338
blight (western) ---- 177, 188, 228, 237, 248, 314, 336, 337, 373, 383
blight (winter) ---- 37, 186, 188, 283, 363
bunchy top ---- 245
chlorosis ---- 311
curling ---- 359
curly top ---- 338, 392
dwarf ---- 228
fern leaf ---- 132, 216, 261
filiform leaf ---- 330
histology ---- 156
"kromnek" or "kat" ---- 262
leaf curl ---- 189
leaf roll ---- 168, 243
lesion ---- 220
mosaic ---- 7, 8, 9, 11, 13, 24, 49, 50, 57, 58, 66, 67, 74, 75 82, 87, 112, 132, 136, 156, 169, 178, 179, 216, 220, 234, 247, 248, 253, 256, 276, 295 297, 298, 308, 311, 329, 346, 347, 354, 368, 371, 373, 386, 396, 389

Lycopersicon esculentum—Continued. Page
rosette 321
streak 9, 25, 48, 49, 56, 117, 175, 194, 200, 201, 252, 320, 338, 347, 368, 381, 382, 383
stripe 8, 9, 382
spotted wilt 8, 38, 67, 76, 117, 171, 184, 275, 280, 293, 320, 321, 342, 350
ring spot 56, 350
virus disease 66, 72, 157, 212, 247, 303, 320
wiry 228
witches' broom 407
yellows 337, 338
yellows (aster) 219
yellows (psyllid) 56, 311
Lygus 162, 232

Macropsis trimaculata 220
Macrosiphum sp. 59, 323
Macrosiphum gei 87, 100, 118, 126, 170, 249, 250, 274, 399
Macrosiphum pisi 284
Macrosiphum pseudosolani 270
Macrosiphum solanifolli 87, 140, 181, 246, 270, 273, 274, 287, 327, 351
Macrosiphum tabaci 10
Macrosiphum tussilaginis 285
Malformation of cotton. (See *Gossypium*)
Malus Malus
bitter pit 11, 36, 68, 77, 178, 244, 344, 400
little leaf 175
rosette 175
stignomose 68
variegation 62
Malvaceae
chlorosis 42, 43, 178
Mandioca mosaic. (See also *Manihot*) 49, 125, 369
Manihot. (See also Mandioca and Manioc) 105, 109, 200, 217, 265, 369
Manila hemp. See *Musa textilis*
Manioc, mosaic. (See also *Manihot* and Mandioca) 61, 123, 157, 176, 200, 217, 287, 358, 409
leprosy 287
Manioks mosaic. (See also *Manihot*) 149, 157, 409, 410
leaf curl 409
Marytynia mosaic 184
Masking 174, 184, 185
Masking raspberry mosaic. See *Rubus*
Matizado. See mosaic of *Saccharum officinarum*
Mealy bug of pineapple 81, 190
Medicago Alfalfa 134, 165, 200, 394
Melanphthalma gibbosa 100
Melanoplus 162

Melilotus alba Page
mosaic ---- 134, 252
ring spot ---- 144, 177
Melon mosaic ---- 116
Metabolism ---- 41, 75, 96
of cabbage ---- 196
of spinach ---- 196
Micrampelis lobata mosaic ---- 115
Micro-chemistry. See Chemistry
Mitochondria ---- 123
Moron of potato ---- 267
Morus. Mulberry
dwarfing ---- 190, 370, 408
mottling ---- 94
variegations ---- 94
Mosaic ---- 10, 12, 18, 20, 21, 24, 28, 62, 66, 82, 88, 111, 112, 113, 121, 127, 152, 165, 173, 178, 230, 247, 252, 256, 257, 260, 281, 310, 344, 346, 347, 390, 392, 393
causes of ---- 68
effects on cells ---- 92, 93, 95, 124, 125
in vitro ---- 28, 252, 281
inactivation of. See inactivation
inhibition by ---- 95
masking of ---- 145, 184, 362, 401
movement of ---- 184, 321
overwintering of. See overwintering
of *Adenoropium gossypifolium* ---- 94
of *Agave* ---- 22
of *Alfalfa* ---- 394
of *Amarillis* ---- 95
of *Anemone* ---- 207, 208
of Aucuba (or aukuba) ---- 50, 57, 74, 75, 135, 136, 220
of bean. (See *Phaseolus vulgaris*)
of Azuki bean ---- 240
of broad bean ---- 153
of lima bean ---- 246
of beets. See *Beta*
of Brassica. See *Brassica*
of bulb plants ---- 36
of Cabbage (chinese) ---- 327, 372
of *Cactaceae* ---- 286
of Cane. See *Saccharum officinarum*
of *Capsicum annuum* ---- 55, 379
of cassava ---- 61, 105, 157, 217, 265, 358, 367
of catnip ---- 267
of celery ---- 117, 294, 395
of *Cichorium intybus* ---- 237
of *Clerodendron fragrans* ---- 54
of clover (red) ---- 134

Mosaic—Continued. Page
of clover (sweet) 134, 252
of *Commelina longicaulis* 94
of *Commelina nudiflora* 117, 395
of *Compositeae* 62
of *Coix lachryma-joli* 64
of *Convolvullaria majalis* 54
of corn 63, 217, 219, 314, 315, 358, 363, 364
of cow pea 344
of *Crotalaria striata* 94, 95
of Crucifer 86, 165
of Cucumber. See *Cucumis sativa*
of Cucurbit. See cucurbit
of *Cucurbitaceae* 134
of dahlia mosaic 67, 161, 237, 307
of *Dolichos biflorus* 379
of *Dolichos lablab* 379
of dew berry 47
of egg plant 286
of *Epiphyllum* 55, 286
of Equisetale (fossil) 171
of *Eucharis amazonica* 95
of fig 90, 91
of gladiolus 118
of grasses. (See Grass).
of greenhouse 116, 117, 165
of *Hippeastrum* 183, 217
of *Hippeastrum Johnsonii* 250
of hops. See *Humulus*
of horse radish 106
of *Hyoscyamus* 184, 256, 339
of *Hypochaeris radicata* 285
of Katahdin (potato) 28
of *Lathyrus adoratus* 116
of *Leconium corni* 369
of legumes 116, 134, 240
of leptomonas 273
of lettuce 133, 193, 274, 307
of *Lilium* sp. 169
of lily 101, 169, 176, 280
of lily of the valley 54
of logan berry 408
of *Lycium* 184
of *Lycopersicon*. See *L. esculentum*
of *Manihot appi* 105, 217
of *Manihot dulcis* 200
of *Manihot glaziovii* 109, 217
of *Manihot palmata* 369
of *Manihot utilissima* 200
of manioks 149

Mosaic—Continued. Page
of mandioca 369
of manioc 123, 287, 358
of *Marytinia* 184
of *Medicago Arabica* 134
of *Melilotus alba* 134
of *Micrampelis lobata* 115
of muskmelon 167
of mustard 327
of narcissus 255
of *Nicandra physaloides* 144
of *Nicotiana glutinosa*. See also *Nicotiana* 391
of *Pachyrhizus* 140
of *Panicum* 102
of pea 54, 116, 284, 353
of peach 190
of peanut 139, 246
of pear 287
of pepper 330, 379
of Petunia 11
of *Phyllocactus gaertneri* 286
of *Physalis* sp. 102, 156, 262, 391
of *Physalis peruviana* 340
of *Phytolacca decandra* 11
of *Pisum sativum* 54, 116
of potato. See *Solanum tuberosum*
of *Prunella vulgaris* 231
of raspberry. See *Rubus*
of *Rhododendron* 285
of rhubarb 112
of *Rhipsalis rosea* 286
of roses 254, 394
of rugose of potato 198 210
of rugose of weeds 129, 380
of rye 251
of sandal (leaf curl) 382
of *Setaria aurea* 102
of *Sida* 94, 219
of *Sinamar* 140
of *Solanum nodiflorum* 346
of soy bean 155, 205
of spinach 62, 323, 389, 400
of strawberry 48, 49, 168
of sweet potato 136, 174, 272, 315, 394
of *Syntherisma* 288
of tobacco. See *Nicotiana tabacum*
of tomato. See *Lycopersicon esculentum*
of tropical plants 125
of tulips 125, 248, 249
of turnip 156

Mosaic—Continued. Page
of *Vicia faba* 134
of vine (grape) 285, 368
of watermelon 391, 392
of wheat 123, 251, 252, 256, 383, 392
of weeds 129
of wild hosts of sugar-cane mosaic 63
properties of 10
rare case of, in sugar cane 406
temperature studies of 112
yellow mosaic of wheat 252
yellow mosaic of tomato 50, 57
yellow mosaic of tobacco 126, 195
Mosaico. See Mosaic
"Mosaikfäule". See Mosaic
"Mosaikkrankheit". See Mosaic
Mottling disease. See also mosaic 325
of mulberry 94
of *Evonymous* 313
of sugar cane 86, 132, 145, 360, 361, 376
of tomato 132
Mulch (straw) 159
Mulberry. See *Morus*
Musa (banana)
bunchy top 16, 17, 20, 22, 23, 71, 74, 107, 132, 160, 235, 273, 275, 286, 332, 342, 392, 410
mosaic 279
Musa textilis (Abacá)
bunchy top 23, 25, 36, 71, 75, 76, 155, 182, 190, 256, 257, 277, 278, 286, 344, 373
mosaic 22
Muskmelon mosaic 106, 167
Mustard, mosaic 327
Mycetozoa of tobacco 202
Myzus circumflexus 181, 232, 249, 399
Myzus fragaefolii 293, 399, 408
Myzus pelargonii 249
Myzus persicae 11, 41, 61, 67, 87, 100, 126, 129, 140, 159, 170, 171, 180, 181, 212, 232, 248, 249, 261, 270, 294, 301, 314, 324, 327, 348, 373, 381, 384, 389, 399
Myzus pseudosolani 100, 181, 270
Myzus solani 232

Narcissus 180, 248
grey disease 255
mosaic 255
virus 170

Page

Necrobiosis of potato ---- 65
Necrosis. See also acronecrosis, netnecrosis, pseudonet necrosis
of potato ---- 35, 43, 65, 137, 138, 147, 149, 159, 205, 281, 298, 317, 318, 324, 325, 328, 329, 332, 378
of strawberry ---- 168
of tobacco ---- 51
Nepeta mosaic ---- 267
Net necrosis. See also acronecrosis, necrosis, pseudonetnecrosis ---- 135, 149, 159, 301, 328
frost ---- 201
of potato ---- 35
of tomato ---- 303
acronecrosis ---- 43, 318
acropetal necrosis ---- 43
leaf roll ---- 43
leptonecrosis ---- 298
necrobiosis ---- 65
phloemnecrosis ---- 19, 170, 356, 357
pseudonecrosis ---- 205
top necrosis ---- 282, 283, 301
Nephotettix apicalis ---- 154, 372
Nettle head of hops ---- 122
Nettle leaf on currant ---- 227
Nicandra mosaic ---- 144
Nicandra physalodes ---- 56, 144, 262
Nicotiana angustifolia ---- 195
Nicotiana glauca ---- 11, 374
Nicotiana glutinosa ---- 56, 74, 75, 183, 321, 405
mosaic of ---- 74, 391
Nicotiana macrophyla ---- 61
Nicotiana paniculata ---- 194
Nicotiana rustica ---- 61, 374, 405
Nicotiana sylvestris ---- 57, 220
Nicotiana tabacum
calico ---- 88, 158, 179, 187, 369
chlorosis ---- 88, 163
"contagium vivum fluidum" ---- 45, 46
crinkle ---- 263, 374, 375
crinkle dwarf ---- 263
curl ---- 61, 367, 375
dwarf ---- 263
enation ---- 194
fern leaf ---- 261
frenching ---- 380
hunchback ---- 354
kroepoek ---- 205
"kroepoek-ziekte" ---- 374, 375
kromnek or kat ---- 262
leaf curl ---- 185, 205, 367

Nicotiana tabacum—Continued. Page
leaf spot ---- 45, 46, 94, 168, 185
mosaic ---- 10, 11, 13, 18, 23, 25, 32, 41, 44, 45, 51, 60, 71, 73, 82, 83, 88, 89, 94, 96, 104, 110, 112, 117, 119, 122, 123, 126, 127, 128, 136, 139, 150, 152, 153, 154, 158, 160, 161, 163, 164, 165, 166, 168, 174, 176, 179, 180, 183, 184, 185, 187, 188, 191, 194, 195, 196, 197, 198, 208, 213, 220, 223, 229, 231, 233, 236, 240, 241, 243, 244, 251, 252, 253, 261, 262, 264, 265, 266, 275, 276, 280, 281, 285, 288, 290, 295, 297, 298, 302, 309, 307, 321, 324, 325, 330, 339, 341, 353, 358, 360, 361, 369, 371, 372, 374, 375, 377, 379, 380, 381, 383, 387, 388, 391, 397, 404, 405
Mycetozoa ---- 202
"Nielle" of tobacco ---- 45, 62, 110, 288
Plasmodiophora tabaci ---- 202, 230
"pochenkrankheit" ---- 110, 191
powdery of ---- 354
ring spot ---- 57, 144, 152, 158, 177, 182, 193, 195, 199, 295, 296, 382, 402, 405
Rotterdam B-disease ---- 195
spot ---- 124, 191, 213
necrosis ---- 51, 199
streak ---- 117, 347
stripe ---- 61
string leaf ---- 88
veinbanding ---- 158
virus diseases ---- 27, 28, 61, 72, 125, 157, 158, 181, 183, 184, 196, 197, 198, 215, 232, 251, 297, 303, 330, 367, 374, 377, 380, 381, 382, 392
white ---- 122
witches' broom ---- 407
yellow mosaic of tobacco ---- 126, 195
Nicotiana tomentosa ---- 94
Nicotiana viscosa ---- 11
Northiella sacchari ---- 23

Ocean spray witches' broom ---- 409
Okra. See *Abelmuschus esculentus*
Orchids ---- 262
Onion ---- 294
dwarf ---- 88, 118, 119, 177, 257
Ophiola striatula ---- 48
Organisms minute ---- 49
ultramicroscopic ---- 157

Oryza sativus Page
dwarf 153
insects 154, 372
stripe 14, 220
stunt 372
Oxalis stricta 359
Oxidases 71
Oxygenation 87
Overwintering 246, 287, 380, 391
aster yellows 218
cucumber mosaic 115, 116
mosaic of physalis 102
spotted wilt of tomato 171
(See also Spotted wilt of tomato)
tomato mosaic 156
potato mosaic 300
tobacco mosaic 198, 246

Pachyrhizus sp. mosaic 140
Panicum colonum 102
Papilonaceae 259
Para-crinkle of potato 317
Paratriozoa cockerelli 51, 139, 311
Parsley yellows 335, 336
Parsnip yellows 336
Passiflora sp.
bullet disease 274
virus 51
woodiness 274, 275
Pea. See *Pisum sativum*
Peanut. See *Arachis*
Peach. See *Prunus*
Pear see *Pyrus*
Pecan. See *Hicoria*
Pelargonium 43, 237, 285, 290, 385
curl 154, 326
"Pomme de terre". See *Solanum tuberosum*
Pentalonia nigronervosa 159, 190, 235, 277
Peonias 397
Pepo moschata (Pumpkin and squash) 106, 248
Pepper. See *Capsicum*
Peregrinus maidis 67, 357, 358, 367
Perkinsiella vastatrix 278, 279
Perkinsiella saccharicida 267, 278
Persea (avocado)
little leaf 186
chlorosis 186
sun blotch 89, 185, 186, 286

Petunia sp. Page
mosaic 11, 198
ring spot 177, 382, 405
rosette 68
Phaseolus (bean)
curly top 234
mosaic 139, 140, 156, 163, 184, 273, 274, 323
virus 27
Phaseolus lunatus (lima bean) 7
chlorosis 88
mosaic 246
Phaseolus vulgaris (beans)
curly top 79, 234
mosaic 7, 23, 25, 27, 41, 139, 140, 153, 163, 169, 220, 240, 254, 273, 274, 292, 304, 308, 309, 323, 344, 362, 379, 408
virus 27, 78, 79, 235
yellows 352
Phloem necrosis
of beet 267
of coffee 170, 356, 357
of potato 33, 135, 137, 298, 299, 398
Phony peach. See *Prunus*
Photosynthesis 92, 125
Phyllocactus gaertneri, mosaic 286
Physalis
bunchy top 245
mosaic 102, 156, 245, 256, 262, 274, 340, 380, 391
ring spot 382
virus 215
Physio-Chemical studies 27
Physiology 41, 42, 59, 74, 75, 108, 119, 145, 194, 306, 308, 316, 325, 346, 351, 370, 374, 377, 394, 399
Phytolacca decandra, mosaic 11, 126
Phytomonas 356, 357
Phytomoeba sacchari 254
Piesma quadrata 60, 129, 402
Pickel (white) 88
Pine apple. See *Ananas*
Pisum sativum
mosaic 54, 116, 284, 353
streak 230
Pittosporum tobira variegata 77, 78
Plantains. (See also *Musa*)
bunchy top 23, 36, 71, 155, 182, 190, 286, 344
Plasma mosaic 262
Plasmodiophora tabaci 202, 230
Plastids 176, 290

Page

Plum. See *Prunus*
Poeciloscytus cognatus ---- 276
Potash ---- 64
  Effect on potato leaf roll ---- 74
"Pochenkrankheit" of tobacco ---- 110, 191
Poinsettia curl ---- 285
Pokeweed. See *Phytolacca decandra*
Pollen ---- 274, 309
Potato. See *Solanum tuberosum*
Powdery of tobacco ---- 354
Primula virus ---- 154, 237
Properties of tobacco mosaic ---- 10
Protozoa ---- 33, 37, 49, 116, 126, 202, 211, 215 222, 254, 273, 285, 291, 297, 372
Prunella vulgaris, mosaic ---- 231
Prunus (Apricot, cherry, peach and plum)
  Aphids ---- 180, 181
  Apricot ---- 23, 25, 42
  Cherry ---- 25, 175, 308, 311, 313
  chlorosis ---- 88
  little leaf ---- 25, 175
  little peach ---- 36, 52, 53, 74, 91, 236, 247, 292, 344, 345, 390
  mosaic ---- 190, 368
  peach ---- 26, 36, 71, 88, 313, 381
  phony peach ---- 12, 26, 71, 105, 189, 190, 204, 272, 379
  plum ---- 23, 36, 42, 313, 381
  red suture ---- 26
  rosette ---- 52, 139, 145, 175, 246, 276, 292, 345, 390
  spike ---- 186, 345
  virus ---- 42, 82, 381
  yellows ---- 25, 36, 37, 47, 52, 53, 74, 89, 91, 145, 160, 219, 220, 236, 247, 248, 269, 269, 276, 288, 292, 296, 331, 344, 345, 368, 369, 373, 380, 390
Purification of virus ---- 28, 51, 66, 127, 251, 252, 388, 392
Pseudacacia (Black locust)
  brooming ---- 174, 192
Pseudococcus ---- 81, 191, 280
Pseudonet necrosis of potato ---- 301, 302
  (See also acronecrosis, necrosis and netnecrosis.)
Psorosis of citrus ---- 144, 236
Psyllid
  potato ---- 139, 311
  yellows ---- 51, 56, 311, 337
Ptelea ---- 43
Pumpkin. See *Pepo moschata*

Pyrus Page
Pear bitter pit ---- 77, 244
lithiasis ---- 77
new disease ---- 287

Radiation ---- 127
Radicula armoracia (horse radish) ---- 106
Ranunculaceae virus ---- 382
Raphanus raphanistrum virus ---- 51
Raspberry. See *Rubus*
Rate of spread of a virus ---- 162
"Rayas amarillas". See sugar-cane mosaic
Red clover ---- 134, 182, 197
Rejuvenation ---- 108
Resistance ---- 63, 64, 78, 133, 254, 295
Reversion. See *Ribes*
Rhamnus ---- 78, 287
Rheum mosaic (rhubarb) ---- 112
Rhipsalis rosea, mosaic ---- 286
Rhododendron mosaic ---- 285
Rhopalosiphum prunifoliae ---- 118
Rhubarb. (See *Rheum*)
Ribes
big bud ---- 227
chlorosis ---- 89
reversion ---- 12, 226, 227, 239, 312, 313, 316
nettle leaf ---- 227
virus ---- 316
Rice. See *Oryza sativus*
Ring spot
aging of ---- 177
cantaloupe ---- 382
clover (red) ---- 197
Melilotus alba ---- 144
muskmelon ---- 106
Petunia ---- 177, 382, 405
tobacco. See *Nicotiana tobacum*
Río Grande disease of lettuce ---- 77, 218
Robina pseudacacia (black locust) ---- 37, 116, 174, 192, 392
Rosa sp. ---- 397
Rose
chlorosis ---- 105, 260, 261, 273, 274, 397
chlorotic spot ---- 105
dieback ---- 167
mosaic ---- 254, 394
virus ---- 167, 287
Rosette
of apple ---- 175
of apricot ---- 23

Rosette—Continued.. Page
of *Chrysophyllum* 68
of brambles. See *Rubus*
of *Cainito* 68
of *Calendula* 68
of *Calliopsis* 68
of *Chenopodium murale* 175
of *Hordeum* 252
of *Lagerstroemia* 68
of peach. See *prunus*
of peanut. See *Arachis*
of pecan. See *Hicoria*
of raspberry. See *Rubus*
of wheat 196, 250, 392
Rotterdam B disease of tobacco 195
"Rube". See beet.
Rubus 12, 25, 27, 47, 48, 54, 74, 88, 89, 96, 110, 111, 112, 151, 163, 166, 168, 173, 180, 186, 201, 203, 231, 232, 256, 266, 275, 305, 306, 316, 341, 345, 346, 358, 361, 362, 373, 401, 403, 408, 410
Black-berry
dwarf 408
leaf curl 48, 166
rosette 48, 305
run out 305, 410
Raspberry
yellows 88, 186, 361
virus disease 47, 96, 306
Rugose mosaic of potato 129, 198, 210, 381
Rumex
acetosa, curl 402
crispus, leaf deformity 61, 78
lanceolatus, chlorosis & mosaic 163
obtusifolius, chlorosis & mosaic 61, 163
virus 163
Running out. (See also degeneration of potato) 317, 328, 395
Running of potatoes. See *Solanum tuberosum*
Running out of raspberry. See *Rubus*
Rye. See *Secale*

Secale mosaic 251
Sacharum officinarum (sugar cane) 8, 124
chlorosis 131, 143, 264, 274, 363, 364
chlorotic streak 8, 238
Cuban streak 296
dwarf 46
Fiji disease 15, 16, 23, 46, 102, 161, 174, 213, 218, 225, 226, 233, 237, 254, 255, 267, 278, 296, 310, 405

Sacharum officinarum—Continued. Page
leaf hopper ---- 278
mosaic (including "rayas amarillas", mottling, "matizado", yellow stripe, "Gele strepeziekte", etc.)-- 7, 9, 10, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 29, 31, 32, 33, 34, 37, 38, 39, 40, 46, 49, 58, 62, 63, 64. 65, 69, 70, 76, 82, 83, 84, 85, 86, 90, 91, 92, 94, 97, 98, 99, 100, 101, 102, 103, 104, 106, 107, 108, 109, 111, 113, 114, 115, 118, 122, 123, 124, 129, 130, 131, 132, 133, 134, 136, 140, 141, 142, 143, 145, 150, 151, 152, 155, 157, 163, 166, 167, 169, 171, 172, 173, 178, 179, 195, 199, 200, 203, 204, 206, 213, 214, 217, 218, 219, 221, 222, 224, 225, 226, 231, 232, 233, 238, 239, 241, 242, 243, 244, 245, 248, 253, 254, 255, 257, 258, 259, 261, 263, 271, 277, 278, 283, 284, 289, 296, 298, 303, 305, 308, 309, 311, 312, 314, 315, 316, 322, 330, 331, 339, 340, 342, 343, 352, 354, 358, 359, 360, 361, 362, 363, 364, 365, 366, 369, 370, 373, 375, 376, 379, 383, 384, 386, 387, 390, 400, 401, 402, 403, 404, 405, 406
Sereh ---- 32, 46, 47, 98, 193, 199, 215, 216, 217, 221, 222, 225, 233, 263, 303, 345, 353, 362, 377, 379, 390, 395, 400, 401, 404, 409
streak ---- 21, 39, 114, 115, 139, 154, 238, 245, 250, 282, 296, 307, 340, 363, 364, 365, 366, 367
virus ---- 276, 283
white spots ---- 143
Sacharum robustum ---- 64
Salsify yellows ---- 175
Salivary glands of *Cicadula sexnotata* ---- 113
Sandal. See *Santalum album*
Santalum album
leaf-curl mosaic ---- 382
Spike disease ---- 28, 29, 37, 38, 73, 85, 90, 118, 146, 174, 180, 182, 191, 192, 195, 224, 233, 272, 275, 304, 306, 307, 322, 355, 356, 383, 392, 397
Scaly bark of citrus ---- 144
Schizanthus sp. ring spot ---- 280
Scolecosomes ---- 208
Scolytus regulosus ---- 345
Seeds. See transmission
Senecio hieracifolia ---- 230

Sereh. See *Sacharum officinarum* Page
Serology 52, 165, 166, 298
Sesame curl 392
Sesamum radiatum, virus disease 110
Setaria sp., mosaic 102, 245
Sida 94, 219
Sieve tube degeneration 137
Sinamar mosaic 140
Sipha flava 331
Sisal 358
Slugs 297
Snapdragon. See *Antirrhinum*
Soil 193, 201, 223, 250
Solanaceae 134, 158, 165, 215, 299, 324, 382
Solanum
aculeatissimum 57
calcasii 120
capsicastrum 256, 349, 350
carolinensis 156, 382
commersonii 120
dulcamara 215, 256
maglia 120
melongena (Egg plant)
mosaic of 286
psyllid yellows 51
virus 156
nigrum 321, 339
nodiflorum 256, 339, 346, 347
pseudocapsicum 51
tuberosum
altitude 389
apical leaf roll 329
aucuba mosaic 87, 135, 386
beetle (Colorado) 162
calico 294
classification of virus 302
crinkle 87, 248, 249, 270, 317, 349, 386
curl, curl leaf & leaf curl 12, 13, 15, 16, 44, 76, 77, 83, 100, 112, 135, 150, 171, 182, 185, 186, 189, 208, 260, 261, 279, 283, 290, 293, 300, 325, 341, 343, 373, 398, 406, 407
degeneration (running out) 7, 17, 20, 35, 44, 49, 51, 54, 56, 69, 97, 98, 99, 100, 108, 110, 113, 119, 120, 121, 128, 146, 147, 148, 149, 155, 158, 161, 182, 192, 193, 206, 207, 208, 211, 216, 222, 224, 235, 236, 237, 239, 257, 260, 263, 264, 265, 289, 292, 293, 297, 299, 300, 317, 322, 323, 325, 327, 328, 353, 361, 368, 371, 372, 376, 378, 381, 384, 385, 386, 389, 393, 395, 396, 397, 398, 399, 409

Solanum—Continued.
tuberosum—continued.
Page
desication of tuber---------- 38
dwarf & curly dwarf---------- 14, 15, 38, 40, 71, 96, 111, 128, 145, 162, 163, 248, 267, 298, 299, 301, 314, 378
filosité---------- 15, 121, 286, 289
giant hill---------- 158, 159
histology---------- 33, 65, 378, 384, 385
histology---------- 33, 65, 378, 384, 385
internal brown spot---------- 35
interveinal mosaic---------- 87
katahdin mosaic---------- 28
"kringeriheid"---------- 205, 301
latent virus---------- 201, 212, 329
leaf blotch---------- 185
leaf hopper---------- 224
leaf roll---------- 8, 13, 17, 20, 25, 26, 27, 28, 29, 30, 31, 33, 40, 41, 42, 46, 49, 53, 54, 55, 56, 57, 65, 67, 73, 74, 76, 77, 86, 87, 89, 96, 100, 105, 113, 114, 121, 129, 135, 136, 137, 138, 144, 147, 148, 155, 156, 157, 159, 164, 171, 176, 178, 179, 188, 189, 197, 199, 203, 204, 206, 209, 210, 211, 212, 214, 216, 223, 226, 230, 232, 234, 239, 244, 248, 249, 253, 259, 264, 265 268, 269, 270, 271, 272, 273, 277, 279, 281, 282, 283, 284, 289, 290, 298, 299 300, 302, 310, 311, 313, 314, 316, 322, 324, 325, 326, 327, 328, 330, 341, 348, 349, 350, 353, 355, 362, 367, 368, 374, 375, 378, 383, 386, 389, 395, 398, 399, 406, 408
leaf-rolling mosaic---------- 248
marginal leaf roll---------- 144
moron---------- 267
mosaic---------- 14, 15, 16, 17, 20, 25, 26, 28, 31, 40, 44, 54, 55, 56, 60, 72, 73, 77, 87, 96, 97, 100, 106, 111, 120, 129, 133, 135, 136, 137, 144, 147, 151, 167, 180, 189, 194, 199, 210, 211, 215, 216, 226, 230, 234, 235, 236, 244, 248, 256, 261, 268, 269, 270, 271, 281, 282, 298, 299, 300, 311, 313, 314, 317, 318, 323, 325, 327, 328, 329, 330, 344, 346, 347, 348, 350, 353, 355, 362, 375, 376, 381, 387, 389, 392, 397, 398, 410
para crinkle---------- 42, 317
psyllid---------- 139, 311, 337
psyllid yellows---------- 51, 337

Solanum—Continued.
tuberosum—continued. Page
rejuvenation of 15, 36, 322
ring spot 381
rugose mosaic 25, 129, 198, 210, 249, 314, 381, 386
running out 25, 149, 216, 264, 268, 317, 323, 328, 395, 396
spindle-sprout 40, 248, 361
spindle-tuber 22, 25, 59, 144, 148, 149, 158, 159, 162, 163, 166, 182, 202, 234, 238, 244, 249, 320, 328, 375, 384, 385, 395, 396
streak 87, 169, 212, 302, 328
veinbanding 175
mild mosaic 249, 329, 386
mottling 386
necrosis of all types 33, 35, 135, 137, 147, 149, 205, 281, 282, 283, 298, 299, 301, 302, 317, 318, 324, 325, 328, 332, 398
Stipple streak of potato 34, 35, 301, 314, 318
of Eersteling 301
of kopsiaan 301
of noodeling 301
Streak 185, 300, 301, 302, 317, 318, 328, 386
variegated leaves 359
veinbanding 73. 175, 210
virus diseases 12, 20, 24, 25, 26, 27, 28, 33, 41, 42, 44, 52, 55, 56, 65, 66, 72, 77, 96, 99, 105, 119, 121, 122, 131, 132, 135, 146, 148, 149, 155, 157, 161, 162, 166, 169, 171, 175, 191, 192, 197, 207, 208, 209, 211, 212, 215, 222, 223, 230, 233, 234, 235, 244. 247, 248, 249, 255, 261, 263, 264, 268, 269, 270, 271, 274, 276, 280, 282, 283, 284, 286, 287, 289, 290, 299, 300, 301, 309, 313, 316, 317, 320, 324, 325, 326, 329, 339, 344, 347, 348, 349, 350, 351, 353, 354, 361, 374, 380, 389, 393, 394, 395, 397
virus A 232
virus X 211, 318, 349
virus Y 211, 318, 349
witches' broom 189, 192, 253, 407
yellows 51, 56, 311, 337
yellow spot 31
Soja Max (soy bean) 42, 89, 155, 205
Sonchus oleraceus 230
Sorbus chlorosis 43
Soy Bean. See *Soja Max*
Spectrum of chlorophyll of cane 90

Spike disease — Page
of *Dodonaceae viscosa*. See D. viscosa
of *Lantana camara*. See L. *camara*
of peach — 186
of sandal wood. See *Santalum album*
Spinacia oleracea (Spinach) — 182
blight — 71, 196, 246, 351, 377, 378
deformity — 60
mosaic — 12, 26, 59, 61, 62, 181, 247, 323, 351, 389, 400
virus — 181, 182
Spinat. See *Spinacia*
Spindle sprout of potato — 361
See also *Solanum tuberosum*
Spindle tuber of potato. See *Solanum tuberosum*
Spots of tobacco — 124, 191
Spots (white) of cane — 143
Spots (yellow) of spinach — 62
Spotted wilt of tomato — 8, 38, 76, 171, 184, 275, 280, 293, 321, 342, 350
Sprain of potato — 35, 185
Spread (rate) — 247
Squash. See *Pepo moschata*
Standardization technique — 127
Starch — 37, 57, 91, 137, 179, 223, 233, 322, 374, 398
Stellaria media curly top — 223
Sterilization of soil — 11
Sterility — 214, 215, 218, 261
Sternbergia — 170
Stigmonose of apple — 68
Stomatal infection — 127, 196
Strawberry. See *Fragaria*
Streak of cane. See *Sacharum Officinarum*
of corn. See *Zea mays*
of (Cuban cane) — 296
of *Digitaria* — 365, 366
of *Eleusine indica* — 365, 366
of *Emilia* — 230
of *Pisum sativum* — 230
of Raspberry. See *Rubus*
of tobacco. See *Nicotiana tabacum*
of tomato. See *Lycopersicum esculentus*
in greenhouse — 200
Stream double refraction — 371
Streptosolen jamesonii — 280
String leaf of tobacco — 88
Stripe
of barley — 371
of cane. See *Saccharum officinarum*

Stripe—Continued. Page
of corn ... 67, 358
of daffodils ... 107
of grass ... 226
of potato ... 300
of rice ... 220
of tomato. See *Lycopersicum esculentus*
Strongyloplasma Iwanowskii ... 285
Stunt
A. of *Delphinium* ... 73
of iris ... 67
of rice ... 372
Sualda moquini ... 78
Sugar cane. See *Saccharum officinarum*
Sunblotch. See *Persea*
Sweet clover. See *Melilotus*
Sweet pea ... 372, 397
Sweet potato. See *Ipomoea*
Syntherisma ... 288
Systena elongata ... 162

Tarnish bug ... 162
Tarsonemus fragaria ... 239
Tea ... 73
Temperature ... 55, 56, 58, 112, 139, 177, 197, 201, 241, 247, 257, 266, 280, 295, 308, 332, 334, 362, 366, 376, 401
Tetranychus telarius ... 87, 293, 371
Thrips minuta ... 97
Thrips tabaci ... 81, 87, 100, 184, 194, 229, 230, 293, 321, 349, 350
Tithonia diversifolia, virus ... 51
Tobacco. See *Nicotiana tobacum*
Toleration ... 133
Tomato. See also *Lycopersicon*
big bud ... 321
bunchy top ... 245
See also bunchy-top
Tomotosis of cotton ... 95, 304
Top-Necrosis of potato ... 307
See also *Solanum tuberosum*
Trachelium sp. ring spot ... 280
Transmission ... 134, 144, 148, 153, 172, 215, 239, 255, 306 380, 396, 406, 408
Artificial transmission ... 63, 249
by grafting ... 48, 90, 174, 308, 313, 320, 374
by inoculation ... 349

Transmission—Continued.
Artificial transmission—Continued. Page
by insects ---------- 7, 8, 10, 12, 16, 18, 19, 28, 38, 39, 41, 47, 48, 51, 55, 58, 59, 60, 61, 62, 63, 64, 65, 67, 68, 79, 80, 81, 82, 83, 84, 85, 86, 87, 90, 91, 97, 100, 101, 102, 108, 113, 117, 118, 119, 126, 129, 134, 135, 136, 140, 142, 143, 152, 157, 159, 160, 162, 163, 169, 170, 171, 172, 173, 175, 181, 184, 185, 190, 193, 194, 206, 212, 217, 218, 220, 221, 227, 229, 230, 235, 238, 239, 240, 245, 246, 248, 249, 261, 262, 267, 269, 270, 273, 274, 276, 277, 278, 279, 280, 284, 288, 293, 294, 296, 297, 304, 305, 306, 311, 314, 320, 321, 323, 324, 327, 330, 332, 333, 334, 335, 336, 337, 346, 348, 349, 350, 351, 352, 357, 358, 362, 363, 364, 365, 366, 367, 368, 369, 370, 371, 372, 373, 374, 381, 384, 389, 391, 394, 398, 399, 400, 402, 403, 406, 408, 409
by insect eggs ---------- 154
by knife ---------- 59, 162
by seeds ---------- 49, 115, 127, 135, 136, 177, 205, 219, 220, 246, 273, 274, 304, 308, 382
by slugs ---------- 297
by soil ---------- 117
by weeds ---------- 403
of aster yellows ---------- 220
of avocado sun blotch ---------- 286
of bean mosaic ---------- 59, 292
of cranberry false blossom ---------- 48
of cucumber mosaic ---------- 115, 193
of curly top of beet ---------- 80, 143, 152
of Fiji ---------- 161, 267
of gray disease of narcissus ---------- 255
of hop virus ---------- 54, 374
of lettuce mosaic ---------- 49, 68, 193
of little peach ---------- 91
of peach yellows ---------- 91
of potato leaf roll ---------- 87, 129, 349
of potato streak ---------- 328
of potato to tobacco ---------- 197
of potato witches' broom ---------- 407
of potato virus disease ---------- 215, 244, 280, 394, 398, 399, 406
of raspberry mosaic ---------- 401
of reversion of black currants ---------- 239
of rose ---------- 387
of *Solanaceae* ---------- 324
of soy bean ---------- 205

Transmission—Continued.
Artificial transmission—Continued. Page
of spike of sandal 90
of spindle tuber 328
of spinach 59
of sugar cane 136
of sugar-cane mosaic 331
of sweet-potato mosaic 315
of tobacco mosaic 265, 280
of tobacco ring spot 177
of tomato 280
of wheat mosaic 252
of yellow stripe of cane 221
Trifolium sp. 116, 156
leafhopper of 182
mosaic of red clover 134
mosaic of sweet clover 134, 252
ringspot of red clover 197
Triticum (wheat)
mosaic 123, 250, 251, 256, 392, 393
rosette 196, 250, 251, 392
Tulips 397
breaking 25, 28, 82, 170, 184, 202, 224, 248, 249, 255, 272
falling 293
mosaic 125, 248, 249
Turnip mosaic. See *Brassica*
Trypanoplasts 207, 214
Trypanosomas 211

Unmottled dwarf. (See *Solanum tuberosum*)
Ultra-microscopic organism, virus, etc. 25, 44, 157, 175, 237, 250, 327
Ultra-violet light 183
Ultra virus 44, 175

Vaccinium, false blossom (cranberry) 44, 45, 48, 113, 150, 151, 201, 287, 323, 338, 354, 359, 400, 401
leaf hopper 44, 113
Variegations 42, 43, 59, 154, 175, 221, 241, 263
of *Abutilon* 94, 205, 309
of apple 62
of citrus 292
of leaves 154
of *Morus* 94
of *Pittosporium tobira* 77, 78
of potato 353
of *Sida* 94
Veinbanding of potato 73, 175, 219
Vein streak of tobacco 196
Verbesina altemifolia 144

Page
Vicia faba ---- 59, 134, 256, 274
Vigna sesquipedalis, mosaic ---- 274
Vigna unguiculata mosaic ---- 156, 344
Vinca sp. rosette ---- 68
Vine
leaf roll ---- 291
mosaic of ---- 285, 368, 369
Viola tricolor, virus ---- 237
Virosis
of plum peach & cherry ---- 313
Virus, longevity ---- 292
nature of ---- 50
Virus A of potatoes ---- 232
Virus diseases (general) ---- 41, 44, 50, 55, 60, 65, 66, 67, 68, 71, 72, 73, 87, 95, 112, 123, 124, 125, 126, 127, 147, 154, 156, 160, 164, 166, 180, 195, 197, 198, 199, 201, 208, 211, 214. 215, 219, 221, 225, 228, 230, 250, 251, 257, 260, 263, 264, 265, 279, 300, 302, 303, 307, 309, 318, 320, 321, 322, 323, 325, 339, 344, 346, 347, 350, 351, 353, 354, 362, 363, 365
of *Ambrosia trifida* ---- 237
of *Antirrhinum* ---- 237
of beet ---- 386
of *Bougainvillea spectabilis* ---- 237
of *Calendula officinalis* ---- 51
of *Campanulales* ---- 382
of cassava ---- 110, 264
of *Crocus* ---- 170
of cucumber ---- 212
of *Cucurbitaceae* ---- 382
of currants ---- 316
of daffodil ---- 167
of *Delphinium* ---- 176, 189, 237, 382, 397
of *Fragaria* ---- 173
of *Fresia* ---- 170
of *Fuchsia* ---- 237
of gladiolus ---- 170
of greenhouse ---- 212
of *Hippeastrum* ---- 235
of *Hyoscyamus* ---- 171
of lily ---- 110, 361
of *Locheria rosea* ---- 110
of Narcissus ---- 255
of *Passiflora* sp. ---- 51
of peanut ---- 364
of pepper ---- 156
of *Physalis* ---- 215

Virus diseases (general)—Continued. Page
of peach 82, 381, 384, 388, 391, 398
of raspberry 47, 96, 306
of *Ranunculaceae* 382
of *Raphanus raphanistrum* 51
of *Sesamum radiatum* 110
of tobacco 27, 72, 181, 183, 197, 297, 367
of tomato 72, 212, 220
property of 199
Vitis, from grape and vine 291, 368, 369
mosaic 285, 368, 369
"Vouille blanche" of tobacco 110

Walnut 155, 169
Warm water treatment 48, 63, 142
Watermelon mosaic 106, 391
Weeds
leaf roll of 129
rugose mosaic 129, 381
virus 380, 403
Wheat. See *Triticum*
White fly. See *Aleurodidae*
White heart of lettuce 218
White pickle of cucumber 88, 96, 193, 256
White tobacco 122, 124
Wild hosts of cane mosaic 63
Wilt
of aster 202, 234
of rose 167
of pineapple 81, 190
of tomato (spotted). See *Lycopersicum esculentus*
Wiry tomato 228
Witches' broom
of *Holodiscus discolor* 409
of locust 192, 392
of ocean spray 409
of potato 189, 192, 253, 407
of strawberry 408
of tomato 407
Woodines of passion flower 274, 275

X-virus of potato 57, 211
Xanthosis of strawberry 293

Y-virus 57
Yellow dwarf
of onions 118, 177, 257
of potato 40, 145, 267
Yellow disease of beans 352
of fall dandelion 264

Page
Yellow edge of strawberry ---- 173, 239
Yellow flat of lilies ---- 176
Yellow mosaic
of tobacco ---- 126, 195
of tomato ---- 50, 57
of wheat ---- 252
Yellow pickle ---- 375
Yellow stripe of daffodil ---- 107, 167
of dandelion (fall) ---- 264
Yellow disease of *Leontodon autumale* (fall dandelion) ---- 264
Yellow spot
of pine apple ---- 81, 190, 229
of spinach ---- 62
Yellow stripe. See *Saccharum officinarum* (mosaic)
of grass ---- 226
transmission ---- 217, 221
(See also transmission.)
Yellow tomato blight (Western) ---- 248
Yellow top
of alfalfa ---- 165
Yellows
of alfalfa ---- 200
of asters ---- 113, 153, 175, 202, 218, 219, 220, 234, 279
of beets ---- 377
of carnation ---- 287, 288
of carrot ---- 220, 336
of celery ---- 175, 220, 335
of *Chrysanthemum* ---- 273
of egg plant (psyllid) ---- 51
of *Erigeron* ---- 246
of lettuce ---- 335
of parsley ---- 336
of peach ---- 36, 37, 47, 53, 74, 91, 145, 160, 219, 220, 236, 247, 248, 263, 288, 292, 344, 331, 345, 368, 369, 373, 390
of psyllid ---- 51, 337
of raspberry ---- 186, 256
of salsify ---- 185
of strawberry ---- 293, 294
of tomato ---- 219, 311, 337, 338
of walnut ---- 169
of Y-virus ---- 57

Zea mays (corn) ---- 235
chlorosis ---- 109, 340
dwarfing ---- 397
mosaic ---- 63, 217, 219, 315, 357, 363, 364, 391
streak ---- 173, 213, 315, 365, 366, 367

Zea mays—Continued. Page
stripe ---- 67, 358
yellowing ---- 397

Zinnia elegans
kroepoek ---- 205
Zinnia leaf curl ---- 205, 240
Zosmenus cuadratus ---- 402
"Zukerrube". See beet
Zyzyphus ---- 180

AUTHORS' INDEX

	Page
Abbeg, F. A.	79
Abbott, E. V.	7
Adam, D. B.	7
Adams, J. F.	7, 236
Adams, R. L.	7
Afzal, H. M.	8, 189
Agee, H. P.	8
Ahr, J.	8
Ainsworth, G. C.	8
Alamo Ibarra, R.	9
Aben, A. O.	9
Alexander, J.	9
Alfaro, J.	9
Allard, H. A.	10, 158
Allen, F. W.	11
Alston, R. A.	12
Amaral, A. de	12
Amos, J.	12
Anders, C. B.	393
Anderson	12
Anderson, E. J.	201
Anderson, F. G.	12
Anderson, H. W.	12, 89
Anderson, J.	12
Anderson, P. J.	89
Andrews, F. W.	239
Angood, E.	163
Annaud, P. M.	13
Anonymous	13
Anstead, R. D.	29
Appel, O.	29
Appleman, C. O.	31
Arango, R.	31
Arceneaux, G.	32
Archer, W. A.	175
Arendsen, H. S. A.	32
Armstrong, J. K.	126
Arthur, J. M.	32
Arthur, J. C.	32
Artschwager, E. F.	33
Ashby, S. F.	33, 199
Asunción, S.	34
Atanosoff, D.	34
Atwood, G. G.	36
Auchinleck, C. G.	36
Aumiot, J.	36
Averna-Saccá, R.	37
Avery, B. T.	53
Badami, B. S. R.	37
Bailey, E.	201
Bailey, I. W.	37
Bailey, L. H.	37
Baissac, L.	38
Bakke, A. L.	38
Bald, J. G.	38, 320
Ball, E. D.	38, 304
Bancroft, C. K.	199
Barber, C. A.	38
Baribeau, B.	40
Barker, H. D.	40, 272, 352
Barner, J.	65
Barreto, B. T.	40
Barrus, M. F.	40
Barss, H. P.	40
Barton-Wright, E.	41
Basinger, A. J.	333
Batchelor, L. D.	169
Baudys, E.	42
Baur, E.	42
Bawden, F. C.	43, 317
Bayon, H. P.	44
Beale, H. P.	44, 297
Beaumont, A.	44
Beauverie, J.	44
Beauverie, M. A.	44
Bechhold, H.	44
Beckwith, C. S.	44
Bedson, S. P.	45
Beecher, F. S.	337
Behrenz, J.	45
Beijerinck, M. W.	45
Beke, L. von	46
Bell, A. F.	46, 267
Benecke, F.	46
Bennett, C. W.	47, 232
Berg, A. J. J.	48
Bergman, H. F.	48

	Page
Berkeley, G. H.	48
Bersch, W.	49
Bensaude, M.	**49**
Betancourt, P. E.	49
Bewley, W. F.	49, 57, 285
Bielert,	325
Bijl, P. A. van der	50
Bijlert, A. van	51
Binkley, A. M.	51
Biourge, P.	51
Birkeland, J. M.	51
Birmingham, W. A.	52
Bisby, G. R.	52, 216
Bisland, R. B.	32
Black, L. M.	145
Black, O. F.	377
Blackman, V. H.	52, 69
Blake, M. A.	52
Blakeslee, A. F.	53
Blanchard, E.	53
Bland, J. O. W.	45
Blaringhem, L. F.	54
Blattny, C.	54, 369
Blodgett, F. M.	55
Blood, H. L.	56, 117, 311, 249, 392
Boas, F.	56
Bodnár, J.	114
Bohm, F.	57
Bohme, R. W.	57
Bohutinsky, G.	57
Bokma de Boer, B.	209
Bolas, B. D.	50, 57
Bolley, H. L.	32
Bonazzi, A.	58
Boncquet, P. A.	58, 352
Bonde, R.	59, 149, 329
Böning, K.	59
Booberg, K. G.	61
Borg, P.	61
Borisevich, G. F.	61
Bottomley, A. M.	365, 366
Bouriquet, A. G.	61
Bouygues, H.	62
Bovell, J. R.	62
Boycott, A. G.	62
Bradford, F. C.	62
Brandenbourg, E.	62
Brandes, E. W.	62, 373
Breda de Haan, J. von	64
Breemen, P. J. von	64
Brehmer, G. von	65
Brewer, P. H.	66, 216
Brick	67
Brierley, P.	67, 248
Brierley, W. B.	67
Brittlebank, C. C.	67
Britton-Jones, H. R.	67
Brock, J. A.	68
Brooks, A. J.	68
Brooks, C.	68
Brooks, F. T.	68
Brown, B. A.	69
Brown, N. E.	69
Brown, W.	69
Broz, O.	209
Bruner, S. C.	69, 104, 150
Brunnich, J. C.	71
Brunton, L. A.	386
Bryant, M. W.	71
Bryce, G.	71
Buchwald, N. F.	71
Bunzel, H. H.	71, 377
Bunzli, H.	356
Burger, O. F.	72
Burkholder, W. H.	72
Burnett, G.	72, 176, 201
Busch, H. J.	73
Busch, W.	73
Butler, E. J.	73
Butler, O.	73
Butler, O. W.	198
Button, H. F.	74
Caesar, L.	74, 304
Caldwell, J.	74
Calinissa, M. R.	75
Caluwe, P. de	76
Calvino, M.	76
Campbell, D. C.	76
Campbell, E. G.	76
Camuñas, M.	76
Capinpin, J. M.	379
Carne, W. M.	76, 400
Carpenter, C. W.	77
Carrante, V.	77
Carroll, E.	77
Carriére, E. A.	77
Carsner, E.	78, 357

	Page
Carter, W.	79
Casagrandi, O.	81
Cation, D.	82
Cayley, D. M.	82
Ceresa, G.	82
Chamberlain, J. C.	13
Chapman, G. H.	**82, 363**
Chapple, J.	83
Chardon, C. E.	83
Chatterju, N. C.	85
Cheal, W. F.	85
Chevalier, A.	85, 132
Chill, T.	85
Christensen, R. J.	372
Christie, G. I.	85
Chu, H.	85
Chupp, C. D.	40
Church, M. B.	85
Ciferri, R.	85
Clark, C. F.	329
Cláusen	86
Clayton, E. E.	86
Cleare, Jr. L. D.	87
Cleveland, C. R.	87
Cleveland, L. R.	87
Cleveringa, O. J.	87
Clinch, P.	87
Clinton, G. P.	88
Cockerham, G.	89, 163
Coit, J. E.	89
Colby, A. S.	89
Cole, J. R.	9
Coleman, L. C.	90
Collard, H.	160
Colón, E. D.	90
Condit, I.	90
Connors, C. H.	52, 53
Cook, M. T.	52, 91
Cook, O. F.	95
Cooley, L. M.	96
Coons, G. H.	96, 215
Cooper, T.	96
Corbett, W.	50
Cordingley, H.	96
Cormany, C. E.	79
Corneli, E.	97
Costa Lima, A. da	97
Costantin, J.	97
Cottier, W.	100
Cotton, A. D.	100
Cottrell-Dormer, W.	101
Cottrell, R. H.	102
Cowdry, E. V.	102
Cowgill, H. B.	102
Crawford, R. F.	102, 139
Crawley, J. T.	102
Craybill, C. H.	309
Crispeyn, C. P.	36
Cross, W. E.	103
Cruz, F. B.	104
Cunningham, G. C.	104
Currie, F.	398, 399
Curtiss, C. F.	105
Curzi, M.	105
Cuthbertson, D. C.	105
Cutler, G. H.	105
Dade, H. A.	105
Dafert, F. W.	105
Dale, H. H.	105
Damf, A.	106
Dana, B. F.	106, 189
Daniel, L.	106
Darlington, H. R.	107
Darnell-Smith, G. P.	107
Dastur, J. F.	107
Davidson, W. D.	108
Davis, E. F.	108
Davis, E. W.	108
Davis, J. J.	108
Davis, R. L.	108
Davis, W. H.	109
Davy, E. D.	358
Decoux, L.	109
Deerr, N.	109
Deighton, F. C.	109
Delacroix, E. G.	110
De Long, D. M.	110
Delplace, E.	110
Demarre, J. B.	110, 343
Detmers, F.	110
Dey, P. K.	111
Dickson, B. T.	111
Dickson, J. G.	201
Dickson, T.	112
Dijk, J. van	113
Dijk, M. D.	113, 299
Dix, W.	113

	Page
Dobroseky, I. D.	113
Dobrosky, I. B.	113
Dobrosky, T. L.	113
Doby, G. K. von	114
Dodds, H. H.	114
Dodge, B. O.	115
Doolittle, S. P.	115
Dorst, J. C.	117, 299
Dosdall, L.	118
Doty, R. E.	118
Dover, C.	118
Down, E. E.	274
Downson, W. J.	280
Doyer, L. C.	118
Drake, C. J.	118
Dschounkowsky, E.	119
Duboys, C.	119
Duchartre, P.	119
Ducomet, V.	119
Duffield, C. W.	122
Dufrénoy, J.	122
Duggar, B. M.	126, 196
Dungan, G. H.	251, 392
Dunlap, A. A.	128
Duriez, C.	128
Durrel, L. W.	128
Dvorak, M.	128
Dyckerhoff, F.	129
Dykstra, T. P.	129, 248, 249
Eardley, C. M.	321
Earle, F. S.	129, 373
East, E. M.	131
Eastham, J. W.	131
Eastwood, H. W.	132
Eberhardt	132
Eckerson, S. H.	132, 216, 250
Edgerton, C. W.	132, 375
Egiz, S. A.	134
Elcock, H. A.	79
Elliot, H. G.	77
Elliot, J. A.	134
Elmer, O. H.	134
Elze, D. L.	135, 300, 301, 302
Emmerez de Charmoy, D. 'd	136, 213
Ensign, M. R.	136
Eristavi, E. M.	136
Esau, K.	137, 234
Esmarch, F.	137
Euler, H. von	138
Eustace, H. J.	361
Evans, I. B. P.	139
Evans, P.	139
Eyer, J. R.	139
Eyles, F.	139
Faes, H.	139
Fajardo, T. G.	139
Fallada, O.	140
Faris, J. A.	140
Farvuharson, C. O.	141
Fawcett, G. L.	103, 141
Fawcett, H. S.	144
Fenne, S. B.	144
Ferdinandsen, C.	144
Ferguson, J. H.	144
Fernow, K. H.	55, 56, 144
Ferraris, T.	145
Fife, J. M.	145
Figueroa, C. A.	145
Filho, A. F. O.	145
Finch, A. H.	145
Findley, W. M.	146
Fisher, C. E. C.	146
Fisher, D. F.	68
Fletcher, T. B.	146
Flexner, S.	146
Flint, W. P.	146
Foëx, E.	120, 146, 299
Folsom, D.	147, 327, 328, 329
Forsbeck, F. G.	281
Forsberg, A.	138
Forsteneichner, F.	149
Fortún Martínez, G.	150
Foster, A. C.	150
Fowlie, P.	115
Fracanzani, G. A.	150
Fracker, S. B.	150
Frank, A.	151
Frank, A. B.	150
Franklin, H. J.	151
Freeman, E. M.	151
Freeman, W. G.	151
Freiberg, G. W.	152
Freise, F. W.	152
Freitag, J. H.	152, 336
Froberville, L. F. de	152
Fromme, F. D.	152, 402

	Page
Fukano, H.	153
Fukushi, T.	153
Fuller, C.	154
Fulmek, L.	154
Funaoka, S.	154
Gadd, C. H.	155
Gaget, J.	155
Gandía Córdova, R.	155
Garbowski, L.	155
Garcke	155
Gard, M.	155
Gardner, M. W.	66, 155, 205, 216, 304
Gates, R. R.	157
Gaylor, F. C.	157
Geerts, J. M.	157
Gentner, G.	179
Gertz, O.	157
Ghésquiere, J.	157, 217
Ghimpu, V.	157
Giddings, N. J.	158
Gigante, R.	158
Gilbert, A. H.	158
Gilbert, W. W.	115, 159
Gildeham, E. J.	388
Gloyer, W. O.	159
Goddard, E. J.	160
Gold, T. S.	160
Golding, F. D.	160
Goldstein, B.	160
Goldsworthy, M. C.	161
Gontiére, J. F.	161
Goot, P. van der	161
Gopalaswami, N. G.	355
Goseco, A. P.	161
Goss, R. W.	161
Gounaux, C. B.	163
Gowdey, C. C.	163
Grainger, J.	96, 163
Gram, E.	164
Granovsky, A. A.	165, 200
Grant, T. J.	165, 199
Gratia, A.	165
Gratz, L. O.	166
Gravier, G.	166
Green, S. B.	166
Gregory, C. T.	157, 167
Grey, R. M.	167
Griffiths, D.	167
Grieve, B. J.	167
Groene, F. de	167
Grove, A.	167
Grubb, N. H.	168, 173
Guba, E. F.	168
Guezé, P.	136
Gulyás, A.	168
Güssow, H. T.	168
Gutermann, C. E. F.	169, 280
Haar, A. W. van der	299
Haas, A. R. C.	169
Haasis, F. W.	174
Hach, O.	169
Haddon, F. C.	169
Haegele, R. W.	170
Hall, A. D.	170
Hall, C. J. J. van	170
Halle, T. G.	171
Hallier, E.	171, 279
Hallowell, E. A.	182
Hamann	171
Hamblac, H.	171
Hamblin, C. O.	171
Hamilton, M. A.	171
Hanken, H. A.	172
Hansen, A. A.	167
Hansford, C. G.	172
Hanson, A. P.	173
Haring, C. M.	173
Harreveld, Ph. van	173
Harrington, F. M.	173
Harris, H. M.	118
Harris, R. V.	173
Hart, W. C.	174
Harter, L. L.	174
Hartley, C. P.	174, 192
Hartung, W. J.	58, 330, 333
Hartwell, B. F.	396
Harvey, R. B.	174
Haskell, R. J.	174
Handuroy, P.	175
Hatton, R. G.	12
Hause, H.	175
Hawkins, L. A.	327, 377, 378
Hawley, I. M.	72, 175
Hayes, T. R.	175
Heafford, R. M.	164

	Page
Heald, F. D.	175
Hédin, L.	176
Hedlund, T.	176
Heileman, W.	223
Heim, R.	176
Heim, I.	176
Heintzel, K. E.	176
Hell, W. F. van	176
Henderson, C. F.	13, 334
Henderson, L. F.	177
Henderson, R. G.	177
Henderson, W. J.	177, 257
Henning, E. J.	178
Herbert, D. A.	178
Heribert-Nilsson, N.	178
Hernández, A.	178
Hernández, C. C.	278
Hernández Torres, C.	178
Hertzsch, W.	138, 178
Heuberger, J. W.	178, 276
Hildebrandt, F. M.	327
Hiltner, L.	179
Himmelbaur, W.	179
Hind, R. R.	179
Hinson, W. M.	179
Hite, B. H.	158
Hockey, J. F.	180, 305
Hodgson, C. M.	180
Hodgson, W. E. H.	44, 180
Hoffman, D. C.	281
Hoffman, M.	180
Hoggan, I. A.	180, 199
Holden, J. A.	182
Hole, R. S.	182
Hollaender, A.	127
Holland, T. H.	182
Hollins, E.	182
Hollrung, M.	182
Holmes, F. O.	183
Holmes Smith, E.	184
Hoog, J.	184
Hopkins, J. C. F.	185
Hori, S.	185
Horne, A. S.	185
Horne, W. T.	90, 185, 286, 308
Horsfall, J. L.	186
Howard, A.	186
Howard, R. F.	395
Howdeniensis	186
Howitt, J. E.	186, 363
Hunger, F. W. T.	187
Hungerford, C. W.	188, 292
Huntley, F. A.	189
Hurley, F.	256
Hurtado, E. A.	278
Husain, M. A.	189
Hutchins, L. M.	189
Hutson, J. C.	190
Hutton, S. B.	44
Iachevskii, A.	190
Ichitkawa, N.	190
Ikeno, J.	190
Illingworth, J. F.	190
Ito, S.	191
Iwanowski, D.	191
Iyengar, A. V. V.	191
Iyengar, B. N.	192
Iyengar, C. S. R.	37
Iyengar, J. V. V.	322
Jackson, A. B.	48, 192
Jackson, H. S.	192
Jackson, L. W. R.	192
Jaczwski, A. L.	192
Jaggar, I. C.	193
Jaguenaud, G.	193
Jankins, E. H.	179
Janse, J. M.	193
Janssen, J. J.	193
Jarrett, P. H.	194
Jehle, R. A.	194
Jensen, H.	194
Jensen, J. H.	194
Jiménez Núñez, E.	195
Jivanna Rao, P. S.	195
Jochems, S. C. J.	195
Jodidi, S. L.	196, 377
Joest, E.	196
Joglar Rodríguez, F.	196
Johnson, A. G.	196
Johnson, B. P.	127, 196
Johnson, E. M.	197, 380, 381, 382
Johnson, G. W.	197
Johnson, J.	180, 197, 201, 307
Jöhnssen, A.	324
Johnston, J. R.	199
Joley, L.	62

	Page
Joly, R. L.	200
Jones, F. R.	116, 200
Jones, G. H.	200
Jones, H. A.	338
Jones, L. K.	72, 200
Jones, L. R.	201
Jones, P. M.	202, 230
Jones, W.	202
Jordi, E.	202
Jorstad, I.	203
Jorgensen, C. A.	203
Kaiser, P.	203
Kamerling, Z.	203
Karatschewsky, I. K.	313
Karrer, J. L.	126
Karunakara Menon, C.	203
Kasai, M.	204
Keller, W.	79
Kellerman, K. F.	204
Kellog, E. H.	196, 377
Kelly, J. W.	377
Kelly, N. L.	204
Kendrick, J. P.	66, 155, 157, 205, 392
Kerling, L. C. P.	205
Keur, J. Y.	205
King, C. J.	205
Kinney, A.	206
Kinnison, A. F.	145
Kirk, T. W.	206
Kirkpatrick, T. W.	206
Kirschner, R.	206
Klaphaak, P. J.	63
Klapp, E. L.	206
Klebahn, H.	207
Klemm, M. J.	208
Klinkowski, M.	259
Knight, R. C.	12
Knight, T. A.	208
Knorr, P.	208
Knowlton, G. F.	208
Kobus, J. D.	209
Koch, K. L.	210
Kock, G.	210
Koeslag, F. D.	211
Kofoid, C. A.	211
Kóhler, E.	211
Köning, C. J.	212
Koningsberger, V. J.	213
Kopke, E. W.	225
Kopp, A.	213
Kornauth, K.	209, 210, 214
Kostoff, D.	214
Kotila, J. E.	96, 215
Kottman, G.	215
Kramer. S. P.	215
Krantz, F. A.	216
Kranzlin, G.	216
Kraybill, H. H.	66, 132, 216, 354, 373
Kreitz, W.	29, 30
Krieckner, E. O. O. von	206
Krüger, K.	216
Krüger, W.	216
Krumbhaar, C. C.	32
Kufferath, H.	217
Kuijper, J.	221
Kulkarni, G. S.	217
Kunkel, L. O.	217
Kuribayashi, K.	220
Kuster, E.	221
Kuyper, J.	221
Kutin, A.	222
Labergerie	222
Labo, A.	222
Lacey, M. S.	222
Lackey, C. F.	79, 222
Lafrenz, E.	223
Lamkey, E. M. R.	223
Lang, W.	223
Laphem, M. H.	223
Larose, E.	223
Larrimer, W. H.	250
Larsen, L. D.	223
Laske, C.	223
Latham, H. A.	224
Laubert, R.	224
Lawrence, W. H.	224
Leach, J. G.	151, 224
Leak, G. D.	224
Lebard, P.	98, 224
Ledeboer, F.	224, 400
Ledingham, J. C. G.	225
Lee, H. A.	225, 274
Leefmans, S.	226
Lees, A. H.	226
Lehman, S. G.	228, 403
Leighty, C. E.	196, 392

	Page
Lek, H. A. A. van der	298
Leonion, M. S.	274
Le Pelley, R. H.	228, 317
Leroux, L.	228
Lesley, J. W.	228, 338
Lesley, M. C. M.	228
Levshin, A. M.	228
Lewin, C. J.	228
Lewis, R. D.	9
Lewton, B. L.	228
Likhité, V. N.	229, 302
Lind, J.	229
Lindemuth, H.	229
Lindstrom, E. W.	72
Linford, M. B.	229
Linhart, G.	230
Link, A. M. de S.	230
Link, G. K. K.	230
Lipschutz. B.	230
Liro, J. I.	231
Ledewijks Jr., J. A.	231
Locsin, C.	225
Loew, O.	231
Lojkin, M.	231
Loomis, H. F.	205
López Domínguez, F. A.	231
Loree, R. F.	232
Loughnane, J. B.	232
Lowe, C. C.	79
Ludewig, K.	232
Ludtke, M.	232, 324
Lushington, P. M.	233
Lyman, G. R.	233
Lyon, H. L.	233
Mac Callum, W. G.	234
Mac Clement, D.	234
Mackenzie, D.	234
Mackie, W. W.	234
Mac Lennan, A. H.	234
Mac Leod, D. J.	234
Macley, K. S.	196
Mac Millan, H. G.	235
Macoun, W. T.	235
Madden, G. O.	49
Magee, C. J. P.	160, 235
Magrou, J.	235
Maige, L. A.	235
Malcom, P.	190
Malhotra, R. C.	235
Malpeaux	236
Mandelson, L. F.	236
Mandenburg, E. C.	236
Manns, T. F.	236
Marañón, G.	140
Marchal, E.	236
Marins Ramos, C. S.	238
Márquez, S. L.	237
Marre, E.	237
Marshall, W.	237
Martin, C.	237
Martin Jr., G. H.	174, 237
Martins, J. P.	238
Martin, W. P.	238
Martyn, E. B.	239
Martyn, T.	261
Marudarajan, D.	239
Marx, T.	239
Mason, T. G.	200
Massee, A. M.	12, 168, 239
Massée, G.	239
Massey, R. E.	239
Mathur, R. N.	240
Matsumoto, T.	240
Mattei, G. E.	241
Matthews, W. H.	241
Matz, J.	241
Maublanc, A.	243
Maupas, A.	243
May, D. W.	243
May, W.	243
Mayer, A. E.	243
Mayr, C.	8
Mc Alpine, D.	244
Mc Bain, A.	41
Mc Call, T. M.	244
Mc Callan, E. A.	244
Mc Clean, A. P. D.	245, 366
Mc Clintock, J. A.	246
Mc Cormick, F. A.	89
Mc Cubbin, W. A.	246
Mc. Curry, J. B.	305
Mc Donald, J.	248
Mc Kay, M. B.	248
Mc Kay, R.	269, 270
Mc Kenny Hughes, A. W.	249
Mc Kinley, E. B.	250
Mc Kinney, H. H.	116, 196, 250

	Page
Mc Kinney Jr., W. H.	283
Mc Larty, H. R.	252
Mc Lean, W.	253
Mc Murran, S. M.	253
Mc Murtrey, Jr., J. E.	253
Mc Rae, W.	253
Mc Rostie, G. P.	111, 254
Mc Whorter, F. P.	67, 254, 394
Medalla, M. G.	225, 226, 255
Meer, J. H. H. van der	255
Megau, W. J.	256
Melchers, L. E.	256
Melhus, I. E.	256
Menecacci, M.	257
Mendiola, N. B.	257
Menéndez Ramos, R.	257
Menezes Sobrino, A.	259
Merkel, L.	259
Merkenschlager, F.	239, 259
Merril, E. D.	260
Meulen, J. G. J. van der	260
Meyer, F. H.	260
Meyer, H.	260
Mezey, G.	230
Mil, P. J.	134
Milbraith, D. G.	260
Miles, H. W.	261
Miles, L. E.	261
Milhoffer, S.	261
Miller, J.	261
Miller, M.	201
Miller, P.	261
Milward, J. G.	261
Mitra, M.	261
Mogendorff, N.	261
Molisch, H.	262
Molz, E.	262
Monteith Jr., J.	182
Montilla, E.	225
Moore, E. S.	262
Moquette, J. P.	263
Mordaunt, C.	263
Mordvinzeff, A. I.	136
Moreland, C. C.	133, 263
Morgenthaler, O.	368
Mori, N.	263
Moritz, O.	139
Morren, E.	263
Morris, H. E.	249, 263, 407
Morse, E. W.	263
Morse, S. F.	263
Morse, W. J.	263
Morstatt, H.	264
Moss, E. G.	404
Mosséri, V.	264
Mottet, S.	264
Moulton, S. C.	196
Moutia, A.	265
Moyer, A. J.	178
Muir, F. A. G.	265
Mulder, E.	265
Müller,	265
Muller, A. S.	72
Muller, D.	265
Muller, H. C.	265
Muller, H. R. A.	265
Müller, K. R.	265
Müller, W.	266, 324
Mulvania, M.	197, 266
Mumford, E. P.	266
Muncie, J. H.	267
Mungomery, R. W.	267
Muraviov, V. P.	267
Muravjev, V. P.	267
Murphy, P. A.	268, 301
Murray, H. L.	74
Murray, J. C.	271
Murray, P. W.	172, 271
Murton, W. A.	271
Musschenbroek, S. C. van	271
Myers, J. G.	271
Nakata, K.	272
Narasimhan, M. J.	272
Narasimhamurthy, N.	272
Narayana, M.	322
Neal, D. C.	40, 272
Needham, C. W.	272
Neger, F. W.	273
Nelson, R.	273
Neuwirth, F.	274
Newcombe, F. C.	274
Newell, J. M.	32
Newhall, A. G.	274
Newman, C. C.	274
Newton, W.	274
Nillsen, O.	203
Nishimura, M.	274

	Page
Noble, R. J.	274
Nolla, J. A. B.	275
Noordanus, G.	275
Norris, R. V.	275
North, D. S.	276
Northrop, J. H.	281
Norton, J. B. S.	276
Novinenko, A. I.	276
Nowell, W.	199, 277
Oberstein, O.	277
Ocampo, R.	225
Ocfemia, G. O.	257, 277
Oehmichen, C.	279
Ogilvie, L.	279
Olaas, G.	52
Oldaker, G. E. W.	280
Olitsky, P. K.	280
Oliver y Lugo, F.	281
Oortwijn, B. J. G.	281, 298, 301, 302
Orian, G.	283
Orton, C. R.	283
Orton, W. A.	283
Osborn, H. T.	284
Osborn, J. G. B.	392
Osner, G. A.	192
Osterspey,	284
Otero Braguertt, J.	284
Owen, F. W.	79
Page, R. L.	284
Pagliano, T. C. L.	285
Palm, B. T.	285
Pantanelli, En.	285
Pape, H.	285
Paravicini, E.	286
Parisot	285
Park, D. A.	79
Park, M.	286
Parker, E. R.	185, 286
Pascalet, M.	287
Passy, P.	287
Patch, E. M.	287
Peacock, W. M.	287
Pearsall, W. H.	96, 287
Peltier, G. L.	161, 287
Pemberton, C. E.	288
Penhallow, D. P.	288
Perreau	62, 288
Perret, C.	53, 146, 289
Perry, F. R.	56
Pestana, A. C.	289
Peters, L.	290
Peterson, P. D.	290
Pethybridge, G. H.	290
Petre, A. W.	388
Petri, L.	291
Phillips, J. L.	292
Pieper, W.	292
Pierce, W. D.	304
Pierce, W. H.	292
Pinazzoli, F.	293
Pinkhof, M.	293
Pitt, W.	293
Pittman, H. A.	77, 293, 320, 321
Plakidas, A. G.	293
Poeteren, N. van	294
Pompeu de Amaral, A.	294
Poole, R. F.	294
Porter, D. R.	294
Porter, R. H.	294
Presant, E. W.	234
Price, C	79
Price, W. C.	295
Prince, W.	296
Priode, C. N.	152, 296
Pritchett, G. H.	296
Proida, P. A.	297
Prunet, I. A.	297
Purdy, H. A.	44, 297
Puttemans, A.	298
Putterill, V. A.	298
Quanjer, H. M.	135, 282, 298
Quar, Sut Ni	303
Raciborski, M.	303
Racicot, H. N.	304
Ragaswami, S.	356
Ralder, J. M.	168
Raleigh, W. P.	329
Rama Rao, D. A.	304
Rand, F. V.	283, 304
Rands, R. D.	305
Rangel, E.	305
Rankin, W. H.	96, 305
Rao, P. S.	306
Rao, R.	307

	Page
Rao, Y. V. S.	307
Rapson, C. J.	307
Rathschlag, H.	307
Ravn, F. K.	144, 307
Rawlins, T. E.	262, 307, 271
Ray, J.	308
Read, W. H.	308
Reddick, D.	308, 362
Reddy, C. S.	257
Redpath, W. H.	309
Reed, H. S.	309
Reesema Geerts van	309
Reid, K. W.	309
Reiling, H.	309
Riencke, R.	309
Reinking, O. A.	310
Reitmair, O.	214, 310
Remy, L.	310
Remy, T.	310
Rengaswamy, S.	174
Reyes, G. M.	255, 311
Reynolds, E. S.	311
Richards, B. L.	56, 249, 311
Richardson Kuntz, P.	312
Richter, K.	325
Ridler, W. F. F.	312
Rietsema, J.	313
Riha, J.	313
Rikker, R. S.	202
Rischkow, V. L.	313
Rivas, D.	313
Rivers, T. M.	314
Rizhkov, V. L.	313
Robb, W.	314
Robbins, W. W.	314
Robertson, N. F.	314
Robertson, J. N.	314
Roehlin, E. J.	66, 314
Roque, A.	275
Rosa, J. T.	314
Rosen, H. R.	315
Rosenfeld, A. H.	315
Rosenthal, H.	316
Rostrup, S.	144, 164, 229
Rouzinoff, P. G.	316
Roxas, M. L.	316
Rozhdestvenskij, N. A.	316
Ruggles, A. G.	316
Runehjelm, D.	138
Rusconi, A.	316
Rutgers, A. A. L.	317
Salaman, R. N.	317
Salmon, E. S.	234, 318
Samson, R. W.	66, 216, 320
Samuel, G.	38, 320
Sandford, G. B.	105
Sandsten, E. P.	322
Sartory, G.	322
Sastri, B. N.	322, 355
Saunders, A. R.	322
Sauri, F.	322
Savastano, G.	323
Sawyer, Jr., W. H.	359
Sayer, W.	323
Schaffnit, J.	59, 323
Schander, R.	324
Schek, A.	325
Scherffius, W. H.	325
Schertz, F. M.	325
Schewe, O.	326
Schleh	326
Schlesinger, M.	44
Schlumger, O.	326
Schmid, A.	326
Schmidt, A.	326
Schmidt, E. W.	326
Schneider, G.	310
Scholz, W.	326
Schribane,	327
Schribau, E.	327
Schulenberger, O.	30, 31
Schultz, E. W.	327
Schultz, E. S.	148, 149, 166, 327
Schultz, G.	329
Schwartz, M. B.	329
Schwarze, C. A.	52, 330
Schweiner, J.	330
Schweizer, G.	330
Schwing, E. A.	330, 333
Sein, Jr., F.	330
Selby, A. D.	331
Semple, D. M.	331
Serrano, F. B.	255, 332, 373
Soubert, E.	332
Severin, H. C.	334
Severin, H. H. P.	152, 211, 332, 370
Shaposhnikov, J. J.	336

	Page
Shapovalov, M.	336
Shattuck, G. C.	369
Shaw, H. B.	338
Shear, C. L.	201, 338
Shear, W. V.	339
Sheffield, F. M. L.	339
Shepard, E. F. S.	339
Shevchenko, I. S.	340
Shevchenko, L. M.	341
Shevchenko, V. I.	341
Shireff, J.	341
Sieg, F. L.	341
Sieger, E.	341
Siemaszko, W.	341
Silberschmidt, K.	303, 341
Simmonds, J. H.	342
Simonetto, M.	342
Simpson, J.	343
Sirotina, M.	343
Sirrine, F. A.	361
Skeete, C. C.	343
Skinner, J. J.	343
Skorie, V.	344
Skuderna, A. W.	79
Slosson, E. E.	344
Small, W.	344
Smith, A. C.	344
Smith, A. J. M.	344
Smith, A. M.	314
Smith, C. E.	344
Smith, E. F.	344
Smith, E. H.	351
Smith, F. E. V.	346
Smith, F. F.	346
Smith, F. T.	247, 346, 401
Smith, J. H.	234, 339, 346
Smith, K. M.	347
Smith, L. B.	246, 351
Smith, R. E.	351
Smith, R. C.	352
Smyth, E. G.	352
Snell, K.	353
Snyder, W. C.	353
Sokal, N.	51, 353
Sollewijn, G.	354
Solovieva, N. V.	353
Soltwedel, F.	353
Somazawa, K.	241
Sorauer, P. C.	353
Soriano, S.	354
Sornay, P. de	354
Sorokin, H.	354
Southwell, H.	354
Spaeth, C. P.	354
Speare, A. T.	354
Spieckermann, A.	355
Spisar, K.	355
Sprecher, A.	355
Sreenivasaya, M.	272, 304, 322, 355
Srejanni, J.	123
Stahel, G.	356
Stahl, C. F.	78, 357, 391
Stakman, E. C.	224, 358
Stamatinis, N.	123
Stanley, W. M.	358
Standford, E. E.	358
Staner, P.	358
Starrett, R. C.	358
Stell, F.	359
Stepanoff, K. M.	359
Stevens, F. D.	305
Stevens, N. E.	359
Stevenson, F. J.	329
Stevenson, J. A.	199, 359
Stewart, F. C.	361
Stewart, V. B.	308, 362
Stift, A.	154, 362
Stock, J .E. van der	362
Stockes, I. E.	32
Stone, G. E.	363
Stone, R. E.	186, 363
Stoneberg, H. F.	363
Storey, H. H.	363
Stormer, K.	265, 367
Stout, A. B.	368
Stout, G. L.	368, 372
Stover, W. G.	368
Stranviak, F.	368
Strong, R. P.	369
Stuart, W.	369
Stucky, H. P.	369
Sturgis, W. C.	369
Subramanian, L. S.	254
Summer, C. B.	117
Sumner, E. M.	305, 369
Sundaranaman, S.	370
Suzuki, U.	370
Swezey, O. H.	265

	Page
Swezy, O.	211, 335, 370
Swieten, H. J.	370
Szembel, S.	371
Taggart, W. G.	133
Takada, K.	371
Takahashi, W. N.	371
Takami, N.	372
Takimoto, S.	272, 372
Taliaferro, W. H.	230
Tate, H. D.	118
Taubenhaus, J. J.	372
Taylor, G. M.	372
Taylor, W. A.	372
Tchirch, A.	372
Tedin, H.	372
Tehon, L. R.	373
Tempany, H.	373
Teodoro, N. G.	373
Thatcher, R. W.	373
Thiébaut	322
Thomas, E. E.	168
Thomas, R.	384
Thomas, W. W.	333
Thompson, H. S.	373
Thornberg, W. S.	373
Thornton, M. H.	273
Thrupp, T. C.	374
Thung, T. H.	301, 302, 374
Tice, C.	375
Tiebout, G. L.	133
Tiedjens, V. A.	375
Tiesenhausen, M.	324
Tilford, P. E.	375
Tims, E. C.	133, 134, 375
Tisdale, W. B.	375
Tolaas, A. G.	375
Tollenaar, D.	376
Tomei, B.	376
Tompkins, C. M.	322, 376
Tower, W. V.	376
Townley, J.	376
Townsend, C. H. T.	7, 377
Townsend, C. O.	376
Trabut	377
Treub, M.	377
Trochin, J.	377
Trotter, A.	377
Troude, M. J.	377
True, R. H.	196, 377
Tryon, H.	107
Tsen-Cheng	378
Tsienen, S. J.	378
Tsuji, R.	378
Turan, W. E.	48
Turner, W. F.	379
Unite, J. O.	257, 379
Uphof, J. C. T.	379
Uppal, B. N.	379
Utra, G. de	379
Uzel, H.	379
Valeton, T.	379
Valle Zeno, R. del	379
Valleau, W. D.	380
Vanha, J. V.	382
Venkata Rao, M. G.	382
Vanterpool, T. C.	382
Varadaraja Iyengar, A. V.	383
Vasters, J.	383
Vaughan, E. K.	383, 409
Vélez, R.	383
Venkatarama Ayyar, K. R.	383
Venkatraman, T. S.	383
Verhoeven, W. B. L.	384
Vermillion, M. T.	368
Verplancke, G.	358, 384
Verret, D. F. C.	386
Verret, J. A.	386
Verteuil, J.	386
Verwoerd, L.	386
Vestal, E. F.	257
Veve, R. A.	83, 386
Vibert,	387
Vidal, L. F.	387
Villamin, V.	387
Villillo, G.	387
Vicent, C. L.	387
Vinson, C. G.	231, 387, 392
Vochting, H. von	389
Vogel, I. H.	389
Voges, E.	389
Volk, A.	389
Volkart, A.	389
Vukulov, V.	55
Wade, B. L.	408

	Page
Wager, V. A.	389
Wagner, M.	389
Wahl, B.	390
Waite, M. B.	390
Wakefield, E. M.	390
Wakker, J. H.	32, 390
Walbach, S. B.	390
Waldmann, O. P. G.	391
Walker, M. N.	116, 391
Wallace, G. B.	392
Wallace, J. M.	272, 392
Wallman, F.	392
Wann, F. B.	56, 392
Ware, W. M.	234, 319, 392
Warner, M. F.	249
Waters, C. E.	392
Waters, H. A.	13
Watson, H. E.	392
Watt, R. D.	392
Waugh, J. G.	392
Webb, R. W.	196, 250, 251, 392
Weber, G. F.	393
Weber, H.	323
Wedgeworth, H. H.	393
Weimer, J. L.	394
Weiss, F. A.	255, 394
Wellensiek, S. J.	394
Welles, C. G.	225
Wallington, R.	395
Wellman, F. L.	117, 395
Wennink, C. S.	395
Went, F. A. F. C.	390, 395
Werner, H. O.	395
Werth, E.	396
Wessels, P. H.	396
West, E.	393
Westerdijk, J.	396
Weston Jr. W. H.	131, 397
Whetzel, H. H.	397
Whipple, O. B.	397
White, R. P.	397
Whitehead, T.	397
Whitney, W. A.	174
Wichmann, W.	400
Wickens, G. W.	400
Wieler, A.	400
Wilbrink, G.	400
Wilcox, R. B.	401
Wiles, D. R. D.	402
Wille, J.	402
Williams, C. B.	402
Williams, R. D.	234
Wingard, S. A.	152, 211, 402
Winter, J. D.	316, 403
Wolcott, G. N.	7, 403
Wolf, F. A.	73, 403
Wolk, P. R.	404
Wollman, E.	404
Wolzogen-Kühr, C. A. H. von	404
Wood, E. J. F.	102, 404
Wood, J. I.	174, 175
Woods, A. F.	405
Woods, M. W.	405
Woodworth, H. E.	406
Worle,	8
Wortley, E. J.	268, 406
Wright, A.	96
Wright, W. P.	406
Wuthrich, E.	406
Yen Yuen Ting	406
Yoder, P. A.	406
Young, A.	407
Young, P. A.	249, 263, 407
Zaprometoff, N. G.	408
Zaumeyer, W. J.	408
Zedtwitz, W. von	408
Zeller, S. M.	408
Zeylstra, H. H.	409
Ziegler, O.	409
Zimmermann, A.	409
Zundel, G. L.	410

ERRATA

PARTIAL BIBLIOGRAPHY ON VIRUS DISEASES OF PLANTS

Journal of Agriculture of the University of Puerto Rico

Vol. XVIII, Nos. 1–2, p. 1–410, 1934

Page Citation

7—1st—should read "La Enfermedad del Mosaico" o "Matizado" de la caña de azúcar. (Sugar cane "Mosaic" or "Mottling" disease).

8—7th—knollensarbe should read Knollenfarbe.

8—7th—bezielund should read beziehung.

9—1st—investigtion should read investigation.

9—3rd—**Alben, A. O., Cole, J[ohn] R., & Lowis, R. D.** should read **Alben, A[rthur] O[tto], Cole, J[ohn] R., & Lewis, R. D.**

9—4th—Insert Phytopathology before **22**(12):

12—2nd—**Amaral, Afranio de** should read **Amaral, A[velardo] Pompen de.**

12—2nd—Pempen de mosaico should read O mosaico.

12—3rd—**Hatton, R[oland] G[eorge]** should read **Hatton, R[onald] G[eorge].**

12—5th—Pflanzen-Schutz should read Pflanzenschutz.

23—8th—departament should read department.

25—2nd—Blatrollkrankheit should read Blattrollkrankheit.

26—8th—"Red structure" should read "Red suture".

27—4th—Landwirtschaflichen should read Landwirtschaftlichen.

27—6th—bekamft should read bekampft.

28—2nd—Kartoffell should read Kartoffel.

29—9th—insert Mitt. after Forst.

29—10th—Derseitige should read Derzseitige.

30—6th—Forest should read Forst.

31—4th—Kartoffelstanden should raed Kartoffelstauden.

31—4th—foliage should read stalks.

31—6th—Kartoffeleruten should read Kartoffellernten.

36—3rd—**1932–33**: 11 should read 1932–33, **11**: 49–70.

36—6th—A period should be inserted after bibliography.

37—1st—Moestias should read molestias.

40—6th—**M[ortimer]** should read **M[ortier].**

42—4th—Phytopathologické should read Fytopathologické.

Page Citation

42—7th—infectiöse should read infektiöse.
43—1st—infectiose should read infektiose.
43—3rd—infecktiose should read infektiöse.
43—4th—*morginatae* should read *marginatæ*.
45—3rd—wirstch should read wirtsch.
45—3rd—Versuch should read **Versuchsst.**
45—3rd—Justs should read Just's.
45—5th—Veeckziek should read Vleckziek.
45—5th—Tabaklanden should read Tabaksbladen.
45—5th—Verlag should read Verslag.
45—6th—Should read Bemerkung zu dem Aufsatz von Herrn Iwanowski uber die mosaikkrankheiten der Tabakspflanze.
45 7th—insert principle after d'un.
45—7th—feulles should read feuilles.
45—7th—Achiv. should read Archiv.
45—7th—Extractes should read Exactes.
46—1st—insert der after Ursache.
46—1st—Flekenkrankheit should read Fleckenkrankheit.
46—2nd—einer should read zur.
46—9th—Proefhemingen should read Proefnemingen.
47—1st—Mogetijk should read mogelijk.
47—9th—After **125** insert 32 p.,
50—1, 2, 3—Cheshut should read Cheshunt.
51—1st—viekziekte should read vleksiekte.
51—1st—Plantutuin should read Plantentuin.
51—4th—vaie should read vraie.
54—2nd—*sativus* should read *sativum*.
56—9th—Kartoffelabbauses should read Kartoffelabbaus.
57—1st—Blattroll Krankheit should read Blattrollkrankheit.
57—3rd—Unterasuchungen should read Untersuchungen.
58—1, 2—Cheshut should read Cheshunt.
59—5th—Gebtet should read Gebiet.
59—6th—Dent. should read Deut.
59—7th—Before Gebt. insert Forsch. a. d.
60—1st—An die: should read Ans den:
60—3rd—Aüspragung should read Ausprägung.
60—3rd—vegetationsziet should read vegetationzeit.
60—3rd—Blätter Pflanzenb. n. should read Prakt. Blätter Pflanzenb. n.
60—4th—pflanzenzlicher should read pflanzicher.

Page Citation

60—6th—Schädlige should read Schädlinge.

60—6th—Prakt. Blatt. Pflanzend. u. Schuts. should read Prakt. Blätt. Pflanzenb. u. Schutz.

61—1st—insert a comma after *Nicotiana tabacum.*

61—3rd—Multa should read Malta.

64—3rd—ziekt should read siekte.

64—5th—Voolooping should read Voorloopige.

64—5th—Mosaiickziekte should read Mosaickziekte.

64—8th—Pasoervean should read Pasoeroean.

65—2nd—vije should read vrije.

65—6 and 8—Uber should read Ueber.

65—7th—Bekampfungsmethoder should read Bekämpfungsmethoden.

65—7th—Forstschitte should read Fortschritte.

65—8th—verschiedner should read verschiedener.

66—1st—allgmeinen should read allegemeinen.

66—2nd—mikrocheminsche should read mikrochemische.

67—2nd—Philosolophy should read Philosophy.

67—9th—**Britton-Jones, H. R.** should read **Briton-Jones, H. R.**

71—4th—Omm virussy gdomme should read Om virus sygdomme.

73—4th—Zukerrübe should read Zuckerrübe.

75—4th—**Calinissa M. R.** should read **Calinisan, Melanio R.**

78—2nd—Is by **Carsner, Eubanks** not by **Carriers, E. A.**

78—9th—*murals* should *murale.*

80—4th—Is by **Carsner, Eubanks** not by **Carter, Walter.**

81—1st—U. S. Tech. Bull. **206**: should read U. S. D. A. Tech. Bull. **206.**

81—3rd—Ecom. should read Econ.

82—3 and 4—Are by **Carter, Walter** not by **Cayley, Dorothy M.** both are properly cited on pages 80 and 81.

82—8th—Insert and after Mosaic.

85—3rd—Sus should read Sur.

87—2nd—Should be omitted, is a repetition of the succeeding one.

87—3rd—*Epitetrix* should read *Epitrix.*

88—1st—troubies should read troubles.

90—1st—1918 should read 1917.

94—2nd—Undescribe should read Undescribed.

98—5th—degerated should read degenerated.

99—3rd—agronomical should read agronomic.

99—4th—cucious should read curious.

101—5th—Should be omitted it is not a virus disease paper.

Page Citation

103—1st—Kavangeric should read Kavangerie.
105—8th—Bercht should read Bericht.
105—8th—Anftretens should read Auftretens.
105—8th—Komitees should read Kommittees.
105—8th—Land-wirtschaftlich Versuchsweseen in Oesier should read Landwirtschaftlich Versuchswesen in Oeser.
110—2nd—Zeithschrf. should read Zeitschr.
110—6th—**Demarre** should read **Demaree.**
111—9th citation—**Mc Rostie, A[ordon] P[eter]** should read **Mc Rostie, G[ordon] P[eter]**.
111—11th—Omit.
113—1st—shjmisierkte should read slijmzekte.
113—8th—Canberry should read Cranberry.
118—5th—Gesundheitsszuztand should read Gesundheitszutand.
119—2, 3, 4—l'Quest should read l'Oest.
119—6th—Innoculation should read Inoculation.
119—7th—Semmis should read Semis.
123—3rd—planties dans less should read plantes dans les
125—8th—cirus should read virus.
126—2 & 3—ztechr. should read ztschr.
128—8th—**Dvorak M[ayne]** should be read **Dvorak M[ayme]**.
129—1st—Zukerrubenkeimlingen should read Zuckerrubenkeimlingen.
129—1 & 2—Auzeig should read Anzeig.
129—2nd—hervorberufehe should read hervorgeruende.
129—2nd—eine should read eine.
129—2nd—Viruskranheit should read Viruskrankheit.
137—5th—blattrollkankon should read Blattrollkranken.
137—6th—Organ should read Organe.
137—7th—Gas. should read Ges.
139—1st—chlorophydefekte should read Chlorophylldefekte.
146—5th—conton should read canton.
149—1st—**18**: 14–29 should read **17**: 14–29.
149—8th—March should read **11**(3): 65–69.
153—7th—tranmission should read transmission.
154—4th—*apicolis* should read *apicalis*.
154—6th—orschienene should read erschienene.
154—6th—Feiden should read Feinden.
157—8th—Tutunului should read Tutumului.
161—9th—**Peltier, Geo[ge] L[eon]** should read **Peltier, Geo[rge] L[eon]**.

Page Citation

163—4th—**1924** should read **1923**.
164—5th—ealsstedets should read avlsstedets.
165—1, 2, 3—Mosaikeyge should read Mosaiksyge.
165—6th—in those non-solanaceous host should read in non- solanaceous hosts.
166—2nd—sorologique should read serologique.
168—6th—Hovard should read Howard.
170—4th—*sternbergia* should read *Sternbergia.*
173—5th—bladuizen should read bladluizen.
175—4th—untravirus should read ultravirus.
175—5th—1893 should read 1873.
176—8th—Onderzoekingen should read Onderzoekingen.
177—4th—*vitrio* should read *vitro.*
178—3rd—Sortshesistens should read Sortsresistens.
179—4th—Starkeanhanfung should read Starkeanhaufung.
179—5th—Uraschen should read Ursachen.
179—5th—Kartoffelstanden should read Kartoffelstauden.
179—5th—Pflanzenzcht. should read Pflanzenzucht.
179—6th—Blattrolkrankheit should read Blattrollkrankheit.
186—4th—After **182** :insert 22 p.,
187—7th—**17**: 10–16 should read **17**: 1–16.
188—1st—Int's should read Uit's.
188—1st—Teymannia should read Teysmannia.
188—4th—Ceschouwingen should read beschouwingen.
190—4th—**Iachevskü** should read **Iachevskii**.
190—4th—Kratkü should read Kratkii.
190—4th—sostoianüa should read sostoianiia.
190—4th—uchenüa should read ucheniia.
190—4th—mikologü should read mikologii.
190—6th—Studies über eimen should read Studien über einen.
190—6th—trigheit should trahigkeit.
191—2nd—After **Viktorowitch** insert **& Polowzoff, W.**
191—2nd—sielstwo should read sredstwo.
191—2nd—Tabaspflanze should read Tabakspflanze.
191—3rd—zurie should read zwrie.
191—3rd—Inspér should read Impér.
191—7th—Pflanzendrank should read Pflanzenkrank.
192—6th—**Jackwski A[rthur] L[ouis]** should read **Jaczewski, A[rthur] L[ouis].**
196—4th—Ueberkannte Ifektionsstoffe. should read Unbekannte Infektionestoffe.

Page Citation

198—8th—everwinter should read overwintering.
201—6th—After **240** insert p. 54–55,
201—11th—Omit.
203—1st—Jabresher should read Jahresber.
203—1st—Shule should read Schule.
203—4th—Plateave should read Planteavel.
203—5th—Obstr-u should read Obst-u
203—6th—onvodoenden groei should read onvoldoenden groeiën.
203—6th—rilt als gevig should read riet als gevlog.
204—4th—Forsch. should read For.
204—5th—tomosis should read tomotosis.
205—3 & 4—Krigerigheid should read Kringerigheid.
205 3rd—Landbouwhooschool should read Landbouwhoogeschool.
205—4th—pseudenetnecrosis should read pseudo-netnecrosis.
206—2nd—blattrollrankheit should read blattrollkrankheit.
206—2nd—Landro Presesse should read Landrw. Presse
206—2nd—Pflanzenkranklr. should read Pflanzenkrank.
206—8th—Ukologie should read Ökologie.
206—8th—Before Pflanzenzucht insert Pflanzenbau.
206—8th—After Pflanzenzucht insert u. Pflanzenzucht.
207—1st—German should read Germany.
207—1st—Abban should read Abbau.
207—1st—Leistungsiibers-pannungen should read Leistungsüberspannungen.
207—1st—After Pflanzenbau insert a comma.
207—2nd—inen should read innen.
207—3rd—Plant should read Planta.
207—4th—des should read der.
207—4th—Deussche should read Deutsche.
207—4th—Gesellech should read Gesellsch.
207—5th—Demonstrationen should read Demonstrations.
207—6th—Auschluss should read Ansschluss.
207—6th—Viruskrangheiten should read Viruskrankheiten.
207—6th—Plants should read Planta.
207—7th—anschluss should read ansschluss.
207—7th—be (in the annotation) should read he.
207—7th—founded should read found.
208—3rd—Ostenrop should read Osteneurop.
208—5 & 6—Kartoffelban should read Kartoffelbau.
208—6th—Forschunsinst should read Forschunssinst.
209—2nd—Bull. **23** should read Bull. **234**.

Page Citation

209—4th—Is by **Koch, G[ustav], & Kornauth, K.** not by **Kobus, J[acob] D[erk] & Bokma de Boer, B.**

209—4th—Ubertrangung should read Ubertragung.

209—5th—van should read on.

209—5th—Pflanzenschuzstation should read Pflanzenschutzstation.

209—5th—Komitees um should read Kommittees zum.

209—6th—Ztrchr. should read Ztrschr.

209—6th—**16**(3) : 140, 1923 should read **16**(3) : 89–140, 1913.

209—7th—Monattish should read Monatsh.

209—8th—dei should read bei.

210—4th—Sastgut should read Saatgut.

211—1st—Abban should read Abbau

211—1st—Kartoffelban should read Kartoffelbau.

211—3rd—**Keeslag, F. D.** should read **Koeslag, F. D.**

211—3rd—Geselch should read Geselsch.

211—5th—for Kartoffelbans should read fur Kartoffelbau.

211—6th—viruskrakheiten should read viruskrankheiten.

211—8th—Untersuchugen should read Untersuchungen.

212—2nd—au should read an.

212—2nd—Zeuschtdienst should read Pflan.

212—4th—Untersuchgen should read Untersuchungen.

212—7th—Hollandasch should read Hollandish.

213—1st—Fleckenodor should read Flecken odor.

214—2nd—Blatrollkrankheit should read Blattrollkrakheit.

214—3rd—Blattrollkranheit should read Blattrollkrankheit.

214—3rd—sightigung should read sichtigung.

214—3rd—Vervreitung should read Verbreitung.

216—2nd—Zrscht. Pfnazenkrank. should read Ztschr. Pflanzenkrank.

216—3rd—**Kraybill, Henry R[esist]** should read **Kraybill, Henry R[eist].**

216—5th—**Brewer B[earl] H[arvey]** should read **Brewer P[earl] H[arvey].**

216—7th—Physiologie should read Physiologie.

216—7th—Landow should read Landw.

216—7th—Pflanzenzan should read Pflanzenbau.

216—9th—Belenchtung should read Belechtung.

217—2nd—Zucherrohres should read Zuckerrohres.

219—9th—sida should read Sida.

221—1st—vergleichbere should read vergleichbare.

221—1st—Erscheiungen should read Erscheinungen.

221—7th—Denwl. should read Dendrol.

Page Citation

221—11th—Warwaterbehandeling should read Warmwaterbehandeling.

222—2nd—voorfgaande hegroeiing should read voorafgaande begroeiing.

223—2nd—Rech. should read Res.

223—2nd—After *Stellaria media* insert Virulent.

224—2nd—Bllattrollkrankheit should read Blattrollkrankheit.

224—8th—l'humité should read l'humidité.

227—3 & 6—Asliton should read Aston.

228—6th—those bodies should read these bodies.

229—3rd—Stuchen uber die segennante Panashure und uber einge should read Studien über die sogennante Panaschure und über einige.

229—6th—Gatenbance should read Gartenbau.

229—7th—den should read dor.

229—8th—Studies should read Studien.

229—8th—Erschenungen should read Erscheinungen.

230—7th—tabacco should tobacco.

231—1st—mikroskopich should read mikroskopisch.

231—1st—filtriebaren should read filterbaren.

231—3rd—Ledewijks should read Lodewijks.

231—6th—Is by **Loew, [Carl Benedict] Oscar** not by **Lojkin, Mary**.

232—8th—Utersuchungen uber should read Untersuchungen über.

232—8th—Kenntnuis should read kenntnis.

232—8th—Stoffwechsees should read Stoffwechsels.

239—4th—Mitteiking should read Mitteilung.

239—4th—verlanf should read verlauf.

239—4th—Forswirtsch should read Forstwirtsh.

239—7th—**Massée, G.** should read **Massee, G.**

244—1st—Mosaikrakheit should read Mosaikrankheit.

252—1st—Is by **Blood H[erbert] L[oren]** on page 56 not by **Mc Kinney, H. H.**

256—1st—*Capsicatrum* should read *Capsicastrum*.

259—6th—Kückzug should read Rückzug.

260—6th—**Milbraith D[avid] G[allens]** should read **Milbrath D[avid] G[allens]**.

262—3rd—Zukerrübe should read Zuckerrübe.

262—3rd—Zuckerrübenkranheit should read Zuckerrübenkrankheit.

262—4th—ud should read und.

262—4th—Mosaikrankheit should read Mosaikkranheit.

262—4th—Umschan should read Umschau.

Page Citation

263—2nd—Sets should read Iets.
263—3rd—nature should read natura. (In the Italian title)
264—6th—Attersschwache should read Altersschwache.
264—6th—Abban should read Abbau.
264—7th—Angeu should read Angew.
265—5th—Vechselseitige should read Wechselseitige.
265—5th—Famile should read Familie.
266—7th—Kentroll should read Kontroll.
267—4th—Zukerrübe should read Zuckerrübe.
272—4th—**Sreenivasaya, N.** should read **Sreenivasaya, M.**
273—4th—Forstwisch. should read Forstwirtschr.
274—3rd—Rudschau should read Rundschau.
277—5th—Zukerrübe should read Zuckerrübe.
277—5th—America should read Amerika.
284—5th—Kin should read Ein.
284—5th—Ertag should read Ertarg.
285—2nd—Is by **Paine, S. G., & Bewley, W. F.** not by **Pagliano, T.C.L.**
285—8th—Krankreit should read Krankheit.
286—8th—**Parker, E. R. & Horne, Wm. F.** should read **Parker E[rwin] R., & Horne, W[illiam] T[itus].**
291—1st—Recerche should read ricerca.
291.—1st—repporto should read rapporto.
291—2nd—Nuave should read Nuova.
292—2nd—infecttiva should read infettiva.
292—6th—**Hugenford** should read **Hungerford.**
293—7th—*Testranyschus* should read *Tetranyschus.*
294—4th—Omit, is repeated on page 12 where it belongs.
297—7th—precipiting should read precipitin.
298—4th—di should read die.
298—5th—Ztrchr. should read Ztschr.
299—1st—sonanceen should read solanaceen.
302—6th—Euifluss should read Einfluss.
304—1st—staet van Sland Plantentium should read stadt van S'land Plantentiun.
304—2nd—Is by **Rankin W[illiam] H[oward]** not by **Raciborski M.** insert on p. 306.
305—4th—**1926**: 26–24, should read **1926**: 24–26.
306—9th—Transfer under **Jivanna Rao, P. S.** page 195. Omit, is its right place.
307—4th—Mosaik krankheit should read Mosaikkrankheit.

Page Citation

307—4th—Omit the N after Blumen.
309—10th—del should read der.
310—7th—Komitees should read Kommittees.
310—10th—Beobactilungen should read Beobanctungen.
311—1st—Ammer should read Amt.
313—7th—Erzymwirkung should read Enzymwirkung.
314—8th—**Rochlin** is also spell **Rokhlina**.
314—8th—Kartoffel pflanzen should read Kartoffelpflanzen.
317—1st—krulziekye should read krulziekte.
322—8th—Is by **Schander, R.** not by **Sauri, F.**
322—8th—Mitted should read Mittls.
323—5th—intrezellularen should read intrazellularen.
323—6th—Is by **Boning K[arl]** not by **Schaffnit, J.** Omit, is in its right place p. 59.
323—6th—wechselseitize should read wechselseitige.
324—3rd—Untersuchugen should read Untersuchungen.
324—9th—maw should read man.
325—1st—Untersuchugen should read Untersuchungen.
325—4th—insert der before Kartoffel.
325—5th—Omit Degeneration serscheinungen after andere.
325—5th—Fortwirtsch should read Forstwirtsch.
325—6th—Krauselkheit should read Krauselkrankheit.
326—3rd—**Schlumger** should read **Schlumberger.**
326—3rd—Fahre should read Jahre.
325—8th—Mossaikkrankheit should read Mosaikkrankheit.
326—9th—forschungsergebuiss should read forschungsergebris.
326—9th—ibren beziechung zum eizen should read ihren beziehung zum eisen.
326—9th—Vorlanfige voroffenlichung should read Vorläufige voroffenlichung.
327—1st—feseilles should read feuilles.
327—2nd—**Schribau, Emile** should read **Schribaux, Emile.**
330—4th—Atiolegie should read Aetiolegie.
339—6th—**Shepard, E[dward] F[rederick] S[isnett]** should read, **Shepherd, E[dward] F[rederick] S[isnett].**
341—7th—Nikotingenhalt should read Nikotininhalt.
342—1st—in fizierenden should read infizierenden.
351—7th—diseases should read disease (aster yellows).
353—2nd—**34** :should read **24.**
353—8th—Augabliche should read Augenbliche.
353—8th—Kartoffel-epidemic should read Kartoffel-epidemie.

Page Citation

353—9th—Standpunht should read Standpunkt.

355—1st—Jaarhg, should read Jahrg.

355—2nd—Jahrbuche should read Jahrbucher.

355—3rd—mosaikrankheit bet should read mosaikkrankheit bei.

355—5th—mycrocopiques should read microscopiques.

356—2nd—Ocurrence should read Occurrence.

357—1st—dessiebrohrenkrakheit should read der siebrohrenkrankheit.

357—1st—Karfubaumes should read kafeebaumes.

357—5th—**Stahl, C[orwin] E[loyd]** should read **Stahl, C[orwin] F[loyd].**

359—2nd—Astrakham should read Astrakhan.

362—9th—Bakt I should read Bakt II.

365—7th—**6**: 89, should read **6**: 38–39,

366—1st—(*Cicadula*) should read (*Cicadulina*).

368—4th—Abban should read Abbau.

368—4th—wiederanffrischung should read wiederauffrischung.

370—1st—**Sundaranaman** should read **Sundararaman.**

370—6th—(Rickettsit-like) should read (Rickettsia-like).

370—8th—Rickttsia-like should read Rickettsia-like.

370—9th—**Swieten, H. J.** should read **Swieten, H. J. van**

371—1st—*Tetrannychus* should read *Tetranychus.*

371—3rd—**Takada** should read **Takata.**

371—3rd—Eoli should read Coll.

374—1st—**Trupp** should read **Thrupp.**

374—2nd—ondersoch should read onderzoek.

374—3rd—mitgeverd should read uitgevoerd.

374—4th—Inefectives should read Infective.

374—5th—by should read bij.

374—5th—Kulr-an should read Krul-en

374—5th—corzaken should read oorzaken.

374—5th—vors-tenandsche Tobak should read vors-otenlandsche Tabak.

375—1st—Kurl-en should read Krul-en

375—1st—cirkle should read crinkle.

377—9th—Indigége should read Indigéne.

379—3rd—Neus should read neue.

379—7th—pultivieten should read kultivierten.

382—5th—Monatschefte should read Monatshefte.

Page Citation

383—2 & 3—**Varadaraja Iyengar, A. V.** should read **Iyengar, A. V. Varadaraja** cited on page 191 where it belongs.

383—3rd—(n. d.) should read Journ. Ind. Inst. Sci. **16A**(13) : 137–152, 1933.

384—2nd—Plantenkieten should read Plantenziekten.

385—1st—paries should read parties.

385—1st—Bol. should read Bot.

386—3rd—viroses should read viruses.

386—3rd—After :2–10, insert (2) : 22–32,

389—5th—After Luft insert und

389—6th—Abvan should read Abbau.

390—1st—Budensanstalt should read Bundesanstalt.

390—1st—Pflanzenschultz should read Pflanzenschutz.

390—8th—East should read Oost.

391—1st—Unsishtbare should read unsichtbare.

391—1st—Virusfoschung should read Virusforschung.

391—1st—moghichkeit should read moglichkeit.

391—1st—Kunsthche should read Kunstliche.

391—1st—Vermechrung should read Vermehrung.

391—1st—netravisibler should read nitchtvisibler.

393—7th—**Wedkeworth** should read **Wedgworth.**

393—8th—Missouri should read Mississippi.

894—6th—Omit Met een Hollanden samenvatting.

394—7th—invlved should read invled.

396—6th—Klimatologye should read Klimatologie.

396—6th—Ackerbanes should read Ackerbaues.

397—1st—verreiningung should read Vereiningung.

397—10th—**Whitehead, T[athan]** should read **Whitehead T[atham].**

400—1st—Gumüsebau should read Gemüsebau.

400—2nd—Urasäche should read Ursache.

400—3rd—Umschan should read Umschau.

400—5th—grumösen should read gumösen.

400—5th—Verstofungen should Verstopfungen.

400—5th—suckerrohres should read zuckerrohres.

401—2nd—Zuckrindunst should read Zuckerind.

402—3rd—durk should read durch.

402—3rd—erzengte should read erzeugte.

403—10th—**Legman** should read **Lehman.**

404—4th—gezichtspint should read gezichtspunt.

Page Citation

404—4th—Culture should read Cultur.
404—6th—mokroflora should read mikroflora.
406—2nd—insert mosaic after potato.
409—6th—1935 should read 1931.
409—6th—Oekelogie should read Oekologie.
409—7th—Incert **von** after **P[hilipp]**.

o-O-o

SAN JUAN, P. R.
BUREAU OF SUPPLIES, PRINTING, AND TRANSPORTATION
1935